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Schiemann et al.

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(54) GENES AND PROTEINS ASSOCIATED WITH ANGIOGENESIS AND USES THEREOF

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U.S.C. 154(b) by 1199 days.

(21) Appl. No.: 11/542,670

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(65) **Prior Publication Data**

US 2007/0243214 A1 Oct. 18, 2007

Related U.S. Application Data

(60) Provisional application No. 60/772,694, filed on Sep. 30, 2005, provisional application No. 60/816,969, filed on Jun. 27, 2006.

(51) Int. Cl.

 A61K 39/00
 (2006.01)

 A61K 38/16
 (2006.01)

 C12N 5/00
 (2006.01)

(52) **U.S. Cl.** **424/9.1**; 424/227.1; 435/6; 435/32; 435/375; 514/8

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(57) ABSTRACT

Disclosed is a panel of biomarkers associated with angiogenesis, and the use of such biomarkers (genes, proteins, homologues and analogs thereof) to regulate angiogenesis. Methods for identifying compounds useful for regulating angiogenesis and conditions related thereto are disclosed.

2 Claims, 14 Drawing Sheets

FIG. 1A

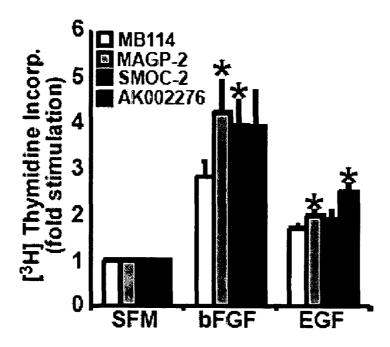


FIG. 1B

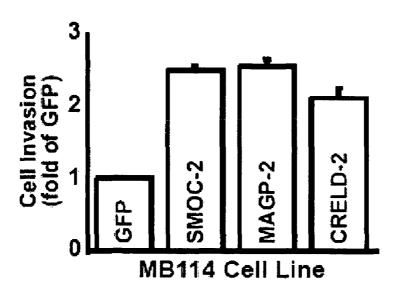


FIG. 1C

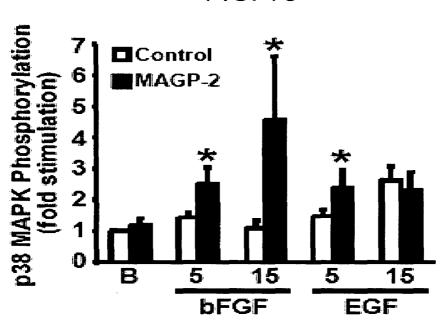


FIG. 1D

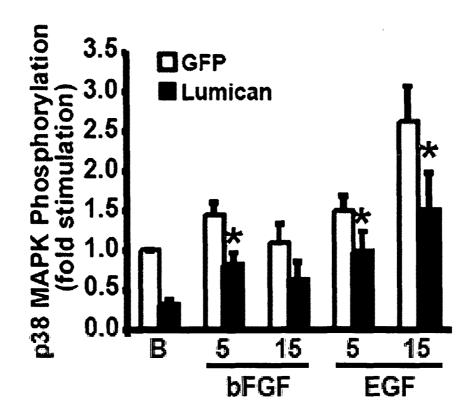


FIG. 1E

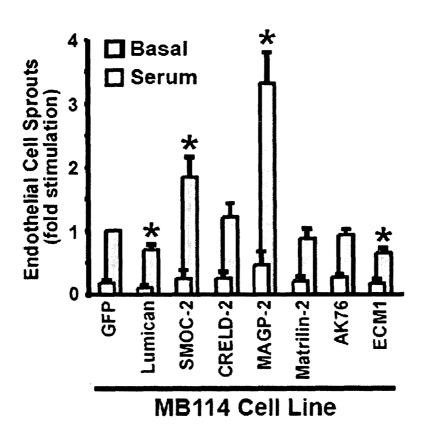


FIG. 2A

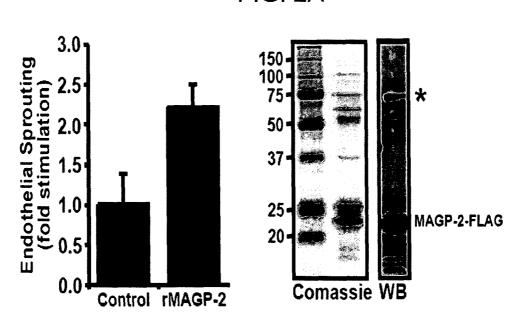


FIG. 2B

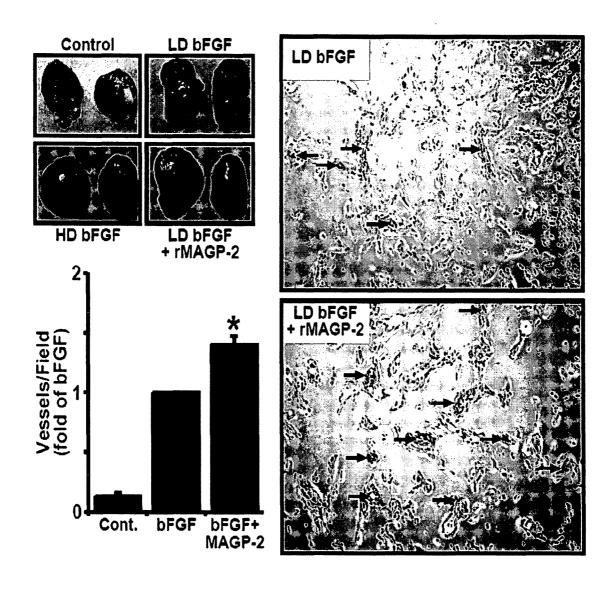


FIG. 3A

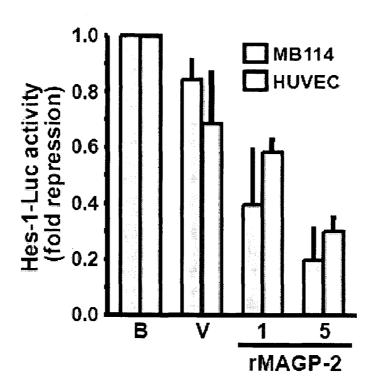


FIG. 4A

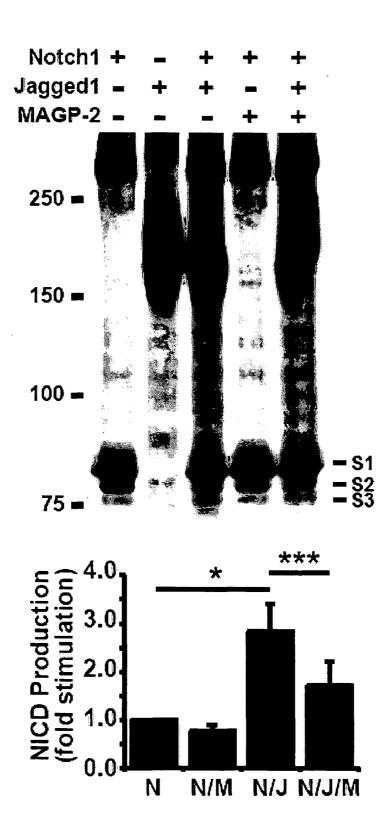


FIG. 4B

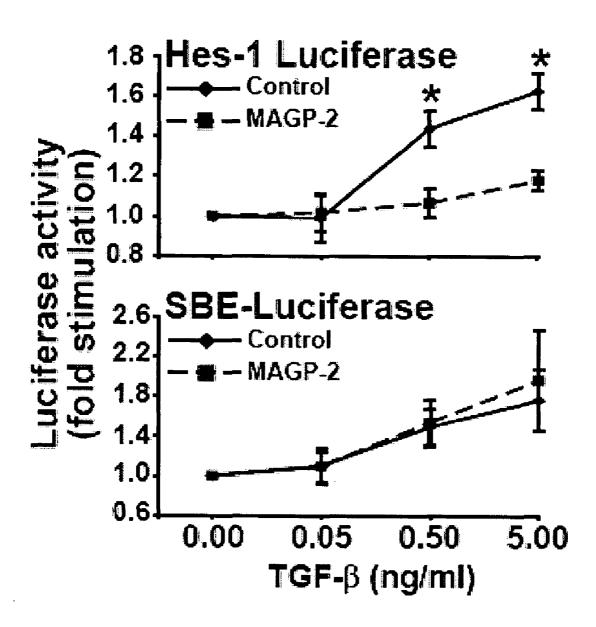


FIG. 5A

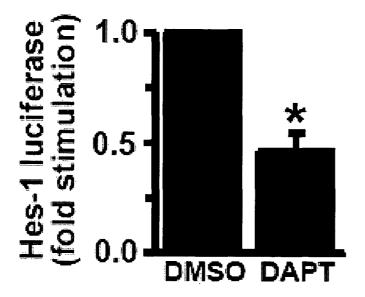


FIG. 5B

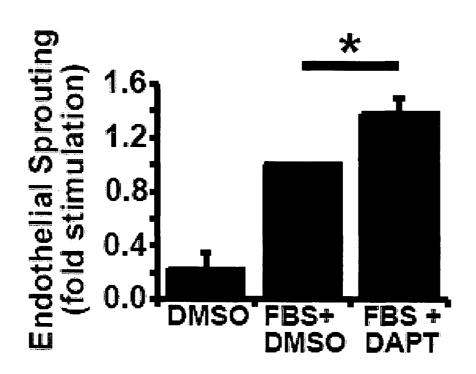


FIG. 5C

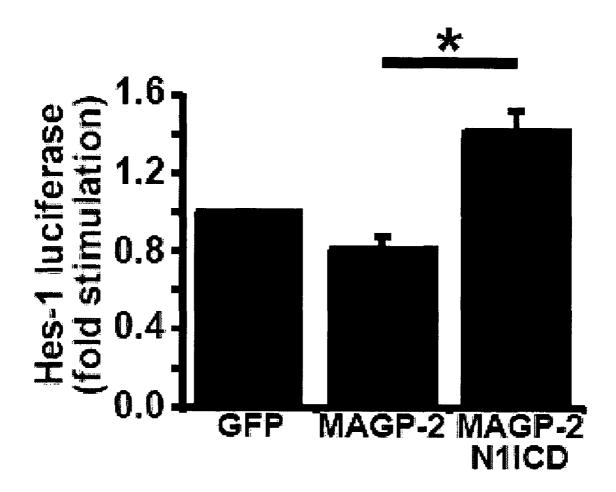
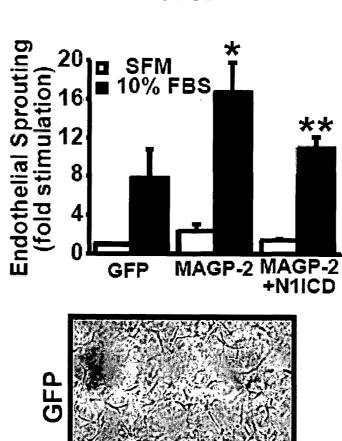


FIG. 5D



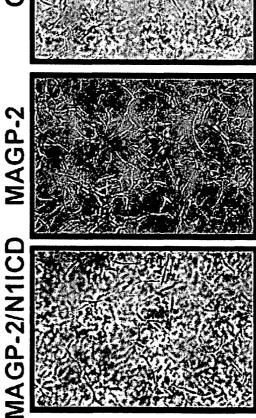


FIG. 6

Tubulogenesis, h

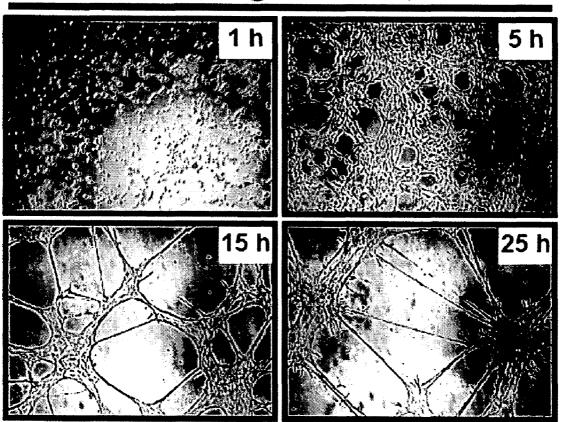


FIG. 7A

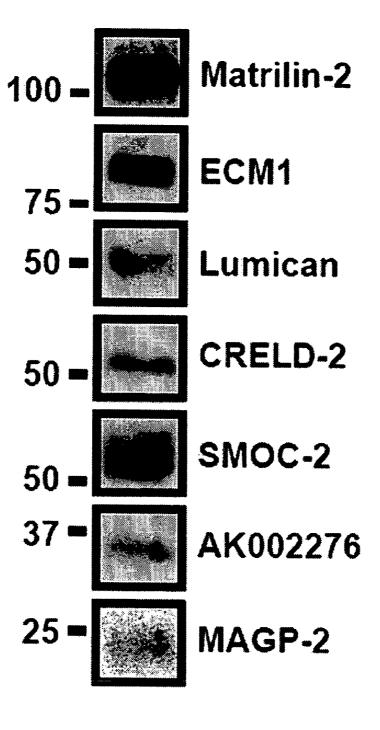


FIG. 7B

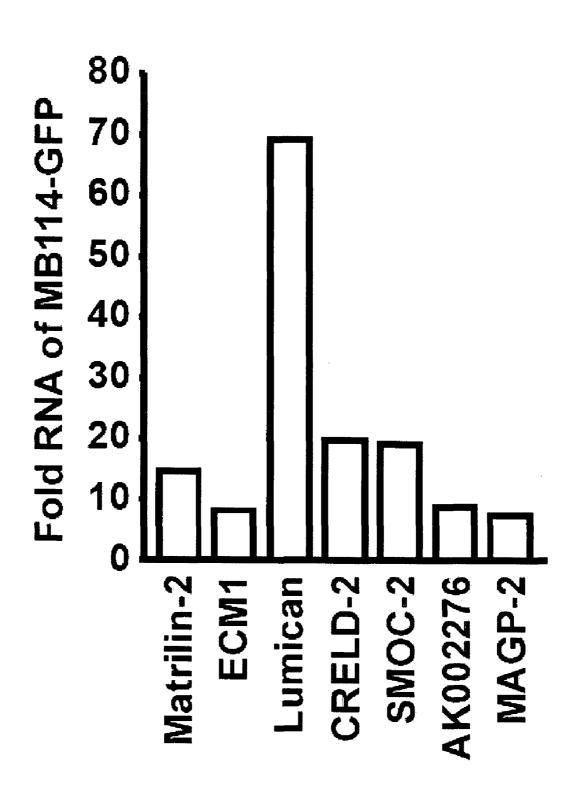
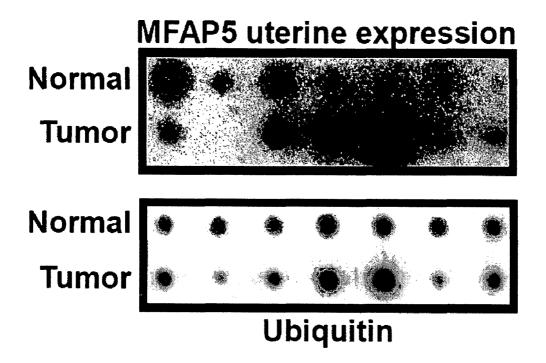


FIG. 8



MFAP5 expression in uterine tumors

Altered 86%(6/7 cases)

Increased 67%(4/6 cases)

GENES AND PROTEINS ASSOCIATED WITH ANGIOGENESIS AND USES THEREOF

CROSS-REFERENCE TO RELATED APPLICATIONS

This application claims the benefit of priority under 35 U.S.C. §119(e) from U.S. Provisional Application No. 60/722,694, filed Sep. 30, 2005 and from U.S. Provisional Application No. 60/816,969, filed Jun. 27, 2006. The entire disclosure of each of U.S. Provisional Application No. 60/722,694 and U.S. Provisional Application No. 60/816,969 is incorporated herein by reference in its entirety.

GOVERNMENT SUPPORT

This invention was made in part with government support under NIH Grant No. CA095519 and NIH Grant No. CA99321, each awarded by the National Institutes of Health. The government has certain rights to this invention.

REFERENCE TO SEQUENCE LISTING

This application contains a Sequence Listing submitted on a compact disc, in duplicate. Each of the two compact discs, which are identical to each other pursuant to 37 CFR §1.52 (e)(4), contains the following file: "Sequence Listing", having a size in bytes of 266 KB, recorded on 2 Oct. 2006. The information contained on the compact disc is hereby incorporated by reference in its entirety pursuant to 37 CFR §1.77 ³⁰ (b)(4).

FIELD OF THE INVENTION

The present invention generally relates to genes and proteins, including homologues and agonist or antagonist analogs thereof, as targets for regulating angiogenesis. The present invention also relates to methods to identify regulators of angiogenesis using such biomarkers, and methods related thereto.

BACKGROUND OF THE INVENTION

Angiogenesis is the process whereby new blood vessels are formed from preexisting vessels; it is a highly regulated event 45 that encompasses a coordinated cascade of gene expression and repression, and one that is influenced by many factors, including a variety of environmental cues provided by the extracellular matrix (ECM) (Sottile, 2004; Stupack and Cheresh, 2002). Cancer cells play a vital role in eliciting 50 many of these environmental cues in part via their ability to produce and secrete numerous angiogenic factors and proteases that create tumor microenvironments conductive to angiogenesis (Bissell et al, 2002; Pupa et al, 2002; Sottile, 2004). Although previously believed to be innocent bystand- 55 ers during angiogenic reactions, it is becoming increasingly apparent that endothelial cells (ECs) also make important contributions to the activation and resolution of angiogenesis. Indeed, ECs generate a variety of environmental cues that shape and remodel tumor and vascular microenvironments, 60 ultimately leading to altered vessel development (Davis and Senger, 2005; Sottile, 2004). Unfortunately, the molecular mechanisms whereby ECs and the molecules they secrete actively direct angiogenesis activation and resolution remain to be determined definitively. It is known that tumor angiogenesis depends upon the coordinated cooperation between cancer and endothelial cells (ECs), and results in the forma2

tion and infiltration of new vessels into tumor microenvironments, thereby providing developing tumors with a source of nutrients and oxygen, as well as a route for cancer cell metastasis (Carmeliet and Jain, 2000; Folkman and Shing, 1992). Failure to establish these cancer: EC connections prevents the development and progression of small, innocuous cancer growths, and as such, tumors remain in a dormant, benign state (Bergers and Benjamin, 2003; Hanahan and Folkman, 1996). Recently, significant inroads in understanding of the role of cancer cells in mediating tumor angiogenesis and EC activation have taken place. Indeed, cancer cells actively induce tumor angiogenesis via their ability to produce and secrete a variety of pro-angiogenic factors (Liotta and Kohn, 2001; Stupack and Cheresh, 2002), a process 15 known as the angiogenic switch (Bergers and Benjamin, 2003; Hanahan and Folkman, 1996). In contrast, comparably little is known concerning the role of ECs during this process, particularly the functional consequences of their ability to remodel vascular and tumor microenvironments during angiogenesis. Although ECs are known to remodel their microenvironment by secreting various extracellular proteases, such as MMPs (matrix metalloproteases), ADAMs (a disintegrin and metalloprotease domain), and ADAMTS (a disintegrin and metalloprotease domain with thrombospondin motifs; Stupack and Cheresh, 2002), a thorough understanding of how these molecules and their stromal targets mediate angiogenesis activation or resolution remains incompletely understood. Thus, identifying and characterizing novel proteins secreted by angiogenic ECs will offer important insights into the role of the endothelium in mediating angiogenesis, as well as its potential to be targeted therapeutically to prevent tumor angiogenesis. Specifically, mapping and defining the EC secretome will significantly enhance understanding of angiogenesis, as well as identify novel therapeutic agents and/or targets that can be exploited to prevent tumor angiogenesis and metastasis in cancer patients.

SUMMARY OF THE INVENTION

One embodiment of the present invention relates to a method to regulate angiogenesis in cells or a tissue of a patient. The method comprises regulating the expression or biological activity in the cells or tissue of any one or more biomarkers selected from a biomarker represented in any one or more of Table I, Table IV, Table V, and/or Table VI.

In one aspect of this embodiment, the biomarkers are any one or more of the biomarkers in Table VI. In another aspect of this embodiment, the biomarkers are any one or more of the biomarkers selected from: ADAMts7, CRELD-2, Decorin, ECM1, Inhibin β -b, Integrin α -3, Integrin α -6, Lipocalin-7, Lox1-3, Lumican, MAGP-2, Matrilin-2, Nephronectin, SerpinE2, and/or SMOC-2.

In another aspect of this embodiment, the biomarkers are any one or more of the biomarkers selected from: 0610007C21Rik, apoptosis related protein APR-3, 1810014L12Rik, Cd14 (encoding CD14 antigen represented herein by SEQ ID NO:5 and SEQ ID NO:6), Cd38 (comprising a nucleic acid sequence represented herein by SEQ ID NO:7 and encoding CD38 antigen); Cd53 (encoding CD53 antigen represented herein by SEQ ID NO:8 and SEQ ID NO:9), Emp2 (encoding epithelial membrane protein represented herein by SEQ ID NO:10 and SEQ ID NO:11), Fcgrt (encoding Fc receptor (IgG, alpha chain transporter) represented herein by SEQ ID NO:12 and SEQ ID NO:13), Islr (encoding immunoglobulin superfamily containing leucine-rich repeat represented herein by SEQ ID NO:14 and SEQ ID NO:15); Lrp2 (comprising a nucleic acid sequence repre-

sented herein by SEQ ID NO:16 and SEQ ID NO:17 and encoding low density lipoprotein receptor-related protein 2); Ly6a (encoding lymphocyte antigen 6 complex, locus A represented herein by SEQ ID NO:18); P2rx4 (encoding purinergic receptor P2X, ligand-gated ion channel 4, represented 5 herein by SEQ ID NO:19 and SEQ ID NO:20; Pcdhb9 (encoding protocadherin beta 9 represented herein by SEQ ID NO:21 and SEQ ID NO:22); Ptpre (encoding protein tyrosine phosphatase receptor type E represented herein by SEQ ID NO:23 and SEQ ID NO:24); Slc4a3 (encoding solute carrier 10 family 4 (anion exchanger) member 3, represented herein by SEQ ID NO:25 and SEQ ID NO:26); and/or Tmc6 (encoding transmembrane channel-like gene family 6, represented herein by SEQ ID NO:27).

In another aspect of this embodiment, the biomarkers are 15 any one or more of the biomarkers selected from: 9130213B05Rik (encoding a protein represented herein by SEQ ID NO:29); C1s (encoding complement component 1, s subcomponent, represented herein by SEQ ID NO:34 and SEQ ID NO:35); C3 (encoding complement component 3 20 represented herein by SEQ ID NO:30 and SEQ ID NO:31); Cfh (comprising a nucleic acid sequence represented herein by SEQ ID NO:32 and SEQ ID NO:33 and encoding complement component factor h); Col9a3 (comprising a nucleic acid sequence represented herein by SEQ ID NO:36 and SEQ ID 25 NO:37 and encoding procollagen, type IX, alpha 3); Grem1 (encoding cysteine knot superfamily 1, BMP antagonist 1, represented herein by SEQ ID NO:38 and SEQ ID NO:39); Lox13 (encoding lysyl oxidase-like 3, represented herein by SEQ ID NO:40 and SEQ ID NO:41); MAGP-2 (comprising a 30 nucleic acid sequence represented herein by SEQ ID NO:123 and SEQ ID NO:124 and encoding microfibrillar associated protein 5, represented herein by SEQ ID NO:42 and SEQ ID NO:43); Mglap (encoding matrix gamma-carboxyglutamate ID NO:45); Naga (encoding N-acetyl galactosaminidase, alpha, represented herein by SEQ ID NO:46 and SEQ ID NO:47); Nbl1 (encoding neuroblastoma, suppression of tumorigenicity 1, represented herein by SEQ ID NO:48 and SEQ ID NO:49); Ngfb (encoding nerve growth factor, beta, repre-40 sented herein by SEQ ID NO:50 and SEQ ID NO:51), Npnt (represented herein by SEQ ID NO:52 and SEQ ID NO:53 and encoding nephronectin); Olfm1 (encoding olfactomedin 1, represented herein by SEQ ID NO:54 and SEQ ID NO:55); and/or U90926 (encoding a protein represented herein by 45 SEO ID NO:56)

Any combinations of any of the above-identified biomarkers are included in the invention. In a preferred aspect of this embodiment, the biomarker is MAGP-2.

In one aspect, the step of regulating comprises contacting 50 the cells or tissue of from the patient with an antagonist of the biomarker. In another aspect, the step of regulating comprises contacting the cells or tissue of from the patient with the biomarker or a biologically active homologue or agonist thereof. In another aspect, the step of regulating comprises 55 expressing a recombinant nucleic acid molecule encoding the biomarker or a homologue thereof in the tissue of the patient.

In one aspect of this embodiment, angiogenesis is upregulated. Such an aspect of the invention can be used to treat a patient that has vascular deficiencies, cardiovascular disease, 60 or would benefit from stimulation of endothelial cell activation and stabilization of newly formed microvessels or other vessels, such as in ischemia or stroke.

In another aspect of this embodiment angiogenesis is downregulated. Such an aspect of the invention can be used to 65 treat conditions that are characterized or caused by abnormal or excessive angiogenesis, including, but are not limited to:

cancer (e.g., activation of oncogenes, loss of tumor suppressors); infectious diseases (e.g., pathogens express angiogenic genes, enhance angiogenic programs); autoimmune disorders (e.g., activation of mast cells and other leukocytes); vascular malformations (e.g., Tie-2 mutation); DiGeorge syndrome (e.g., low VEGF and neuropilin-1 expression); HHT (e.g., mutations of endoglin or LK-1), cavernous hemangioma (e.g., loss of Cx37 and Cx40); atherosclerosis; transplant ateriopathy; obesity (e.g., angiogenesis induced by fatty diet, weight loss by angiogenesis inhibitors); psoriasis; warts; allergic dermatitis; scar keloids; pyogenic granulomas; blistering disease; Kaposi sarcoma in AIDS patients; persistent hyperplastic vitreous syndrome (e.g., loss of Ang-2 or VEGF164); diabetic retinopathy; retinopathy of prematurity; choroidal neovascularization (e.g., TIMP-3 mutation); primary pulmonary hypertension (e.g., germline BMPR-2 mutation, somatic EC mutation); asthma; nasal polyps; inflammatory bowel disease; periodontal disease; ascites; peritoneal adhesions; endometriosis; uterine bleeding; ovarian cysts; ovarian hyperstimulation; arthritis; synovitis; osteomyelitis; and/or osteophyte formation.

Another embodiment of the present invention relates to a method to reduce tumorigenicity in a patient, comprising regulating the expression or biological activity of any one or more biomarkers selected from a biomarker represented in any one or more of Table I, Table IV, Table V, and/or Table VI. In one aspect of this embodiment, the biomarkers are any one or more of the biomarkers in Table VI.

In another aspect of this embodiment, the biomarkers are any one or more of the biomarkers selected from: ADAMts7, CRELD-2, Decorin, ECM1, Inhibin β -b, Integrin α -3, Integrin α-6, Lipocalin-7, Lox1-3, Lumican, MAGP-2, Matrilin-2, Nephronectin, SerpinE2, and/or SMOC-2.

In another aspect of this embodiment, the biomarkers are (gla) protein represented herein by SEQ ID NO:44 and SEQ 35 any one or more of the biomarkers selected from: 0610007C21Rik, apoptosis related protein APR-3, 1810014L12Rik, Cd14 (encoding CD14 antigen represented herein by SEQ ID NO:5 and SEQ ID NO:6), Cd38 (comprising a nucleic acid sequence represented herein by SEQ ID NO:7 and encoding CD38 antigen); Cd53 (encoding CD53 antigen represented herein by SEQ ID NO:8 and SEQ ID NO:9), Emp2 (encoding epithelial membrane protein represented herein by SEQ ID NO:10 and SEQ ID NO:11), Fcgrt (encoding Fc receptor (IgG, alpha chain transporter) represented herein by SEQ ID NO:12 and SEQ ID NO:13), Islr (encoding immunoglobulin superfamily containing leucinerich repeat represented herein by SEQ ID NO:14 and SEQ ID NO:15); Lrp2 (comprising a nucleic acid sequence represented herein by SEQ ID NO:16 and SEQ ID NO:17 and encoding low density lipoprotein receptor-related protein 2); Ly6a (encoding lymphocyte antigen 6 complex, locus A represented herein by SEQ ID NO:18); P2rx4 (encoding purinergic receptor P2X, ligand-gated ion channel 4, represented herein by SEQ ID NO:19 and SEQ ID NO:20; Pcdhb9 (encoding protocadherin beta 9 represented herein by SEQ ID NO:21 and SEQ ID NO:22); Ptpre (encoding protein tyrosine phosphatase receptor type E represented herein by SEQ ID NO:23 and SEQ ID NO:24); Slc4a3 (encoding solute carrier family 4 (anion exchanger) member 3, represented herein by SEQ ID NO:25 and SEQ ID NO:26); and/or Tmc6 (encoding transmembrane channel-like gene family 6, represented herein by SEQ ID NO:27).

> In yet another aspect of this embodiment, the biomarkers are any one or more of the biomarkers selected from: 9130213B05Rik (encoding a protein represented herein by SEQ ID NO:29); C1s (encoding complement component 1, s subcomponent, represented herein by SEQ ID NO:34 and

SEQ ID NO:35); C3 (encoding complement component 3 represented herein by SEQ ID NO:30 and SEQ ID NO:31); Cfh (comprising a nucleic acid sequence represented herein by SEQ ID NO:32 and SEQ ID NO:33 and encoding complement component factor h); Col9a3 (comprising a nucleic acid 5 sequence represented herein by SEQ ID NO:36 and SEQ ID NO:37 and encoding procollagen, type IX, alpha 3); Grem1 (encoding cysteine knot superfamily 1, BMP antagonist 1, represented herein by SEQ ID NO:38 and SEQ ID NO:39); Lox13 (encoding lysyl oxidase-like 3, represented herein by 10 SEQ ID NO:40 and SEQ ID NO:41); MAGP-2 (comprising a nucleic acid sequence represented herein by SEQ ID NO:124 and SEQ ID NO:125 and encoding microfibrillar associated protein 5, represented herein by SEQ ID NO:42 and SEQ ID NO:43); Mglap (encoding matrix gamma-carboxyglutamate 15 (gla) protein represented herein by SEQ ID NO:44 and SEQ ID NO:45); Naga (encoding N-acetyl galactosaminidase, alpha, represented herein by SEQ ID NO:46 and SEQ ID NO:47); Nbl1 (encoding neuroblastoma, suppression of tumorigenicity 1, represented herein by SEO ID NO:48 and SEO 20 ID NO:49); Ngfb (encoding nerve growth factor, beta, represented herein by SEQ ID NO:50 and SEQ ID NO:51), Npnt (represented herein by SEQ ID NO:52 and SEQ ID NO:53 and encoding nephronectin); Olfm1 (encoding olfactomedin 1, represented herein by SEQ ID NO:54 and SEQ ID NO:55); 25 and/or U90926 (encoding a protein represented herein by SEQ ID NO:56).

Any combinations of any of the above-identified biomarkers are included in the invention. In a preferred aspect of this embodiment, the biomarker is MAGP-2.

Another embodiment of the present invention relates to a method to identify a compound that regulates angiogenesis. The method includes the steps of: (a) detecting an initial level of the expression or activity of one or more biomarkers in a cell or soluble product derived therefrom, wherein the biomarker is a biomarker selected from a biomarker represented in any one or more of Table I, Table IV, Table V, and Table VI; (b) contacting the cell with a test compound; (c) detecting a level of the biomarker expression or activity in the cell or soluble product derived therefrom after contact of the cell with the 40 compound; and, (d) selecting a compound that changes the level of biomarker expression or activity in the cell or soluble product therefrom, as compared to in the absence of the compound and/or as compared to the initial level of biomarker expression or activity, as a compound that regulates 45 angiogenesis.

In one aspect of this embodiment, the biomarkers are any one or more of the biomarkers in Table VI.

In another aspect of this embodiment, the biomarkers are any one or more of the biomarkers selected from: ADAMts7, 50 CRELD-2, Decorin, ECM1, Inhibin β -b, Integrin α -3, Integrin α -6, Lipocalin-7, Lox1-3, Lumican, MAGP-2, Matrilin-2, Nephronectin, SerpinE2, and/or SMOC-2.

In another aspect of this embodiment, the biomarkers are any one or more of the biomarkers selected from: 55 0610007C21Rik, apoptosis related protein APR-3, 1810014L12Rik, Cd14 (encoding CD14 antigen represented herein by SEQ ID NO:5 and SEQ ID NO:6), Cd38 (comprising a nucleic acid sequence represented herein by SEQ ID NO:7 and encoding CD38 antigen); Cd53 (encoding CD53 60 antigen represented herein by SEQ ID NO:8 and SEQ ID NO:9), Emp2 (encoding epithelial membrane protein represented herein by SEQ ID NO:10 and SEQ ID NO:11), Fcgrt (encoding Fc receptor (IgG, alpha chain transporter) represented herein by SEQ ID NO:12 and SEQ ID NO:13), Islr 65 (encoding immunoglobulin superfamily containing leucine-rich repeat represented herein by SEQ ID NO:14 and SEQ ID

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NO:15); Lrp2 (comprising a nucleic acid sequence represented herein by SEQ ID NO:16 and SEQ ID NO:17 and encoding low density lipoprotein receptor-related protein 2); Ly6a (encoding lymphocyte antigen 6 complex, locus A represented herein by SEQ ID NO:18); P2rx4 (encoding purinergic receptor P2X, ligand-gated ion channel 4, represented herein by SEQ ID NO:19 and SEQ ID NO:20; Pcdhb9 (encoding protocadherin beta 9 represented herein by SEQ ID NO:21 and SEQ ID NO:22); Ptpre (encoding protein tyrosine phosphatase receptor type E represented herein by SEQ ID NO:23 and SEQ ID NO:24); Slc4a3 (encoding solute carrier family 4 (anion exchanger) member 3, represented herein by SEQ ID NO:25 and SEQ ID NO:26); and/or Tmc6 (encoding transmembrane channel-like gene family 6, represented herein by SEQ ID NO:27).

In yet another aspect of this embodiment, the biomarkers are any one or more of the biomarkers selected from: 9130213B05Rik (encoding a protein represented herein by SEQ ID NO:29); C1s (encoding complement component 1, s subcomponent, represented herein by SEQ ID NO:34 and SEQ ID NO:35); C3 (encoding complement component 3 represented herein by SEQ ID NO:30 and SEQ ID NO:31); Cfh (comprising a nucleic acid sequence represented herein by SEQ ID NO:32 and SEQ ID NO:33 and encoding complement component factor h); Col9a3 (comprising a nucleic acid sequence represented herein by SEQ ID NO:36 and SEQ ID NO:37 and encoding procollagen, type IX, alpha 3); Grem1 (encoding cysteine knot superfamily 1, BMP antagonist 1, represented herein by SEQ ID NO:38 and SEQ ID NO:39); Lox13 (encoding lysyl oxidase-like 3, represented herein by SEQ ID NO:40 and SEQ ID NO:41); MAGP-2 (comprising a nucleic acid sequence represented herein by SEQ ID NO:124 and SEQ ID NO:125 and encoding microfibrillar associated protein 5, represented herein by SEQ ID NO:42 and SEQ ID NO:43); Mglap (encoding matrix gamma-carboxyglutamate (gla) protein represented herein by SEQ ID NO:44 and SEQ ID NO:45); Naga (encoding N-acetyl galactosaminidase, alpha, represented herein by SEO ID NO:46 and SEO ID NO:47); Nbl1 (encoding neuroblastoma, suppression of tumorigenicity 1, represented herein by SEQ ID NO:48 and SEQ ID NO:49); Ngfb (encoding nerve growth factor, beta, represented herein by SEQ ID NO:50 and SEQ ID NO:51), Npnt (represented herein by SEQ ID NO:52 and SEQ ID NO:53 and encoding nephronectin); Olfm1 (encoding olfactomedin 1, represented herein by SEQ ID NO:54 and SEQ ID NO:55); and/or U90926 (encoding a protein represented herein by SEQ ID NO:56).

Any combinations of any of the above-identified biomarkers are included in the invention. In a preferred aspect of this embodiment, the biomarker is MAGP-2.

Another embodiment of the invention relates to a method to identify a compound useful for inhibition of tumor growth or malignancy. The method includes the steps of: (a) detecting an initial level of the expression or activity of one or more biomarkers in a cell or soluble product derived therefrom, wherein the biomarker is a biomarker represented in any one or more of Table I, Table IV, Table V, and Table VI; (b) contacting the tumor cell with a test compound; (c) detecting a level of biomarker expression or activity in the tumor cell or soluble product derived therefrom after contact of the tumor cell with the compound; and, (d) selecting a compound that changes the level of the biomarker expression or activity in the tumor cell or soluble product therefrom, as compared to the initial level of biomarker expression or activity, toward a baseline level of biomarker expression or activity established

from a non-tumor cell, wherein the selected compound is predicted to be useful for inhibition of tumor growth or malignancy

Yet another embodiment of the present invention relates to a method for assessing the presence of tumor cells or potential 5 therefore in a patient. The method includes the steps of: (a) detecting a level of expression or activity of the expression or activity of one or more biomarkers in a test sample from a patient to be diagnosed, wherein the biomarker is a biomarker represented in any one or more of Table I, Table IV, Table V, 10 and Table VI; and (b) comparing the level of expression or activity of the biomarker in the test sample to a baseline level of biomarker expression or activity established from a control sample. Detection of a statistically significant difference in the biomarker expression or activity in the test sample, as 15 compared to the baseline level of biomarker expression or biological activity, is an indicator of the presence of tumor cells or the potential therefore in the test sample as compared to cells in the control sample.

In one aspect of this embodiment, the step of detecting 20 comprises detecting biomarker mRNA transcription by cells in the test sample. For example, such a step of detecting can be performed by a method selected from, but not limited to, polymerase chain reaction (PCR), reverse transcriptase-PCR (RT-PCR), in situ hybridization, Northern blot, sequence 25 analysis, gene microarray analysis, and detection of a reporter gene. In one aspect, the step of detecting comprises detecting biomarker protein in the test sample. For example, such a step of detecting can be performed by a method selected from, but not limited to, immunoblot, enzyme-linked immunosorbant 30 assay (ELISA), radioimmunoassay (RIA), immunoprecipitation, immunohistochemistry and immunofluorescence. In one aspect, the step of detecting comprises detecting biomarker biological activity in the test sample. For example, such a step of detecting can be performed by a method selected 35 from, but not limited to, measuring proliferation of cells expressing the biomarker, measuring angiogenic sprouting of cells expressing the biomarker, and measuring migration and invasion ability of endothelial cells expressing the biomarker.

In one aspect of this embodiment, the test sample is from a 40 source selected from the group consisting of: breast, kidney, ovary, colon, and uterus, in the patient. In another aspect, the test sample is from a patient being diagnosed for cancer and wherein the baseline level is established from a negative control sample that is established as non-tumorigenic.

In one aspect of this embodiment, the baseline level is established by a method selected from the group consisting of: (1) establishing a baseline level of biomarker expression or activity in an autologous control sample from the patient, wherein the autologous sample is from a same cell type, tissue 50 type or bodily fluid type as the test sample of step (a); (2) establishing a baseline level of biomarker expression or activity from at least one previous detection of biomarker expression or activity in a previous test sample from the patient, wherein the previous test sample was of a same cell type, 55 tissue type or bodily fluid type as the test sample of step (a); and, (3) establishing a baseline level of biomarker expression or activity from an average of control samples of a same cell type, tissue type or bodily fluid type as the test sample of step (a), the control samples having been obtained from a popula- 60 tion of matched individuals.

Yet another embodiment of the invention relates to an assay kit for assessing angiogenesis or the presence of tumor cells in a patient, comprising: (a) a reagent for detecting the expression or activity of a biomarker in a test sample, wherein the 65 biomarker is a biomarker represented in any one or more of Table I, Table IV, Table V, and Table VI; and (b) a reagent for

8

detecting a control marker characteristic of a cell or tissue type that is in the test sample or that is secreted into the test sample by the cell or tissue. In one aspect, the reagent of (a) is selected from the group consisting of: a hybridization probe of at least about 8 nucleotides that hybridizes under stringent hybridization conditions to a nucleic acid molecule encoding the biomarker or a fragment thereof; an oligonucleotide primer for amplification of mRNA encoding the biomarker or a fragment thereof; and an antibody that selectively binds to the biomarker. In one aspect, the reagent of (b) is selected from the group consisting of: a hybridization probe of at least about 8 nucleotides that hybridizes under stringent hybridization conditions to a nucleic acid molecule encoding the control marker or a fragment thereof; an oligonucleotide primer for amplification of mRNA encoding the control marker or a fragment thereof; and an antibody that selectively binds to the control marker. In one aspect, the reagents of (a) and (b) are suitable for use in a method of detection selected from the group consisting of immunohistochemistry and immunofluorescence.

Yet another embodiment of the invention relates to a method to reduce angiogenesis in cells or a tissue of a patient, comprising decreasing the expression or biological activity of Microfibril-associated glycoprotein-2 (MAGP-2) in the cells or tissue.

Another embodiment of the invention relates to a method to promote angiogenesis in cells or a tissue of a patient, comprising increasing the expression or biological activity of MAGP-2 in the cells or tissue.

Another embodiment of the invention relates to the use of MAGP-2 or a fragment or homologue thereof, or a nucleic acid molecule encoding MAGP-2 or a fragment or homologue thereof, or an agonist or antagonist of MAGP-2, in the preparation of a medicament for the regulation of angiogenesis

BRIEF DESCRIPTION OF THE FIGURES OF THE INVENTION

FIG. 1A is a bar graph shows DNA synthesis (determined by measuring [³H]thymidine incorporation into cellular DNA) in serum-starved MB114 cells stably expressing either GFP or various putative angiogenic agents, stimulated in the absence or presence of either bFGF (50 ng/ml) or EGF (10 ng/ml) for 24 h at 37° C. (data are the mean (±SEM) of five independent experiments for MAGP-2 and SMOC-2, and of three independent experiments of CRELD-2; *, p<0.05; Student's T-Test).

FIG. 1B is a bar graph showing the invasion of MB114 cells expressing either GFP or various putative angiogenic agents through synthetic basement membranes over 48 h using a modified Boyden-chamber assay (data are the mean (±SEM) of three independent experiments; *, p<0.05; Student's T-Test).

FIGS. 1C and 1D are bar graphs showing p38 MAPK phosphorylation in serum-starved MB114 cells expressing MAGP-2 (FIG. 1C) or lumican (FIG. 1D), stimulated with either bFGF (50 ng/ml) or EGF (10 ng/ml) 0-15 min (data are the mean (±SEM) of 5 independent experiments; *, p<0.05; Student's T-Test).

FIG. 1E is a bar graph showing endothelial cell sprouting in MB114 cells expressing either GFP or various putative angiogenic agents (data are the mean (×SEM) of 5 independent experiments for lumican, SMOC-2, CRELD-2, MAGP-2, and Matrilin-2, and of three independent experiments for AK76 and ECM-1; *, p<0.05; Student's T-Test).

FIG. 2A shows that MAGP-2 (MAGP-2 purity was monitored by coomassie staining, and by immunoblotting with anti-FLAG M2 monoclonal antibodies (right panel)) promotes angiogenesis in vivo, as measured by angiogenic sprouting of quiescent MB114 cell monolayers (left panel) 5 (data are the mean (±SEM) of two independent experiments; *, p<0.05; Student's T-Test).

FIG. 2B shows the results of subcutaneous injection of C57BL/6 female mice with Matrigel supplemented either with diluent (D), bFGF (50 ng/ml, LD; or 300 ng/ml, HD), or bFGF (50 ng/ml) in combination with MAGP-2 (1 μg/ml), where plugs were harvested and photographed (left panels), and then fixed, sectioned, and stained with Masson's trichrome to visualize infiltrating blood vessels (right panels; arrows denote blood vessels) (data are the mean (±SEM) of four independent experiments; *, **, ****, p<0.05; Student's Test)

FIG. 3A is a bar graph showing that MAGP-2 inhibits Hes-1 promoter activity in ECs (data are mean (±SEM) of 2 20 independent experiments).

FIG. 3B is a bar graph also showing that MAGP-2 inhibits Hes-1 promoter activity in ECs (data are the mean (±SEM) of four independent experiments; *, ***, ****, p<0.05; Student's Test).

FIG. 4A shows Notch1 cleavage products (upper) and the densitometric analysis of Notch1 NICD production in response to experimental treatments (lower) in human 293T cells transiently transfected with cDNAs encoding Myctagged versions of Notch1, Jagged-1, and MAGP-2 in all combinations as indicated (data are the mean (±SEM) of four independent experiments; *, ***, p<0.05; Student's T-Test; N, Notch1; N/M, Notch1 plus MAGP-2; N/J, Notch1 plus Jagged-1; N/J/M, Notch1, Jagged-1, and MAGP-2).

FIG. 4B shows luciferase activity after stimulation with TGF-β1 in GFP- and MAGP-2-expressing MB114 cells transiently transfected with either pHes1- or pSBE-luciferase, both together with pCMV-β-gal as indicated (data are the mean (±SEM) of 3 independent experiments; *, p<0.05; Stu-dent's T-Test).

FIG. **5**A is a bar graph showing Hes-1 luciferase activity in MB114 cells transiently transfected with pHes1-luciferase and pCMV- β -gal cDNAs, incubated overnight in the absence or presence of DAPT (10 μ M) (data are the mean (\pm SEM) of 45 two independent experiments).

FIG. **5**B is a bar graph showing endothelial angiogenic sprouting in quiescent MB114 cell monolayers induced to form angiogenic sprouts by addition of 10% FBS supplemented with or without DAPT (10 μ M) (data are the mean (\pm SEM) of four independent experiment. (*, p<0.05; Student's T-Test)).

FIG. 5C is a bar graph showing Hes-1 luciferase activity in GFP-, MAGP-2-, and MAGP-2/N1ICD-expressing MB114 cells transiently transfected with pHes1-luciferase and pCMV- β -gal cDNAs (data are the mean (\pm SEM) of two independent experiments).

FIG. **5**D is a bar graph showing endothelial angiogenic sprouting in quiescent monolayers of GFP-, MAGP-2-, and MAGP-2/N1ICD-expressing MB114 cells (bottom shows representative photomicrographs of angiogenic sprouts produced by GFP-, MAGP-2-, and MAGP-2N1ICD-expressing MB114 cells; data are the mean (±SEM) of four independent experiments; *, ***, p<0.05; Student's T-Test).

FIG. 6 is a digitized image showing the time course of angiogenesis in vitro.

10

FIGS. 7A and 7B show retroviral expression of selected potential angiogenic proteins in MB114 cells via detergent-solubilized cell extracts (FIG. 7A) and semi-quantitative real-time PCR (FIG. 7B).

FIG. **8** is a digitized image showing that MAGP-2 is expressed aberrantly in a majority of human uterine tumors.

DETAILED DESCRIPTION OF THE INVENTION

The present invention generally relates to the discovery by the present inventor of several genes, and the proteins encoded thereby, that are associated with angiogenesis. More particularly, the present inventors used microarray analyses to monitor changes in the transcriptome of ECs undergoing angiogenesis when cultured onto tumor-derived basement membranes in vitro. In doing so, the inventors identified 308 genes whose expression was altered at least 3-fold during the angiogenic time course. Of these differentially-expressed genes, 63 encoded for EC secretory proteins and several were shown to mediate pro- or anti-angiogenic activities in vitro (e.g., SMOC-2, secreted MAGP-2 Promotes Angiogenesis modular calcium-binding protein-2; CRELD-2, cysteine-rich with EGF-like domains-1; MAGP-2, microfibril-associated glycoprotein-2; lumican; ECM-1, extracellular matrix protein-1). Expression of one of these genes, MAGP-2 (also known as Microfibrillar associated protein-5 (MFAP-5)), enhanced EC proliferation and p38 MAPK activation stimulated by bFGF, as well as stimulated EC invasion through synthetic basement membranes. The inventors have also demonstrated that MAGP-2 promoted EC sprouting in vitro, and as such, stimulated vessel formation and infiltration into Matrigel plugs implanted into genetically normal mice. Importantly, the inventors show herein that Notch1 activation prevented angiogenesis in vitro, a reaction that was overcome by MAGP-2-mediated antagonism of Notch1 signaling in ECs. Collectively, the inventors' findings have established MAGP-2 as a novel inducer of angiogenesis, doing so in part through its ability to antagonize Notch1 signaling in ECs. In addition, the inventors' findings have identified several additional targets for use in diagnostic, drug discovery and therapeutic applications related to the inhibition or promotion of angiogenesis.

More particularly, in order to increase the understanding of the role of ECs in mediating the remodeling of tumor and vascular microenvironments during pathological angiogenesis, the inventors cultured ECs on tumor-derived basement membranes to induce angiogenesis in vitro, and subsequently performed microarray analyses to identify alterations within the EC transcriptome that accompanied angiogenesis activation. In doing so, they focused specifically on genes that encoded secretory proteins or components of the ECM, which collectively comprised 20% (i.e., 63 out of 308 genes) of the differentially-expressed EC genes identified by the inventors (Table I). The analyses described herein also identified an additional 35 (~11%) membrane-spanning and/or membrane-associated genes, whose expression and activation likely mediate paracrine and/or autocrine signaling in angiogenic ECs. Thus, secreted molecules constituted a significant fraction (~31%) of all differentially regulated EC genes identified herein, thereby highlighting the importance of microenvironment remodeling during angiogenesis. The proportion of differentially-expressed EC genes classified as secretory proteins was similar to those observed in other recent EC transcriptome analyses (Aitkenhead et al, 2002; Bell et al, 2001; Kahn et al, 2000). However, unlike these profiling studies, the present inventors specifically investigated the inductive effect of tumor-derived basement membranes (i.e.,

Matrigel matrices) in regulating gene expression in tubulating ECs, and as such, numerous secretory proteins not previously associated with angiogenesis were identified (see Table I). Moreover, the inventors' identification of known angiogenic genes (Table I) validated this experimental design and gave 5 credence to the notion that many of these newly identified genes may function as bone fide regulators of angiogenesis. Indeed, the present inventors' findings implicate ECM-1 and lumican as mediators of angiostasis, while CRELD-2 and SMOC-2 are proposed herein to function as novel mediators of angiogenesis (see discussion below). The ability of these EC secretory proteins to affect vessel development in vivo, as well as the molecular mechanisms whereby they mediate their pro- or anti-angiogenic activities in ECs can now be evaluated using the guidance provided herein.

An especially important finding of the present study was the inventors' identification of MAGP-2 as a novel mediator of angiogenesis. Indeed, the present inventors show for the first time that MAGP-2 expression stimulates EC proliferation, invasion, and angiogenic sprouting, as well as enhances 20 EC activation of p38 MAPK in response to bFGF and EGF (FIG. 1). Moreover, MAGP-2 is shown to enhance the ability of bFGF to promote neovascularization and vessel infiltration into Matrigel plugs implanted into genetically normal mice (FIG. 2). Mechanistically, MAGP-2 is shown to induce angio- 25 genesis through its ability to inhibit Notch1 processing and activation (FIGS. 3 and 4), an inhibitory reaction that is rescued by constitutive expression of Notch1 NICD (FIG. 5). Collectively, these findings have established MAGP-2 as a novel activator of angiogenesis, doing so in part via its ability 30 to inhibit the Notch1 signaling pathway.

The precise mechanism whereby MAGP-2 antagonizes Notch1 signaling remains to be determined. Recent studies using heterologous cell expression systems have shown MAGP-2 to interact physically with Notch1 and its ligand, 35 Jagged-1, resulting in their shedding from the cell surface (Miyamoto et al, 2006; Nehring et al, 2005). Although the inventors made no attempt to measure Notch1 and/or Jagged-1 extracellular domain shedding in response to MAGP-2, the production of such soluble Notch1 and 40 Jagged-1 extracellular domains readily inhibits Notch signaling (Rebay et al, 1993; Small et al, 2001). In this fashion, MAGP-2 expression was observed to block the ability of Jagged-1 to stimulate Notch1 processing and the production of NICD, thereby preventing transactivation of the Hes1 pro- 45 moter in ECs. Thus, MAGP-2 may promote angiogenesis in part by inducing Notch1 and/or Jagged-1 ectodomain shedding in ECs. In contrast to the present inventors' findings, Miyamoto et al (Miyamoto et al, 2006) recently found that MAGP-2 not only induces Notch1 ectodomain shedding in 50 Cos-7 and NIH-3T3 cells, but also Notch1 processing and NICD production, leading to transcriptional activation of the Hes5 and CSL promoters. The reasons underlying this discrepancy are currently unknown, but most likely reflect differences in the cell types studied (i.e., ECs versus fibroblasts 55 and kidney epithelial cells), as well as differences in microenvironmental factors that may influence the interactions between MAGP-2 and Notch1. In addition, cell-type specific expression of various Notch receptor and ligand combinations may also impact the ability of MAGP-2 to regulate, 60 either positively or negatively, Notch signaling in responsive cells. Indeed, the present inventors, without being bound by theory, believe that MAGP-2 regulates angiogenesis in a context-specific manner via its ability to target both Notch signaling and elastin microfibril networks.

The present inventors' findings demonstrating the ability of MAGP-2 to stimulate angiogenesis by preventing Notch1

12

activation is intellectually credible in light of the established function of Notch in mediating angiostasis (Leong et al, 2002; Liu et al, 2006; Noseda et al, 2004; Williams et al, 2006; Zimrin et al, 1996). Moreover, the inventors recently observed MAGP-2 expression to be abnormally elevated in human uterine cancers (Example 6), and to significantly increase the growth and vascularization of MCA102 fibrosarcomas produced in mice (Albig and Schiemann, unpublished observation). It should be noted, however, that Notch activation also has been shown to stimulate angiogenesis (Leong and Karsan, 2005; Shawber and Kitajewski, 2004), and as such, it cannot yet be ascertained whether MAGP-2 promotes tumorigenesis by alleviating Notch1-mediated angiostasis, or by facilitating Notch1-mediated angiogenesis. The mechanisms whereby Notch mediates such disparate activities in ECs remains unclear, but may reflect a complex integration of cellular and environmental cues. Indeed, Notch signaling is subject to regulation by (i) the relative expression levels of various Notch receptors (Delaney et al, 2005; Duarte et al. 2004); (ii) the extent and form of Notch receptor glycosylation (Haines and Irvine, 2003); (iii) the availability of various Notch ligands within vascular microenvironments; and (iv) the activation of various Notch inhibitors, including MINT, Numb, NRARP, and proteolyzed ligands (Kadesch, 2004). The present inventors' findings herein and those by others (Miyamoto et al, 2006; Sakamoto et al, 2002) clearly show Notch signaling to be influenced by environmental cues, such as those produced by MAGP-2 (demonstrated herein).

Numerous additional EC secretory proteins were identified whose expression was also regulated by angiogenesis (Tables I and VI), suggesting that EC expression of these genes was obligatory for vessel development. Moreover, in vitro assays that modeled key steps in the angiogenic process showed that several these newly identified genes did indeed regulate EC activities-coupled to angiogenesis. For instance, lumican expression was found to inhibit MB114 cell proliferation (data not shown) and angiogenic sprouting (FIG. 1), as well as reduce the ability of bFGF and EGF to activate p38 MAPK in MB114 cells (FIG. 1). Lumican belongs the SLRP (small leucine-rich proteoglycan) family of ECM proteins, which also includes fibromodulin, biglycan, and the angiogenesis antagonist, decorin (Davies Cde et al, 2001; Kao et al, 2006; Sulochana et al, 2005). Genetic ablation of lumican in. mice indicates that this secreted proteoglycan functions in organizing collagen fibrils in the skin and cornea (Chakravarti et al, 1998). Additionally, lumican interacts physically with FasL (Fas-ligand), leading to enhanced Fas expression in and subsequent apoptosis of corneal fibroblasts (Vij et al, 2004; Vij et al, 2005). Recently, elevated lumican expression has been associated with cancers of the pancreas (Ping Lu et al, 2002), breast (Leygue et al, 1998), cervix (Naito et al, 2002), and colon (Lu et al, 2002), suggesting that lumican may promote tumorigenesis in these organs. In stark contrast, lumican expression also has been shown to inhibit the anchorageindependent growth and invasion of B16F1 melanoma cells in vitro, as well as their ability to form tumors in when implanted into mice (Vuillermoz et al, 2004). Thus, lumican also may function in suppressing cancer development and progression. Along these lines, the inventors have found that lumican antagonizes the development and infiltration of vessels in Matrigel plugs implanted into mice, as well as decreases the growth and blood vessel density of MCA102 fibrosarcomas produced in mice (Albig and Schiemann, unpublished observations).

The inventors further showed that ECM-1 is functionally similar to lumican and antagonized angiogenic sprouting by

MB114 cells (FIG. 1). ECM-1 is a broadly distributed glycoprotein that plays important roles in maintaining normal skin structure, function, and homeostasis (Chan, 2004). In humans, loss of function mutations in ECM-1 elicit a rare genetic skin disease called lipoid proteinosis (Chan, 2004; 5 Hamada et al, 2002), whose clinicopathological features are phenocopied in patients with lichen sclerosus, an acquired inflammatory disorder of the skin and mucous membranes associated with the development self-reactive ECM-1 antibodies (Oyama et al, 2003). Interestingly, both skin conditions are characterized by the (i) abnormal development of cutaneous microvessels, and (ii) excessive deposition of basement membrane proteins, leading to thickened mucous and vascular basement membranes (Kowalewski et al, 2005). ECM-1 overexpression is observed in cancers of the breast, 15 esophagus, thyroid, stomach, and colon (Han et al, 2001; Kebebew et al, 2005; Wang et al, 2003), and has been associated with the acquisition of angiogenic (Han et al, 2001) and metastatic phenotypes (Wang et al, 2003). Thus, ECM-1 is an important regulator of basement membrane protein secretion 20 and deposition, and quite possibly, of microenvironment remodeling (Kowalewski et al, 2005; Mirancea et al, 2006). As such, aberrant ECM-1 production likely dysregulates normal microenvironment conditions operant in balancing proand anti-angiogenic signals, leading to altered vessel forma- 25 tion and disease development in humans.

In contrast to lumican and ECM-1, the inventors observed CRELD-2 expression to significantly increase MB114 cell invasion, and to promote a trend towards enhanced angiogenic sprouting (FIG. 1), indicating that this secreted EGF- 30 like domain containing protein may serve to enhance angiogenesis. Along these lines, the inventors found SMOC-2 expression to enhance the proliferative response of MB114 cells to bFGF, and more importantly, to increase MB114 cell invasion and angiogenic cell sprouting (FIG. 1). SMOC-2 and 35 its related molecule, SMOC-1, are widely expressed glycoproteins that localize predominantly to basement membranes, and to various ECM structures (Vannahme et al, 2003; Vannahme et al, 2002). Structurally, SMOCs are defined by a unique, centrally located SMOC domain that is flanked N-ter- 40 minally by follistatin-like and thyroglobulin-like domains, and C-terminally by an extracellular calcium-binding (EC) domain reminiscent of that found in SPARC (Vannahme et al, 2003; Vannahme et al, 2002). Interestingly, proteolytic cleavage of SPARC results in the release of biologically active 45 fragments that can induce angiogenesis (Funk and Sage, 1993; Sage et al, 2003). SPARC, however, also mediates angiostasis by interacting physically with VEGF via its EC domain (Jendraschak and Sage, 1996; Kupprion et al, 1998). Thus, given the functional and structural similarities between 50 SMOC-2 and SPARC, it remains to be determined whether SMOC-2 also mediates pro- and anti-angiogenic activities, and if so, whether these disparate EC activities occur via direct or indirect mechanisms.

Collectively, the inventors' findings indicate that lumican and EMC-1 function as novel angiogenesis antagonists, while CRELD-2 and SMOC-2 function as novel angiogenesis agonists. The molecular mechanisms underlying their ability to impact the activation or resolution of angiogenesis can now be determined.

The present invention more particularly relates to genes, nucleic acid molecules derived therefrom, and proteins or fragments thereof encoded by such genes and nucleic acid molecules, as well as homologues of such genes and proteins and related agents (e.g., antibodies, agonists, antagonists), and the use or targeting of such genes, nucleic acids, proteins, homologues and/or related agents, and/or compositions or

14

formulations comprising the same, in methods related to the inhibition or promotion of angiogenesis, including the inhibition of angiogenesis for the inhibition or treatment of cancer. As discussed above, the present inventors identified 308 genes whose expression in angiogenic ECs was altered ≥3-fold. Of these differentially-expressed genes, 63 genes (~20%) encoded EC secretory proteins (Table I), 35 genes (~11%) encoded transmembrane or membrane-associated proteins (Table V), and 210 genes encoded non-secretory proteins (Table IV). This approach identified several secretory proteins that were previously known to be associated with angiogenesis and/or microenvironment remodeling, including ADAMTS1 (Iruela-Arispe et al, 2003), CTGF (Brigstock, 2002), HGF (Gao and Vande Woude, 2005), MMPs 3 and 9 (Heissig et al, 2003), thrombospondins 1 and 2 (Armstrong and Bornstein, 2003), and TIMP3 (Qi et al, 2003) (Table I, bold type face). In addition, the inventors identified numerous secretory proteins not previously associated with angiogenesis (e.g., Table I, regular text face), all of which are encompassed by the present invention. The inventors verified the differential expression of 19 individual genes by semi-quantitative real-time PCR (see Materials and Methods). These analyses showed significant concordance in the expression profiles measured either by real-time PCR or microarray analyses (Table VI), indicating that these (and other) genes are indeed bona fide targets of angiogenic signaling systems in tubulating ECs.

Accordingly genes that are encompassed by the present invention (as well as nucleic acid molecules derived from or comprising at least a portion of the coding region and/or regulatory region of such genes and any proteins or fragments thereof encoded by such genes) include any of the genes or portions of genes (including ESTs) represented in Table I, Table IV, Table V, and/or Table VI. Preferred genes for use in the present invention include any of the genes presented in regular (non-bold)-type face in Table I or Table V and/or any of the genes in Table VI. The invention also includes the use of nucleic acid molecules derived from or comprising at least a portion of the coding region and/or regulatory region of such genes and any proteins or fragments thereof encoded by such genes. Particularly preferred genes for use in the present invention include any of the genes in Table VI. The invention also includes the use of nucleic acid molecules derived from or comprising at least a portion of the coding region and/or regulatory region of such genes and any proteins or fragments thereof encoded by such genes.

In one embodiment, the invention includes the use of genes encoding any one or more of the following proteins, the genes or nucleic acid sequences therein, or primers used to amplify and identify such genes being identified in Table I and/or Table III and/or Table VI:

rect or indirect mechanisms.

Collectively, the inventors' findings indicate that lumican dEMC-1 function as novel angiogenesis antagonists, while AL359939).

murine ADAMts7 (encoded by a gene comprising the nucleic acid sequence found in GenBank Accession No. AL359939).

human ADAMts7 (encoded by a gene comprising the nucleic acid sequence found in GenBank Accession No. AF140675).

murine CRELD-2 or the human equivalent thereof (murine CRELD-2 encoded by a gene comprising the nucleic acid sequence found in GenBank Accession No. AK017880),

murine Decorin (encoded by a gene comprising the nucleic acid sequence found in GenBank Accession No. NM_007833),

human Decorin (encoded by a gene comprising the nucleic acid sequence found in GenBank Accession No. AH002681),

murine ECM1 (encoded by a gene comprising the nucleic acid sequence found in GenBank Accession No. NM 007899).

human ECM1 (encoded by a gene comprising the nucleic acid sequence found in GenBank Accession No. 5 NP_001415),

murine Inhibin β -b (encoded by a gene comprising the nucleic acid sequence represented herein by SEQ ID NO:97 or SEQ ID NO:98)

human Inhibin β -b (encoded by a gene comprising the 10 nucleic acid sequence found in GenBank Accession No. NM_002193),

murine Integrin α -3 (encoded by a gene comprising the nucleic acid sequence represented herein by SEQ ID NO:99 or SEQ ID NO:100),

human Integrin α -3 (encoded by a gene comprising the nucleic acid sequence found in GenBank Accession No. E16082),

murine Integrin α -6 (encoded by a gene comprising the nucleic acid sequence represented herein by SEQ ID NO:101 20 or SEQ ID NO:102),

human Integrin α -6 (encoded by a gene comprising the nucleic acid sequence found in, for example, GenBank Accession No. AH008066),

murine Lipocalin-7 (encoded by a gene comprising the 25 nucleic acid sequence found in GenBank Accession No. BC005738 and represented herein by SEQ ID NO:103 or SEQ ID NO:104),

human Lipocalin-7 (encoded by a gene comprising the nucleic acid sequence found in GenBank Accession No. 30 NM_022164),

murine Lox1-3 (encoded by a gene comprising the nucleic acid sequence found in GenBank Accession No. NM_013586, the amino acid sequence encoded by which is represented herein by SEQ ID NO:40),

human Lox1-3 (encoded by a gene comprising the nucleic acid sequence found in GenBank Accession No. AAH71865, the amino acid sequence encoded by which is represented herein by SEQ ID NO:41),

murine Lumican (encoded by a gene comprising the 40 nucleic acid sequence found in GenBank Accession No. AK014312),

human Lumican (encoded by a gene comprising the nucleic acid sequence found in GenBank Accession No. AF239660),

murine MAGP-2 (encoded by a gene comprising the nucleic acid sequence found in GenBank Accession No. NM_015776 and represented herein by SEQ ID NO:123, the amino acid sequence encoded by which is represented herein by SEQ ID NO:42),

human MAGP-2 (encoded by a gene comprising the nucleic acid sequence found in GenBank Accession No. AAC83942 and represented herein by SEQ ID NO:124, the amino acid sequence encoded by which is represented herein by SEQ ID NO:43),

murine Matrilin-2 (encoded by a gene comprising the nucleic acid sequence found in GenBank Accession No. BC005429),

human Matrilin-2 (encoded by a gene comprising the nucleic acid sequence found in GenBank Accession No. 60 BC010444)

murine Nephronectin (encoded by a gene comprising the nucleic acid sequence found in GenBank Accession No. AA223007 the amino acid sequence encoded by which is represented herein by SEQ ID NO:52),

human Nephronectin (encoded by a gene comprising the nucleic acid sequence found in GenBank Accession No.

16

NM_001033047, the amino acid sequence encoded by which is represented herein by SEQ ID NO:53),

murine SerpinE2 (encoded by a gene comprising the nucleic acid sequence found in GenBank Accession No. NM_009255),

human SerpinE2 (encoded by a gene comprising the nucleic acid sequence found in GenBank Accession No. BC042628),

murine SMOC-2 (encoded by a gene comprising the nucleic acid sequence found in GenBank Accession No. NM_022315), and

human SMOC-2 (encoded by a gene comprising the nucleic acid sequence found in GenBank Accession No. NM_022138).

The invention also includes the use of nucleic acid molecules derived from or comprising at least a portion of the coding region and/or regulatory region of such genes and any proteins or fragments thereof encoded by such genes, as well as agonists and antagonists of any of such proteins or genes.

In another embodiment, the invention includes the use of genes from Table V encoding any one or more of the following proteins:

murine 0610007C21Rik (GenBank Accession No. AK002276; encoding a protein represented herein by SEQ ID NO:1);

human apoptosis related protein APR-3 (GenBank Accession No. AF144055; encoding a protein represented herein by SEQ ID NO:2);

murine 1810014L12Rik (GenBank Accession No. NM_133706; encoding a protein represented herein by SEQ ID NO:3);

human 1810014L12Rik (GenBank Accession No. NP_055388; encoding a protein represented herein by SEQ ID NO:4);

murine Cd14 (GenBank Accession No. NM_009841; encoding CD14 antigen represented herein by SEQ ID NO:5):

human Cd14 (GenBank Accession No. NP_000638; encoding CD14 antigen represented herein by SEQ ID NO:6);

murine Cd38 (GenBank Accession No. BB256012; comprising a nucleic acid sequence represented herein by SEQ ID NO:7 and encoding CD38 antigen);

murine Cd53 (GenBank Accession No. NM_007651; 45 encoding CD53 antigen represented herein by SEQ ID NO:8);

human Cd53 (GenBank Accession No. NP_000551; encoding CD53 antigen represented herein by SEQ ID NO:9);

murine Emp2 (GenBank Accession No. AF083076; encoding epithelial membrane protein represented herein by SEQ ID NO:10);

human Emp2 (GenBank Accession No. NP_001415; encoding epithelial membrane protein represented herein by 55 SEQ ID NO:11);

murine Fcgrt (GenBank Accession No. NM_010189; encoding Fc receptor (IgG, alpha chain transporter) represented herein by SEQ ID NO:12);

human Fcgrt (GenBank Accession No. NP_004098; encoding Fc receptor (IgG, alpha chain transporter) represented herein by SEQ ID NO:13);

murine Islr (GenBank Accession No. NM_012043; encoding immunoglobulin superfamily containing leucinerich repeat represented herein by SEQ ID NO:14);

human Islr (GenBank Accession No. NP_005536; encoding immunoglobulin superfamily containing leucine-rich repeat represented herein by SEQ ID NO:15);

murine Lrp2 (GenBank Accession No. C80829; comprising a nucleic acid sequence represented herein by SEQ ID NO:16 and encoding low density lipoprotein receptor-related protein 2);

human Lrp2 (GenBank Accession No. NP_004516; comprising a nucleic acid sequence represented herein by SEQ ID NO:17 and encoding low density lipoprotein receptor-related protein 2);

murine Ly6a (GenBank Accession No. BC002070; encoding lymphocyte antigen 6 complex, locus A represented 10 herein by SEQ ID NO:18);

murine P2rx4 (GenBank Accession No. AJ251462; encoding purinergic receptor P2X, ligand-gated ion channel 4, represented herein by SEQ ID NO:19);

human P2rx4 (GenBank Accession No. Q99571; encoding 15 purinergic receptor P2X, ligand-gated ion channel 4, represented herein by SEQ ID NO:20);

murine Pcdhb9 (GenBank Accession No. NM_053134; encoding protocadherin beta 9 represented herein by SEQ ID NO:21):

human Pcdhb9 (GenBank Accession No. AA103495; encoding protocadherin beta 9 represented herein by SEQ ID NO:22);

murine Ptpre (GenBank Accession No. U35368; encoding protein tyrosine phosphatase receptor type E represented 25 herein by SEQ ID NO:23);

human Ptpre (GenBank Accession No. NP_569119; encoding protein tyrosine phosphatase receptor type E represented herein by SEQ ID NO:24);

murine Slc4a3 (GenBank Accession No. NM_009208; 30 encoding solute carrier family 4 (anion exchanger) member 3, represented herein by SEQ ID NO:25);

human Slc4a3 (GenBank Accession No. NP_005061; encoding solute carrier family 4 (anion exchanger) member 3, represented herein by SEQ ID NO:26);

murine Tmc6 (GenBank Accession No. BC004840; encoding transmembrane channel-like gene family 6 represented herein by SEQ ID NO:27).

and/or human Tmc6 (GenBank Accession No. AAH35648; encoding transmembrane channel-like gene 40 family 6 represented herein by SEQ ID NO:28).

The invention also includes the use of nucleic acid molecules derived from or comprising at least a portion of the coding region and/or regulatory region of such genes and any proteins or fragments thereof encoded by such genes, as well 45 as agonists and antagonists of any of such proteins or genes.

In another embodiment, the invention includes the use of genes from Table I encoding any one or more of the following proteins:

murine 9130213B05Rik (GenBank Accession No. 50 BC006604; encoding a protein represented herein by SEQ ID NO:29);

murine C1s (GenBank Accession No. BC022123; encoding complement component 1, s subcomponent, represented herein by SEQ ID NO:34);

human C1s (GenBank Accession No. NM_001734; encoding complement component 1, s subcomponent, represented herein by SEQ ID NO:35);

murine C3 (GenBank Accession No. K02782; encoding complement component 3 represented herein by SEQ ID 60 NO:30);

human C3 (GenBank Accession No. NP_000055; encoding complement component 3 represented herein by SEQ ID NO:31);

murine Cfh (GenBank Accession No. AI987976; comprising a nucleic acid sequence represented herein by SEQ ID NO:32 and encoding complement component factor h);

18

human Cfh (GenBank Accession No. CAA30403; comprising a nucleic acid sequence represented herein by SEQ ID NO:33 and encoding complement component factor h);

murine Col9a3 (GenBank Accession No. BG074456; comprising a nucleic acid sequence represented herein by SEQ ID NO:36 and encoding procollagen, type IX, alpha 3):

human Col9a3 (GenBank Accession No. Q14050; comprising a nucleic acid sequence represented herein by SEQ ID NO:37 and encoding procollagen, type IX, alpha 3);

murine Grem1 (GenBank Accession No. BC015293; encoding cysteine knot superfamily 1, BMP antagonist 1, represented herein by SEQ ID NO:38);

human Grem1 (GenBank Accession No. NP_037504; encoding cysteine knot superfamily 1, BMP antagonist 1, represented herein by SEQ ID NO:39);

murine Lox13 (GenBank Accession No. NM_013586; encoding lysyl oxidase-like 3, represented herein by SEQ ID NO:40);

human Lox13 (GenBank Accession No. AAH71865; encoding lysyl oxidase-like 3, represented herein by SEQ ID NO:41);

murine MAGP-2 (GenBank Accession No. NM_015776; comprising a nucleic acid sequence represented herein by SEQ ID NO:123 and encoding microfibril-associated glycoprotein-2 (also known as microfibrillar associated protein 5), represented herein by SEQ ID NO:42);

human MAGP-2 (GenBank Accession No. AAC83942; comprising a nucleic acid sequence represented herein by SEQ ID NO:124 and encoding microfibrillar associated protein 5, represented herein by SEQ ID NO:43);

murine Mglap (GenBank Accession No. NM_008597; encoding matrix gamma-carboxyglutamate (gla) protein represented herein by SEQ ID NO:44);

human Mglap (GenBank Accession No. AAP36640; encoding matrix gamma-carboxyglutamate (gla) protein represented herein by SEQ ID NO:45);

murine Naga (GenBank Accession No. BC021631; encoding N-acetyl galactosaminidase, alpha, represented herein by SEQ ID NO:46);

human Naga (GenBank Accession No. NP_000253; encoding N-acetyl galactosaminidase, alpha, represented herein by SEQ ID NO:47);

murine Nbl1 (GenBank Accession No. NM_008675; encoding neuroblastoma, suppression of tumorigenicity 1, represented herein by SEQ ID NO:48);

human Nb11 (GenBank Accession No. AAL15440; encoding neuroblastoma, suppression of tumorigenicity 1, represented herein by SEQ ID NO:49);

murine Ngfb (GenBank Accession No. NM_013609; encoding nerve growth factor, beta, represented herein by SEQ ID NO:50);

human Ngfb (GenBank Accession No. AAH32517; encoding nerve growth factor, beta, represented herein by SEQ ID NO:51):

murine Npnt (GenBank Accession No. AA223007; encoding nephronectin and represented herein by SEQ ID NO:52);

human Npnt (GenBank Accession No. NM_001033047; encoding nephronectin and represented herein by SEQ ID NO:53);

murine Olfm1 (GenBank Accession No. C78264; encoding olfactomedin 1, represented herein by SEQ ID NO:54);

human Olfm1 (GenBank Accession No. Q99784; encoding olfactomedin 1, represented herein by SEQ ID NO:55);

and/or murine U90926 (GenBank Accession No. NM_020562; encoding a protein represented herein by SEQ ID NO:56).

The invention also includes the use of nucleic acid molecules derived from or comprising at least a portion of the coding region and/or regulatory region of such genes and any proteins or fragments thereof encoded by such genes.

The genes identified in the Tables herein are identified by 5 name, by GenBank Accession numbers, and by description of the protein, when available. The amino acid sequence for several of the proteins encoded by the genes in the Tables herein are also provided herein. All information associated with the publicly available identifiers and accession numbers 10 in any of the tables described herein, including the nucleic acid sequences of the genes and probes and the amino acid sequences of the proteins encoded thereby, is incorporated herein by reference in its entirety.

Genes and proteins identified in the present invention can 15 also be referred to as "biomarkers". The term "biomarker" as used herein can refer to gene described herein or to the protein encoded by that gene, wherein the gene has been identified as being differentially regulated during angiogenesis. In addition, the term "biomarker" can be generally used to refer to 20 any portion of such a gene or protein that can identify or correlate with the full-length gene or protein, for example, in an assay or other method of the invention.

Microfibril-associated glycoprotein-2 (MAGP-2) is a secreted glycoprotein (25 kDa) that incorporates into and 25 organizes elastin fibril networks by interacting with tropoelastin, and with fibrillins 1 and 2; it also mediates cell adhesion by ligating integrins via its RGD integrin-binding motif (Gibson et al, 1998; Gibson et al, 1999). Abnormally elevated MAGP-2 expression is observed in the skin of systemic sclerosis patients, as well as in mouse models of systemic sclerosis that have associated MAGP-2 expression with excessive matrix deposition of type I collagen (Lemaire et al, 2004; Lemaire et al, 2005). Moreover, skin lesions in systemic sclerosis patients contain aberrant vessel morphologies 35 characteristic of abnormal angiogenesis (Bodolay et al, 2002). In addition, MAGP-2 expression is induced in human T-47DE3 breast cancer cells when treated with progestin (Graham et al, 2005), and in human A549 lung adenocarcinoma cells when implanted into nude mice (Creighton et al, 40 2003). Most recently, MAGP-2 has been shown to interact physically with Notch1 (Miyamoto et al, 2006) and its ligand, Jagged-1 (Nehring et al, 2005), leading to the ectodomain shedding of both molecules from the cell surface.

Human MAGP-2 cDNA has been cloned and described, for 45 example, in Faraco et al. (Genomics. 1995 Feb. 10; 25(3):630-7) and in Gibson et al. (J Biol Chem. 1996 Jan. 12; 271(2): 1096-103). The organization of the human MAGP-2 gene is described in Hatzinikolas and Gibson (J Biol Chem. 1998 Nov. 6; 273(45):29309-14). The organization of the mouse 50 MAGP-2 gene has been described by Frankfater et al. (Mamm Genome. 2000 Mar.; 11(3):191-5). The nucleotide sequence encoding human MAGP-2 is described in the National Center for Biotechnology Information (NCBI) database Accession No. AH007047 (gi:3983462) and is represented herein by 55 SEQ ID NO:124. The amino acid sequence for human MAGP-2 is represented herein as SEQ ID NO:43 and is also found in the NCBI database Accession No. AAC83942 (gi: 3983463). The nucleotide sequences encoding bovine and murine MAGP-2 are also known. The nucleotide sequence 60 encoding murine MAGP-2 is described in NCBI database Accession No. BC025131 (gi: 19264044) and is represented herein by SEQ ID NO:123 and encodes the murine MAGP-2 protein, described in NCBI database Accession No. AAH25131 (gi:19264045), also represented herein by SEQ ID NO:42. The nucleotide sequence encoding bovine MAGP-2 is described in NCBI database Accession No.

20

NM_174386 (gi:31342148) and encodes the bovine MAGP-2 protein, described in NCBI database Accession No. NP_776811 (gi:27805993). All of the information contained in the database accession numbers and in the publications referenced herein is incorporated herein by reference.

In accordance with the present invention, an isolated polynucleotide (also referred to as an isolated nucleic acid molecule) is a nucleic acid molecule that has been removed from its natural milieu (e.g., that has been subject to human manipulation), its natural milieu being the genome or chromosome in which the nucleic acid molecule is found in nature. As such, "isolated" does not necessarily reflect the extent to which the nucleic acid molecule has been purified, but indicates that the molecule does not include an entire genome or an entire chromosome in which the nucleic acid molecule is found in nature. The polynucleotides useful in the present invention are typically a portion of a gene (sense or non-sense strand) of the present invention that is suitable for use as a hybridization probe or PCR primer for the identification of a full-length gene (or portion thereof) in a given sample, to encode a protein or fragment thereof, or as a therapeutic reagent (e.g., antisense). An isolated nucleic acid molecule can include a gene or a portion of a gene (e.g., the regulatory region or promoter), for example, to produce a reporter construct according to the present invention. An isolated nucleic acid molecule that includes a gene is not a fragment of a chromosome that includes such gene, but rather includes the coding region and regulatory regions associated with the gene, but no additional genes naturally found on the same chromosome. An isolated nucleic acid molecule can also include a specified nucleic acid sequence flanked by (i.e., at the 5' and/or the 3' end of the sequence) additional nucleic acids that do not normally flank the specified nucleic acid sequence in nature (i.e., heterologous sequences). Isolated nucleic acid molecule can include DNA, RNA (e.g., mRNA), or derivatives of either DNA or RNA (e.g., cDNA). Although the phrase "nucleic acid molecule" primarily refers to the physical nucleic acid molecule and the phrase "nucleic acid sequence" primarily refers to the sequence of nucleotides on the nucleic acid molecule, the two phrases can be used interchangeably, especially with respect to a nucleic acid molecule, or a nucleic acid sequence, being capable of encoding a protein. Preferably, an isolated nucleic acid molecule of the present invention is produced using recombinant DNA technology (e.g., polymerase chain reaction (PCR) amplification, cloning) or chemical synthesis.

The minimum size of a nucleic acid molecule or polynucleotide of the present invention is a size sufficient to encode a protein having a desired biological activity, sufficient to form a probe or oligonucleotide primer that is capable of forming a stable hybrid with the complementary sequence of a nucleic acid molecule encoding the natural protein (e.g., under moderate, high or very high stringency conditions), or to otherwise be used as a target or agent in an assay or in any therapeutic method discussed herein. If the polynucleotide is an oligonucleotide probe or primer, the size of the polynucleotide can be dependent on nucleic acid composition and percent homology or identity between the nucleic acid molecule and a complementary sequence as well as upon hybridization conditions per se (e.g., temperature, salt concentration, and formamide concentration). The minimum size of a polynucleotide that is used as an oligonucleotide probe or primer is at least about 5 nucleotides in length, and preferably ranges from about 5 to about 50 or about 500 nucleotides or greater (1000, 2000, etc.), including any length in between, in whole number increments (i.e., 5, 6, 7, 8, 9, 10, ... 33, 34, ... 256, $257, \ldots 500 \ldots 1000 \ldots$), and more preferably from about

10 to about 40 nucleotides, and most preferably from about 15 to about 40 nucleotides in length. In one aspect, the oligonucleotide primer or probe is typically at least about 12 to about 15 nucleotides in length if the nucleic acid molecules are GC-rich and at least about 15 to about 18 bases in length if they are AT-rich. There is no limit, other than a practical limit, on the maximal size of a nucleic acid molecule of the present invention, in that the nucleic acid molecule can include a portion of a protein-encoding sequence or a nucleic acid sequence encoding a full-length protein.

According to the present invention, an oligonucleotide probe (or simply, probe) is a nucleic acid molecule which most typically ranges in size from about 8 nucleotides to several hundred nucleotides in length. Such a molecule is typically used to identify a target nucleic acid sequence in a 15 sample by hybridizing to such target nucleic acid sequence under stringent hybridization conditions. As used herein, stringent hybridization conditions refer to standard hybridization conditions under which nucleic acid molecules are used to identify similar nucleic acid molecules. Such standard 20 conditions are disclosed, for example, in Sambrook et al., Molecular Cloning: A Laboratory Manual, Cold Spring Harbor Labs Press, 1989. Sambrook et al., ibid., is incorporated by reference herein in its entirety (see specifically, pages 9.31-9.62). In addition, formulae to calculate the appropriate 25 hybridization and wash conditions to achieve hybridization permitting varying degrees of mismatch of nucleotides are disclosed, for example, in Meinkoth et al., 1984, Anal. Biochem. 138, 267-284; Meinkoth et al., ibid., is incorporated by reference herein in its entirety.

More particularly, moderate stringency hybridization and washing conditions, as referred to herein, refer to conditions which permit isolation of nucleic acid molecules having at least about 70% nucleic acid sequence identity with the nucleic acid molecule being used to probe in the hybridiza- 35 tion reaction (i.e., conditions permitting about 30% or less mismatch of nucleotides). High stringency hybridization and washing conditions, as referred to herein, refer to conditions which permit isolation of nucleic acid molecules having at least about 80% nucleic acid sequence identity with the 40 nucleic acid molecule being used to probe in the hybridization reaction (i.e., conditions permitting about 20% or less mismatch of nucleotides). Very high stringency hybridization and washing conditions, as referred to herein, refer to conditions which permit isolation of nucleic acid molecules having 45 at least about 90% nucleic acid sequence identity with the nucleic acid molecule being used to probe in the hybridization reaction (i.e., conditions permitting about 10% or less mismatch of nucleotides). As discussed above, one of skill in the art can use the formulae in Meinkoth et al., ibid. to calcu- 50 late the appropriate hybridization and wash conditions to achieve these particular levels of nucleotide mismatch. Such conditions will vary, depending on whether DNA:RNA or DNA:DNA hybrids are being formed. Calculated melting temperatures for DNA:DNA hybrids are 10° C. less than for 55 DNA:RNA hybrids. In particular embodiments, stringent hybridization conditions for DNA:DNA hybrids include hybridization at an ionic strength of 6×SSC (0.9 M Na⁺) at a temperature of between about 20° C. and about 35° C. (lower stringency), more preferably, between about 28° C. and about 60 40° C. (more stringent), and even more preferably, between about 35° C. and about 45° C. (even more stringent), with appropriate wash conditions. In particular embodiments, stringent hybridization conditions for DNA:RNA hybrids include hybridization at an ionic strength of 6×SSC (0.9 M 65 Na⁺) at a temperature of between about 30° C. and about 45° C., more preferably, between about 38° C. and about 50° C.,

22

and even more preferably, between about 45° C. and about 55° C., with similarly stringent wash conditions. These values are based on calculations of a melting temperature for molecules larger than about 100 nucleotides, 0% formamide and a G+C content of about 40%. Alternatively, T_m can be calculated empirically as set forth in Sambrook et al., supra, pages 9.31 to 9.62. In general, the wash conditions should be as stringent as possible, and should be appropriate for the chosen hybridization conditions. For example, hybridization conditions can include a combination of salt and temperature conditions that are approximately 20-25° C. below the calculated T_m of a particular hybrid, and wash conditions typically include a combination of salt and temperature conditions that are approximately 12-20° C. below the calculated T_m of the particular hybrid. One example of hybridization conditions suitable for use with DNA: DNA hybrids includes a 2-24 hour hybridization in 6×SSC (50% formamide) at about 42° C., followed by washing steps that include one or more washes at room temperature in about 2×SSC, followed by additional washes at higher temperatures and lower ionic strength (e.g., at least one wash as about 37° C. in about 0.1×-0.5×SSC, followed by at least one wash at about 68° C. in about 0.1×- $0.5 \times SSC$).

PCR primers are also nucleic acid sequences, although PCR primers are typically oligonucleotides of fairly short length that are used in polymerase chain reactions. PCR primers and hybridization probes can readily be developed and produced by those of skill in the art, using sequence information from the target sequence. (See, for example, Sambrook et al., supra or Glick et al., supra).

Knowing the nucleic acid sequences of certain nucleic acid molecules of the present invention allows one skilled in the art to, for example, (a) make copies of those nucleic acid molecules and/or (b) obtain nucleic acid molecules including at least a portion of such nucleic acid molecules (e.g., nucleic acid molecules including full-length genes, full-length coding regions, regulatory control sequences, truncated coding regions). Such nucleic acid molecules can be obtained in a variety of ways including traditional cloning techniques using oligonucleotide probes to screen appropriate libraries or DNA and PCR amplification of appropriate libraries or DNA using oligonucleotide primers. Preferred libraries to screen or from which to amplify nucleic acid molecule include mammali7an genomic DNA libraries. Techniques to clone and amplify genes are disclosed, for example, in Sambrook et al., ibid.

As used herein, reference to an isolated protein or polypeptide in the present invention, including any of the proteins described particularly herein (e.g., any protein encoded by a gene or nucleic acid sequence referenced in Table I, Table IV, Table V, and/or Table VI), includes full-length proteins, fusion proteins, or any fragment or homologue of such a protein. Such a protein can include, but is not limited to, purified proteins, recombinantly produced proteins, membrane bound proteins, proteins complexed with lipids, soluble proteins and isolated proteins associated with other proteins. More specifically, an isolated protein, such as a MAGP-2 (MFAP-5) protein, by way of example, according to the present invention, is a protein (including a polypeptide or peptide) that has been removed from its natural milieu (i.e., that has been subject to human manipulation) and can include purified proteins, partially purified proteins, recombinantly produced proteins, and synthetically produced proteins, for example. As such, "isolated" does not reflect the extent to which the protein has been purified. Preferably, an isolated protein of the present invention is produced recombinantly. In addition, and again by way of example, a "human MAGP-2

protein" or a protein "derived from" a human MAGP-2 protein refers to a MAGP-2 protein (generally including a homologue of a naturally occurring MAGP-2 protein) from a human (*Homo sapiens*) or to a MAGP-2 protein that has been otherwise produced from the knowledge of the structure (e.g., 5 sequence) and perhaps the function of a naturally occurring MAGP-2 protein from Homo sapiens. In other words, a human MAGP-2 protein includes any MAGP-2 protein that has substantially similar structure and function of a naturally occurring MAGP-2 protein from Homo sapiens or that is a 10 biologically active (i.e., has biological activity) homologue of a naturally occurring MAGP-2 protein from Homo sapiens as described in detail herein. As such, a human MAGP-2 protein can include purified, partially purified, recombinant, mutated/modified and synthetic proteins. According to the 15 present invention, the terms "modification" and "mutation" can be used interchangeably, particularly with regard to the modifications/mutations to the amino acid sequence of protein (or nucleic acid sequences) described herein. An isolated protein useful as an antagonist or agonist according to the 20 present invention can be isolated from its natural source, produced recombinantly or produced synthetically.

As used herein, the term "homologue" is used to refer to a protein or peptide which differs from a naturally occurring protein or peptide (i.e., the "prototype" or "wild-type" pro- 25 tein) by minor modifications to the naturally occurring protein or peptide, but which maintains the basic protein and side chain structure of the naturally occurring form. Such changes include, but are not limited to: changes in one or a few amino acid side chains; changes one or a few amino acids, including 30 deletions (e.g., a truncated version of the protein or peptide) insertions and/or substitutions; changes in stereochemistry of one or a few atoms; and/or minor derivatizations, including but not limited to: methylation, glycosylation, phosphorylation, acetylation, myristoylation, prenylation, palmitation, 35 amidation and/or addition of glycosylphosphatidyl inositol. A homologue can have either enhanced, decreased, or substantially similar properties as compared to the naturally occurring protein or peptide. A homologue can include an agonist of a protein or an antagonist of a protein.

Homologues can be the result of natural allelic variation or natural mutation. A naturally occurring allelic variant of a nucleic acid encoding a protein is a gene that occurs at essentially the same locus (or loci) in the genome as the gene which encodes such protein, but which, due to natural variations 45 caused by, for example, mutation or recombination, has a similar but not identical sequence. Allelic variants typically encode proteins having similar activity to that of the protein encoded by the gene to which they are being compared. One class of allelic variants can encode the same protein but have 50 different nucleic acid sequences due to the degeneracy of the genetic code. Allelic variants can also comprise alterations in the 5' or 3' untranslated regions of the gene (e.g., in regulatory control regions). Allelic variants are well known to those skilled in the art.

Homologues can be produced using techniques known in the art for the production of proteins including, but not limited to, direct modifications to the isolated, naturally occurring protein, direct protein synthesis, or modifications to the nucleic acid sequence encoding the protein using, for 60 example, classic or recombinant DNA techniques to effect random or targeted mutagenesis.

According to the present invention, an isolated protein, including a biologically active homologue or fragment thereof, has at least one characteristic of biological activity of 65 activity the wild-type, or naturally occurring reference protein (which can vary depending on whether the homologue or

24

fragment is an agonist or antagonist of the protein, or whether an agonist or antagonist mimetic of the protein is described). In general, the biological activity or biological action of a protein refers to any function(s) exhibited or performed by the protein that is ascribed to the naturally occurring form of the protein as measured or observed in vivo (i.e., in the natural physiological environment of the protein) or in vitro (i.e., under laboratory conditions). Modifications, activities or interactions which result in a decrease in protein expression or a decrease in the activity of the protein, can be referred to as inactivation (complete or partial), down-regulation, reduced action, or decreased action or activity of a protein. Similarly, modifications, activities or interactions which result in an increase in protein expression or an increase in the activity of the protein, can be referred to as amplification, overproduction, activation, enhancement, up-regulation or increased action of a protein. The biological activity of a protein according to the invention can be measured or evaluated using any assay for the biological activity of the protein as known in the art. Such assays can include, but are not limited to, binding assays, assays to determine internalization of the protein and/or associated proteins, enzyme assays, cell signal transduction assays (e.g., phosphorylation assays), and/or assays for determining downstream cellular events that result from activation or binding of the cell surface protein (e.g., expression of downstream genes, production of various biological mediators, etc.).

As used herein, reference to an "agonist" of a given protein refers to any compound that is characterized by the ability to agonize (e.g., stimulate, induce, increase, enhance, or mimic) the biological activity of the naturally occurring protein, and includes any homologue, binding protein (e.g., an antibody), agent that interacts with a protein or receptor bound by the protein, or any suitable product of drug/compound/peptide design or selection which is characterized by its ability to agonize (e.g., stimulate, induce, increase, enhance) the biological activity of the naturally occurring protein in a manner similar to the natural agonist, which is the reference protein.

Similarly, reference to an "antagonist" refers to any compound which inhibits (e.g., antagonizes, reduces, decreases, blocks, reverses, or alters) the effect of a given agonist of a protein (including the protein itself) as described above. More particularly, an antagonist is capable of acting in a manner relative to the activity of the protein, such that the biological activity of the natural agonist or reference protein, is decreased in a manner that is antagonistic (e.g., against, a reversal of, contrary to) to the natural action of the protein. Such antagonists can include, but are not limited to, a protein, peptide, or nucleic acid (including ribozymes, RNAi, aptamers, and antisense), antibodies and antigen binding fragments thereof, or product of drug/compound/peptide design or selection that provides the antagonistic effect.

As used herein, an anti-sense nucleic acid molecule is defined as an isolated nucleic acid molecule that reduces expression of a protein by hybridizing under high stringency conditions to a gene encoding the protein. Such a nucleic acid molecule is sufficiently similar to the gene encoding the protein that the molecule is capable of hybridizing under high stringency conditions to the coding or complementary strand of the gene or RNA encoding the natural protein. RNA interference (RNAi) is a process whereby double stranded RNA, and in mammalian systems, short interfering RNA (siRNA), is used to inhibit or silence expression of complementary genes. In the target cell, siRNA are unwound and associate with an RNA induced silencing complex (RISC), which is then guided to the mRNA sequences that are complementary to the siRNA, whereby the RISC cleaves the mRNA. A

ribozyme is an RNA segment that functions by binding to the target RNA moiety and inactivate it by cleaving the phosphodiester backbone at a specific cutting site. A ribozyme can serve as a targeting delivery vehicle for a nucleic acid molecule, or alternatively, the ribozyme can target and bind to 5 RNA encoding the biomarker, for example, and thereby effectively inhibit the translation of the biomarker. Aptamers are short strands of synthetic nucleic acids (usually RNA but also DNA) selected from randomized combinatorial nucleic acid libraries by virtue of their ability to bind to a predetermined specific target molecule with high affinity and specificity. Aptamers assume a defined three-dimensional structure and are capable of discriminating between compounds with very small differences in structure.

Homologues of a given protein, including peptide and non-peptide agonists and antagonists (analogs), can be products of drug design or selection and can be produced using various methods known in the art. Such homologues can be referred to as mimetics. Various methods of drug design, useful to design or select mimetics or other therapeutic compounds 20 useful in the present invention are disclosed in Maulik et al., 1997, *Molecular Biotechnology: Therapeutic Applications and Strategies*, Wiley-Liss, Inc., which is incorporated herein by reference in its entirety.

As used herein, a mimetic refers to any peptide or non- 25 peptide compound that is able to mimic the biological action of a naturally occurring peptide, often because the mimetic has a basic structure that mimics the basic structure of the naturally occurring peptide and/or has the salient biological properties of the naturally occurring peptide. Mimetics can 30 include, but are not limited to: peptides that have substantial modifications from the prototype such as no side chain similarity with the naturally occurring peptide (such modifications, for example, may decrease its susceptibility to degradation); anti-idiotypic and/or catalytic antibodies, or 35 fragments thereof; non-proteinaceous portions of an isolated protein (e.g., carbohydrate structures); or synthetic or natural organic molecules, including nucleic acids and drugs identified through combinatorial chemistry, for example. Such mimetics can be designed, selected and/or otherwise identi- 40 fied using a variety of methods known in the art.

A mimetic can be obtained, for example, from molecular diversity strategies (a combination of related strategies allowing the rapid construction of large, chemically diverse molecule libraries), libraries of natural or synthetic compounds, 45 in particular from chemical or combinatorial libraries (i.e., libraries of compounds that differ in sequence or size but that have the similar building blocks) or by rational, directed or random drug design. See for example, Maulik et al., supra.

In a molecular diversity strategy, large compound libraries 50 are synthesized, for example, from peptides, oligonucleotides, carbohydrates and/or synthetic organic molecules, using biological, enzymatic and/or chemical approaches. The critical parameters in developing a molecular diversity strategy include subunit diversity, molecular size, and library 55 diversity. The general goal of screening such libraries is to utilize sequential application of combinatorial selection to obtain high-affinity ligands for a desired target, and then to optimize the lead molecules by either random or directed design strategies. Methods of molecular diversity are 60 described in detail in Maulik, et al., ibid.

In a rational drug design procedure, the three-dimensional structure of a regulatory compound can be analyzed by, for example, nuclear magnetic resonance (NMR) or X-ray crystallography. This three-dimensional structure can then be 65 used to predict structures of potential compounds, such as potential regulatory agents by, for example, computer mod-

eling. The predicted compound structure can be used to optimize lead compounds derived, for example, by molecular diversity methods. In addition, the predicted compound structure can be produced by, for example, chemical synthesis, recombinant DNA technology, or by isolating a mimetope from a natural source (e.g., plants, animals, bacteria and fungi).

26

Maulik et al. also disclose, for example, methods of directed design, in which the user directs the process of creating novel molecules from a fragment library of appropriately selected fragments; random design, in which the user uses a genetic or other algorithm to randomly mutate fragments and their combinations while simultaneously applying a selection criterion to evaluate the fitness of candidate ligands; and a grid-based approach in which the user calculates the interaction energy between three dimensional receptor structures and small fragment probes, followed by linking together of favorable probe sites.

In one embodiment, a homologue of a given protein comprises, consists essentially of, or consists of, an amino acid sequence that is at least about 45%, or at least about 50%, or at least about 55%, or at least about 60%, or at least about 65%, or at least about 70%, or at least about 75%, or at least about 80%, or at least about 85%, or at least about 90%, or at least about 95% identical, or at least about 95% identical, or at least about 96% identical, or at least about 97% identical, or at least about 98% identical, or at least about 99% identical (or any percent identity between 45% and 99%, in whole integer increments), to the amino acid sequence of the reference protein. In one embodiment, the homologue comprises, consists essentially of, or consists of, an amino acid sequence that is less than 100% identical, less than about 99% identical, less than about 98% identical, less than about 97% identical, less than about 96% identical, less than about 95% identical; and so on, in increments of 1%, to less than about 70% identical to the naturally occurring amino acid sequence of the reference protein.

As used herein, unless otherwise specified, reference to a percent (%) identity refers to an evaluation of homology which is performed using: (1) a BLAST 2.0 Basic BLAST homology search using blastp for amino acid searches and blastn for nucleic acid searches with standard default parameters, wherein the query sequence is filtered for low complexity regions by default (described in Altschul, S. F., Madden, T. L., Schääffer, A. A., Zhang, J., Zhang, Z., Miller, W. & Lipman, D. J. (1997) "Gapped BLAST and PSI-BLAST: a new generation of protein database search programs." Nucleic Acids Res. 25:3389-3402, incorporated herein by reference in its entirety); (2) a BLAST 2 alignment (using the parameters described below); (3) and/or PSI-BLAST with the standard default parameters (Position-Specific Iterated BLAST. It is noted that due to some differences in the standard parameters between BLAST 2.0 Basic BLAST and BLAST 2, two specific sequences might be recognized as having significant homology using the BLAST 2 program, whereas a search performed in BLAST 2.0 Basic BLAST using one of the sequences as the query sequence may not identify the second sequence in the top matches. In addition, PSI-BLAST provides an automated, easy-to-use version of a "profile" search, which is a sensitive way to look for sequence homologues. The program first performs a gapped BLAST database search. The PSI-BLAST program uses the information from any significant alignments returned to construct a positionspecific score matrix, which replaces the query sequence for the next round of database searching. Therefore, it is to be understood that percent identity can be determined by using any one of these programs.

Two specific sequences can be aligned to one another using BLAST 2 sequence as described in Tatusova and Madden, (1999), "Blast 2 sequences—a new tool for comparing protein and nucleotide sequences", FEMS Microbiol Lett. 174: 247-250, incorporated herein by reference in its entirety. 5 BLAST 2 sequence alignment is performed in blastp or blastn using the BLAST 2.0 algorithm to perform a Gapped BLAST search (BLAST 2.0) between the two sequences allowing for the introduction of gaps (deletions and insertions) in the resulting alignment. For purposes of clarity herein, a BLAST 10 2 sequence alignment is performed using the standard default parameters as follows.

For blastn, using 0 BLOSUM62 matrix:

Reward for match=1

Penalty for mismatch=-2

Open gap (5) and extension gap (2) penalties

gap x_dropoff (50) expect (10) word size (11) filter (on)

For blastp, using 0 BLOSUM62 matrix:

Open gap (11) and extension gap (1) penalties

gap x dropoff (50) expect (10) word size (3) filter (on).

Also included in the present invention are antibodies and antigen binding fragments thereof that selectively bind to any of the proteins associated with angiogenesis described herein, as well as the use of such antibodies and antigen binding fragments thereof in any of the methods described herein. 25 Antibodies that selectively bind to a protein can be produced using the structural information available for the protein (e.g., the amino acid sequence of at least a portion of the protein). More specifically, the phrase "selectively binds" refers to the specific binding of one protein to another (e.g., an antibody, 30 fragment thereof, or binding partner to an antigen), wherein the level of binding, as measured by any standard assay (e.g., an immunoassay), is statistically significantly higher than the background control for the assay. For example, when performing an immunoassay, controls typically include a reac- 35 tion well/tube that contain antibody or antigen binding fragment alone (i.e., in the absence of antigen), wherein an amount of reactivity (e.g., non-specific binding to the well) by the antibody or antigen binding fragment thereof in the absence of the antigen is considered to be background. Bind- 40 ing can be measured using a variety of methods standard in the art including enzyme immunoassays (e.g., ELISA), immunoblot assays, etc.). Antibodies useful in the assay kit and methods of the present invention can include polyclonal and monoclonal antibodies, divalent and monovalent anti- 45 bodies, bi- or multi-specific antibodies, serum containing such antibodies, antibodies that have been purified to varying degrees, and any functional equivalents of whole antibodies. Isolated antibodies of the present invention can include serum containing such antibodies, or antibodies that have been puri- 50 fied to varying degrees. Whole antibodies of the present invention can be polyclonal or monoclonal. Alternatively, functional equivalents of whole antibodies, such as antigen binding fragments in which one or more antibody domains ments), as well as genetically-engineered antibodies or antigen binding fragments thereof, including single chain antibodies or antibodies that can bind to more than one epitope (e.g., bi-specific antibodies), or antibodies that can bind to one or more different antigens (e.g., bi- or multi-specific 60 antibodies), may also be employed in the invention.

Genetically engineered antibodies include those produced by standard recombinant DNA techniques involving the manipulation and re-expression of DNA encoding antibody variable and/or constant regions. Particular examples include, 65 chimeric antibodies, where the V_H and/or V_L domains of the antibody come from a different source to the remainder of the

antibody, and CDR grafted antibodies (and antigen binding fragments thereof), in which at least one CDR sequence and optionally at least one variable region framework amino acid is (are) derived from one source and the remaining portions of the variable and the constant regions (as appropriate) are derived from a different source. Construction of chimeric and CDR-grafted antibodies are described, for example, in European Patent Applications: EP-A 0194276, EP-A 0239400, EP-A 0451216 and EP-A 0460617.

Generally, in the production of an antibody, a suitable experimental animal, such as, for example, but not limited to, a rabbit, a sheep, a hamster, a guinea pig, a mouse, a rat, or a chicken, is exposed to an antigen against which an antibody is desired. Typically, an animal is immunized with an effective 15 amount of antigen that is injected into the animal. An effective amount of antigen refers to an amount needed to induce antibody production by the animal. The animal's immune system is then allowed to respond over a pre-determined period of time. The immunization process can be repeated until the immune system is found to be producing antibodies to the antigen. In order to obtain polyclonal antibodies specific for the antigen, serum is collected from the animal that contains the desired antibodies (or in the case of a chicken, antibody can be collected from the eggs). Such serum is useful as a reagent. Polyclonal antibodies can be further purified from the serum (or eggs) by, for example, treating the serum with ammonium sulfate.

Monoclonal antibodies may be produced according to the methodology of Kohler and Milstein (Nature 256:495-497, 1975). For example, B lymphocytes are recovered from the spleen (or any suitable tissue) of an immunized animal and then fused with myeloma cells to obtain a population of hybridoma cells capable of continual growth in suitable culture medium. Hybridomas producing the desired antibody are selected by testing the ability of the antibody produced by the hybridoma to bind to the desired antigen.

The invention also extends to non-antibody polypeptides, sometimes referred to as antigen binding partners or antigen binding peptides, that have been designed to bind selectively to the protein of interest. Examples of the design of such polypeptides, which possess a prescribed ligand specificity are given in Beste et al. (Proc. Natl. Acad. Sci. 96:1898-1903, 1999), incorporated herein by reference in its entirety.

One embodiment of the present invention relates to a method to identify a compound useful for the inhibition (reduction, decrease) of angiogenesis, which may also be applied to identifying agents useful for inhibition of tumor cell growth, presence, or malignancy. A similar method of the present invention can also be used to identify a compound useful for the promotion (increase, initiation, enhancement) of angiogenesis, which may also be applied to identifying agents useful for conditions in which angiogenesis may be desired (e.g., stroke, ischemia).

Either of such methods generally includes the steps of: (a) are truncated or absent (e.g., Fv, Fab, Fab', or F(ab)₂ frag- 55 detecting an initial level of the expression or activity of one or more genes or proteins encoded thereby (biomarkers) that are associated with angiogenesis as described herein (e.g., any one or more of the genes or the proteins encoded by a gene or nucleic acid sequence referenced in Table I, Table IV, Table V, and/or Table VI, and/or any one or more of the genes or proteins specifically described herein by reference to a particular nucleic acid or amino acid sequence) in a cell or soluble sample or product derived from the cell (e.g., cell supernate); (b) contacting the cell with a test compound; (c) detecting a level of gene or protein expression or activity in the cell (or sample derived therefrom) after contact of the cell with the compound; and, (d) selecting a compound that regu-

lates the level of gene or protein expression or activity in the cell, as compared to prior to contact with the test compound. In one embodiment, the biomarker is a protein, or the gene encoding such protein, selected from: ADAMts7, CRELD-2, Decorin, ECM1, Inhibin β -b, Integrin α -3, Integrin α -6, Lipocalin-7, Lox1-3, Lumican, MAGP-2, Matrilin-2, Nephronectin, SerpinE2, and/or SMOC-2. These genes and proteins have been described in detail above.

In another embodiment, the biomarker is a gene, or the protein encoded by the gene, selected from: 0610007C21Rik, apoptosis related protein APR-3, 1810014L12Rik, Cd14 (encoding CD14 antigen represented herein by SEQ ID NO:5 and SEQ ID NO:6), Cd38 (comprising a nucleic acid sequence represented herein by SEQ ID NO:7 and encoding CD38 antigen); Cd53 (encoding CD53 antigen represented herein by SEQ ID NO:8 and SEQ ID NO:9), Emp2 (encoding epithelial membrane protein represented herein by SEQ ID NO:10 and SEQ ID NO:11), Fcgrt (encoding Fc receptor (IgG, alpha chain transporter) represented herein by SEQ ID 20 NO:12 and SEQ ID NO:13), Islr (encoding immunoglobulin superfamily containing leucine-rich repeat represented herein by SEQ ID NO:14 and SEQ ID NO:15); Lrp2 (comprising a nucleic acid sequence represented herein by SEQ ID NO:16 and SEQ ID NO:17 and encoding low density lipo- 25 protein receptor-related protein 2); Ly6a (encoding lymphocyte antigen 6 complex, locus A represented herein by SEQ ID NO:18); P2rx4 (encoding purinergic receptor P2X, ligand-gated ion channel 4, represented herein by SEQ ID NO:19 and SEQ ID NO:20; Pcdhb9 (encoding protocadherin 30 beta 9 represented herein by SEQ ID NO:21 and SEQ ID NO:22); Ptpre (encoding protein tyrosine phosphatase receptor type E represented herein by SEQ ID NO:23 and SEQ ID NO:24); Slc4a3 (encoding solute carrier family 4 (anion exchanger) member 3, represented herein by SEQ ID NO:25 35 and SEQ ID NO:26); and/or Tmc6 (encoding transmembrane channel-like gene family 6, represented herein by SEQ ID

In yet another embodiment, the biomarker is a gene, or the protein encoded by the gene, selected from: 9130213B05Rik 40 (encoding a protein represented herein by SEQ ID NO:29); C1s (encoding complement component 1, s subcomponent, represented herein by SEQ ID NO:34 and SEQ ID NO:35); C3 (encoding complement component 3 represented herein by SEQ ID NO:30 and SEQ ID NO:31); Cfh (comprising a 45 nucleic acid sequence represented herein by SEQ ID NO:32 and SEQ ID NO:33 and encoding complement component factor h); Col9a3 (comprising a nucleic acid sequence represented herein by SEQ ID NO:36 and SEQ ID NO:37 and encoding procollagen, type IX, alpha 3); Grem1 (encoding 50 cysteine knot superfamily 1, BMP antagonist 1, represented herein by SEQ ID NO:38 and SEQ ID NO:39); Lox13 (encoding lysyl oxidase-like 3, represented herein by SEQ ID NO:40 and SEQ ID NO:41); MAGP-2 (comprising a nucleic acid sequence represented herein by SEQ ID NO:123 and 55 SEQ ID NO:124 and encoding microfibril-associated glycoprotein-2, represented herein by SEQ ID NO:42 and SEQ ID NO:43); Mglap (encoding matrix gamma-carboxyglutamate (gla) protein represented herein by SEQ ID NO:44 and SEQ ID NO:45); Naga (encoding N-acetyl galactosaminidase, 60 alpha, represented herein by SEQ ID NO:46 and SEQ ID NO:47); Nbl1 (encoding neuroblastoma, suppression of tumorigenicity 1, represented herein by SEQ ID NO:48 and SEQ ID NO:49); Ngfb (encoding nerve growth factor, beta, represented herein by SEQ ID NO:50 and SEQ ID NO:51), Npnt 65 (represented herein by SEQ ID NO:52 and SEQ ID NO:53 and encoding nephronectin); Olfm1 (encoding olfactomedin

30

1, represented herein by SEQ ID NO:54 and SEQ ID NO:55); and/or U90926 (encoding a protein represented herein by SEQ ID NO:56).

In yet another embodiment, the biomarker is a gene, or the protein encoded by the gene, selected from any of the genes or proteins specifically identified by a sequence described herein.

Typically, compounds that regulate the expression or activity of the gene or protein in the presence of the compound in the manner that has been associated by the present inventors with angiogenesis can be selected as pro-angiogenic agents or anti-angiogenesis targets (agents that are targets for inhibition in order to inhibit angiogenesis), and compounds that regulate the expression or activity of the gene or protein in the presence of the compound in a manner that is opposite or contrary to the manner that has been associated by the present inventors with angiogenesis, can be selected as anti-angiogenic agents. The method can include a further step of detecting whether a compound selected in (d) has or regulates pro-angiogenic activity or anti-angiogenic activity, such as in a bioassay for angiogenesis described herein.

Detection of the regulation of the expression of a gene (or the protein encoded thereby) in the "manner" associated with the established level of expression for that gene during angiogenesis, at a minimum, refers to the detection of the regulation of a gene that has now been shown by the present inventors to be selectively regulated in during angiogenesis, in the same direction (i.e., upregulation or downregulation) and at a similar or comparable level, as compared to a normal control (the level of expression of the gene that has been or is established under normal, or non-angiogenic conditions). In other words, if "gene X" is upregulated during angiogenesis as compared to a normal control level of expression, then one determines whether the expression of gene X is upregulated in as compared to a normal control, or whether the expression of gene X is more similar to the level of expression of the normal control. In one aspect of the invention, a gene identified as being upregulated or downregulated as compared to a baseline control according to the invention is regulated in the same direction and to at least about 10%, and more preferably at least 20%, and more preferably at least 25%, and more preferably at least 30%, and more preferably at least 35%, and more preferably at least 40%, and more preferably at least 45%, and more preferably at least 50%, and preferably at least 55%, and more preferably at least 60%, and more preferably at least 65%, and more preferably at least 70%, and more preferably at least 75%, and more preferably at least 80%, and more preferably at least 85%, and more preferably at least 90%, and more preferably at least 95%, or even higher (e.g., above 100%) of the level of expression of the gene that has been established during angiogenesis. Statistical significance should be at least p<0.05, and more preferably, at least p<0.01, and more preferably, p<0.005, and even more preferably, p<0.001.

Steps (a) and (c) of the method of the present invention require detection of the biomarker (gene or protein encoded thereby) expression and/or biological activity in a cell or in a sample derived from the cell, such as a cellular extract or supernate. Detection of biomarker expression and/or biological activity can include, but is not limited to: detecting biomarker mRNA transcription (e.g., by polymerase chain reaction (PCR), reverse transcriptase-PCR (RT-PCR), in situ hybridization, Northern blot, sequence analysis or detection of a reporter gene); detecting biomarker translation (e.g., by immunoblot, enzyme-linked immunosorbant assay (ELISA), radioimmunoassay (RIA), immunoprecipitation, immunohistochemistry and immunofluorescence); and/or detecting

biomarker biological activity (e.g., by detecting any of the activities of the particular biomarker, such as enzyme activity, receptor binding, induction of a growth factor, a cell signal transduction event, etc.). The step of detection in step (a) is the control level of biomarker expression or biological activity 5 for a cell to which the detection in step (c) is to be compared and evaluated. The step of detection in step (c) is the experimental level of biomarker expression or biological activity which indicates whether the test compound can change the level of biomarker expression or biological activity in the cell, 10 as compared to the level determined in step (a). In other words, the assay determines whether a given compound is capable of regulating the expression or activity of the biomarker (up or down), and therefore can predicted to regulate angiogenesis.

One can use a tumor cell or a normal, non-tumor cell, such as an endothelial cell, or a sample derived therefrom, in this assay, in order to identify compounds that regulate biomarker-associated angiogenesis, including angiogenesis that is associated with tumor cells, or to identify compounds in order 20 to screen for putative carcinogens.

A cell suitable for use in the present method is any cell which expresses or can be induced to express, a detectable level of the biomarker of interest. A detectable level of biomarker is a level which can be detected using any of the meth- 25 ods for biomarker detection described herein. Since the biomarkers identified herein are expressed by many mammalian cell types, a variety of cell types could be selected. However, it will be appreciated by those of skill in the art that some cell types are more suitable for use in an in vitro assay (e.g., easy 30 to maintain in culture, easy to obtain), and that certain biomarkers may be more readily detectable in some cell types, and therefore, such cell types are preferable for use in the present invention. A preferred cell type to use in the method of the present invention is any cell type that has a high expression or 35 low expression of the biomarker in a tumor cell as compared to a non-tumor cell of the same cell type, or has a high expression or low expression of the biomarker under angiogenic conditions as compared to non-angiogenic conditions, so that a change in biomarker expression or activity is readily 40 detectable. As discussed above, one can also use a sample derived from such a cell, such as a cell extract or cell supemate. Some preferred cells to use in the method of the present invention include, but are not limited to: fibroblasts (and fibrosarcomas), epithelial cells, endothelial cells, and breast, 45 colon, kidney, ovarian or uterine tumor cells and non-tumor cells that endogenously or recombinantly express the biomarker. In one embodiment, a cell suitable for use in any aspect the general assay method is a cell which has been transfected with a recombinant nucleic acid molecule encoding the biom- 50 arker and operatively linked to a transcription control sequence so that the biomarker is expressed by the cell. Methods and reagents for preparing recombinant cells are known in the art.

As used herein, the term "putative regulatory compound" refers to compounds having an unknown or previously unappreciated regulatory activity in a particular process. The above-described method for identifying a compound of the present invention includes a step of contacting a test cell with a compound being tested for its ability to regulate the expression or biological activity of the biomarker. For example, test cells can be grown in liquid culture medium or grown on solid medium in which the liquid medium or the solid medium contains the compound to be tested. In addition, as described above, the liquid or solid medium contains components necessary for cell growth, such as assimilable carbon, nitrogen and micronutrients.

32

The above-described methods, in one aspect, involve contacting cells with the compound being tested for a sufficient time to allow for interaction of the putative regulatory compound with an element that affects biomarker expression and/or biological activity in a cell. Such elements can include, but are not limited to: a nucleic acid molecule encoding the biomarker (including regulatory regions of such a molecule). the biomarker protein, biomarker inhibitors, biomarker stimulators, and biomarker substrates. The period of contact with the compound being tested can be varied depending on the result being measured, and can be determined by one of skill in the art. For example, for binding assays, a shorter time of contact with the compound being tested is typically suitable, than when activity or expression is assessed. As used herein, the term "contact period" refers to the time period during which cells are in contact with the compound being tested. The term "incubation period" refers to the entire time during which cells are allowed to grow prior to evaluation, and can be inclusive of the contact period. Thus, the incubation period includes all of the contact period and may include a further time period during which the compound being tested is not present but during which growth is continuing (in the case of a cell based assay) prior to scoring. The incubation time for growth of cells can vary but is sufficient to allow for the upregulation or downregulation of biomarker expression or biological activity in a cell. It will be recognized that shorter incubation times are preferable because compounds can be more rapidly screened. A preferred incubation time is between about 1 hour to about 48 hours.

The conditions under which the cell or cell lysate of the present invention is contacted with a putative regulatory compound, such as by mixing, are any suitable culture or assay conditions and includes an effective medium in which the cell can be cultured or in which the cell lysate can be evaluated in the presence and absence of a putative regulatory compound. Cells of the present invention can be cultured in a variety of containers including, but not limited to, tissue culture flasks, test tubes, microtiter dishes, and petri plates. Culturing is carried out at a temperature, pH and carbon dioxide content appropriate for the cell. Such culturing conditions are also within the skill in the art. Cells are contacted with a putative regulatory compound under conditions which take into account the number of cells per container contacted, the concentration of putative regulatory compound(s) administered to a cell, the incubation time of the putative regulatory compound with the cell, and the concentration of compound administered to a cell. Determination of effective protocols can be accomplished by those skilled in the art based on variables such as the size of the container, the volume of liquid in the container, conditions known to be suitable for the culture of the particular cell type used in the assay, and the chemical composition of the putative regulatory compound (i.e., size, charge etc.) being tested. A preferred amount of putative regulatory compound(s) comprises between about 1 nM to about 10 mM of putative regulatory compound(s) per well of a 96-well plate.

In one aspect, the present method also makes use of noncell based assay systems to identify compounds that can regulate biomarker expression or biological activity and thereby are predicted to be useful for regulating cell growth. For example, biomarker proteins and nucleic acid molecules encoding the biomarker may be recombinantly expressed and utilized in non-cell based assays to identify compounds that bind to the protein or nucleic acid molecule, respectively. In non-cell based assays the recombinantly expressed biomarker or nucleic acid encoding the biomarker is attached to a

solid substrate such as a test tube, microtiter well or a column, by means well known to those in the art.

In one embodiment, DNA encoding a reporter molecule can be linked to a regulatory element of the biomarker gene (or a gene encoding a protein that directly regulates the biomarker) and used in appropriate intact cells, cell extracts or lysates to identify compounds that modulate biomarker gene expression, respectively. Appropriate cells or cell extracts are prepared from any cell type that normally expresses the biomarker, thereby ensuring that the cell extracts contain the transcription factors required for in vitro or in vivo transcription. The screen can be used to identify compounds that modulate the expression of the reporter construct. In such screens, the level of reporter gene expression is determined in the presence of the test compound and compared to the level of 15 expression in the absence of the test compound.

Following steps (a), (b) and (c) of the method to identify a compound that regulates the biomarker is a step (d) of selecting a compound that regulates (up or down) the level of the biomarker expression or activity in the cell, as compared to in the absence of the compound. Compounds which cause a regulation (increase or decrease) in the level of biomarker expression or biological activity are selected by the present method as being compounds that are predicted to be useful as pro-angiogenesis agents or anti-angiogenesis agents (or targets for regulation of angiogenesis), depending on how the biomarker has been correlated with angiogenesis according to the description provided herein.

Preferably, compounds which are selected in step (d) are compounds for which, after the test cell was contacted with 30 the compound in step (b), the level of biomarker expression or biological activity detected in step (c) was statistically significantly changed (i.e., with at least a 95% confidence level, or p<0.05) as compared to the initial level of biomarker expression or biological activity detected in step (a). Prefer- 35 ably, detection of at least about a 30% change in biomarker expression or biological activity in the cell as compared to initial level results in selection of the compound according to step (d). More preferably, detection of at least about a 50% change and more preferably at least about a 70% change, and 40 more preferably at least about a 90% change, or any percentage change between 5% and higher in 1% increments (i.e., 5%, 6%, 7%, 8% . . .) in biomarker expression or biological activity in the cell as compared to the initial level results in selection of the compound according to step (d). In one 45 embodiment, a 1.5 fold change in biomarker expression or biological activity in the cell as compared to the initial level results in selection of the compound according to step (d). More preferably, detection of at least about a 3 fold change, and more preferably at least about a 6 fold change, and even 50 more preferably, at least about a 12 fold change, and even more preferably, at least about a 24 fold change, or any fold change from 1.5 up in increments of 0.5 fold (i.e., 1.5, 2.0, 2.5, 3.0 . . .) in biomarker expression or biological activity as compared to the initial level, results in selection of the com- 55 pound according to step (d).

It is to be understood that either of steps (a) and (c) of detection in any of the methods to identify a compound described above can result in no detection, or no change in detection, of biomarker expression or biological activity. In 60 addition, since the level of biomarker expression or biological activity in step (a) (i.e., the initial level) is one of the control levels of biomarker for the assay (i.e., in the absence of the test compound), if step (a) reveals no detectable biomarker expression or biological activity, then any detectable level of 65 biomarker expression or biological activity in step (c) is considered to be a positive result and indicative of increased

34

biomarker activity in the cell and the appropriate assessment associated with this result. If the initial level of biomarker expression or biological activity in step (a) is a detectable level, then the level of biomarker expression or biological activity detected in step (c) is evaluated to determine whether it is statistically significantly greater than or less than that of step (a). It is possible that the level of biomarker expression or biological activity in step (c) could be no detectable change, which would indicate that the compound did not increase or decrease biomarker activity. In this scenario, however, it should be determined that the test cell can display an increase or decrease in the particular biomarker expression or biological activity under some conditions (i.e., by contact with a compound known to increase the biomarker activity in the test cell), so that false negatives are not identified.

In one embodiment of this method to identify regulators of biomarkers of the present invention, the method further includes the step of detecting whether the compound selected in step (d) can inhibit tumor cell formation or a characteristic thereof. In this embodiment, the test cell is contacted with the compound as in step (b), and the growth characteristics of the cell before and after contact with the cell are evaluated. Evaluation of cell growth can be by any suitable method in the art, including, but not limited to, proliferation assays (e.g., by measuring uptake of [³H]-thymidine, viewing cells morphologically) and/or evaluating markers of cell growth (e.g., measurement of changes in cell surface markers, measurement of intracellular indicators of cell growth). Such methods are known in the art and are exemplified in the attached examples.

Compounds suitable for testing and use in the methods of the present invention include any known or available proteins, nucleic acid molecules, as well as products of drug design, including peptides, oligonucleotides, carbohydrates and/or synthetic organic molecules. Such an agent can be obtained, for example, from molecular diversity strategies (a combination of related strategies allowing the rapid construction of large, chemically diverse molecule libraries), libraries of natural or synthetic compounds, in particular from chemical or combinatorial libraries (i.e., libraries of compounds that differ in sequence or size but that have the same building blocks) or by rational drug design. See for example, Maulik et al., 1997, supra. Candidate compounds initially identified by drug design methods can be screened for the ability to modulate the expression and/or biological activity of the biomarker using the methods described herein.

Compounds identified by the method described above can be used in a method to regulate angiogenesis, treat a condition or reduce a symptom of a condition in which inhibition of angiogenesis is desirable (e.g.,cancer), or treat a condition or reduce a symptom of a condition in which promotion of angiogenesis is desirable (e.g.,ischemia, stroke), as described herein and any such compounds are encompassed for use in the method described below.

More particularly, according to one embodiment of the present invention, administration of a compound or composition of the invention or targeting of a biomarker of the invention is useful to inhibit the tumorigenicity of a target cell or to inhibit angiogenesis in a tissue of a patient. Typically, it is desirable to inhibit the growth of a target cell (e.g., a tumor) to obtain a therapeutic benefit in the patient. In one embodiment, patients whom are suitable candidates for methods of the present invention include, but are not limited to, patients that have, or are at risk of developing (e.g., are predisposed to), cancer or a lymphoproliferative disease, or any condition in which regulation of angiogenesis might be beneficial. Particular conditions that are characterized or caused by abnor-

mal or excessive angiogenesis, and therefore may be treated using the methods and compositions of the invention include, but are not limited to: cancer (e.g., activation of oncogenes, loss of tumor suppressors); infectious diseases (e.g., pathogens express angiogenic genes, enhance angiogenic pro- 5 grams); autoimmune disorders (e.g., activation of mast cells and other leukocytes); vascular malformations (e.g., Tie-2 mutation); DiGeorge syndrome (e.g., low VEGF and neuropilin-1 expression); HHT (e.g., mutations of endoglin or LK-1), cavernous hemangioma (e.g., loss of Cx37 and Cx40); 10 atherosclerosis; transplant ateriopathy; obesity (e.g., angiogenesis induced by fatty diet, weight loss by angiogenesis inhibitors); psoriasis; warts; allergic dermatitis; scar keloids; pyogenic granulomas; blistering disease; Kaposi sarcoma in AIDS patients; persistent hyperplastic vitreous syndrome 15 (e.g., loss of Ang-2 or VEGF164); diabetic retinopathy; retinopathy of prematurity; choroidal neovascularization (e.g., TIMP-3 mutation); primary pulmonary hypertension (e.g., germline BMPR-2 mutation, somatic EC mutation); asthma; nasal polyps; inflammatory bowel disease; periodontal dis- 20 ease; ascites; peritoneal adhesions; endometriosis; uterine bleeding; ovarian cysts; ovarian hyperstimulation; arthritis; synovitis; osteomyelitis; and osteophyte formation.

In another embodiment of the invention, administration of a compound or composition of the invention or targeting of a 25 biomarker of the invention is useful to promote angiogenesis. Patients whom are suitable candidates for such a method of the invention include, but are not limited to: patients with vascular deficiencies, cardiovascular disease, or patients in whom stimulation of endothelial cell activation and stabilization of newly formed microvessels or other vessels would be beneficial. For example, such conditions include, but are not limited to, stroke, ischemia and related conditions.

Therefore, yet another embodiment of the invention relates to methods to increase or decrease the expression or biologi- 35 cal activity of any one or more of the biomarkers described herein (e.g., Table I, Table IV, Table V, and/or Table VI) in cells (e.g., isolated cells, cells of a tissue, cells in a patient) in order to achieve a goal. This goal can include, but is not limited to, reduction of angiogenesis in a tissue, decreased 40 tumorigenicity of tumor cells, or reduction in the potential for development of tumor cells, enhancement or promotion of angiogenesis in a tissue, or treatment of a disease or condition in which enhanced angiogenesis would be desirable. Such methods generally include the step of increasing or decreas- 45 ing the expression and/or biological activity of one or more biomarkers described herein, as required for a given cell type, in order to achieve the desired result (e.g., inhibition or promotion of angiogenesis, cancer inhibition, etc.). In one embodiment, the biomarker is a protein, or the gene encoding 50 such protein, selected from: ADAMts7, CRELD-2, Decorin, ECM1, Inhibin β -b, Integrin α -3, Integrin α -6, Lipocalin-7, Lox1-3, Lumican, MAGP-2, Matrilin-2, Nephronectin, SerpinE2, and/or SMOC-2.

In another embodiment, the biomarker is a gene, or the 55 protein encoded by the gene, selected from: 0610007C21Rik, apoptosis related protein APR-3, 1810014L12Rik, Cd14 (encoding CD14 antigen represented herein by SEQ ID NO:5 and SEQ ID NO:6), Cd38 (comprising a nucleic acid sequence represented herein by SEQ ID NO:7 and encoding 60 CD38 antigen); Cd53 (encoding CD53 antigen represented herein by SEQ ID NO:8 and SEQ ID NO:9), Emp2 (encoding epithelial membrane protein represented herein by SEQ ID NO:10 and SEQ ID NO:11), Fcgrt (encoding Fc receptor (IgG, alpha chain transporter) represented herein by SEQ ID NO:12 and SEQ ID NO:13), IsIr (encoding immunoglobulin superfamily containing leucine-rich repeat represented

36

herein by SEQ ID NO:14 and SEQ ID NO:15); Lrp2 (comprising a nucleic acid sequence represented herein by SEQ ID NO:16 and SEQ ID NO:17 and encoding low density lipoprotein receptor-related protein 2); Ly6a (encoding lymphocyte antigen 6 complex, locus A represented herein by SEQ ID NO:18); P2rx4 (encoding purinergic receptor P2X, ligand-gated ion channel 4, represented herein by SEQ ID NO:19 and SEQ ID NO:20; Pcdhb9 (encoding protocadherin beta 9 represented herein by SEQ ID NO:21 and SEQ ID NO:22); Ptpre (encoding protein tyrosine phosphatase receptor type E represented herein by SEQ ID NO:23 and SEQ ID NO:24); Slc4a3 (encoding solute carrier family 4 (anion exchanger) member 3, represented herein by SEQ ID NO:25 and SEQ ID NO:26); and/or Tmc6 (encoding transmembrane channel-like gene family 6, represented herein by SEQ ID NO:27).

In yet another embodiment, the biomarker is a gene, or the protein encoded by the gene, selected from: 9130213B05Rik (encoding a protein represented herein by SEQ ID NO:29); C1s (encoding complement component 1, s subcomponent, represented herein by SEQ ID NO:34 and SEQ ID NO:35); C3 (encoding complement component 3 represented herein by SEQ ID NO:30 and SEQ ID NO:31); Cfh (comprising a nucleic acid sequence represented herein by SEQ ID NO:32 and SEQ ID NO:33 and encoding complement component factor h); Col9a3 (comprising a nucleic acid sequence represented herein by SEQ ID NO:36 and SEQ ID NO:37 and encoding procollagen, type IX, alpha 3); Grem1 (encoding cysteine knot superfamily 1, BMP antagonist 1, represented herein by SEQ ID NO:38 and SEQ ID NO:39); Lox13 (encoding lysyl oxidase-like 3, represented herein by SEQ ID NO:40 and SEQ ID NO:41); MAGP-2 (comprising a nucleic acid sequence represented herein by SEQ ID NO:123 and SEQ ID NO:124 and encoding microfibrillar associated protein 5, represented herein by SEQ ID NO:42 and SEQ ID NO:43); Mglap (encoding matrix gamma-carboxyglutamate (gla) protein represented herein by SEQ ID NO:44 and SEQ ID NO:45); Naga (encoding N-acetyl galactosaminidase, alpha, represented herein by SEQ ID NO:46 and SEQ ID NO:47); Nb11 (encoding neuroblastoma, suppression of tumorigenicity 1, represented herein by SEQ ID NO:48 and SEQ ID NO:49); Ngfb (encoding nerve growth factor, beta, represented herein by SEQ ID NO:50 and SEQ ID NO:51), Npnt (represented herein by SEQ ID NO:52 and SEQ ID NO:53 and encoding nephronectin); Olfm1 (encoding olfactomedin 1, represented herein by SEQ ID NO:54 and SEQ ID NO:55); and/or U90926 (encoding a protein represented herein by SEQ ID NO:56).

In yet another embodiment, the biomarker is a gene, or the protein encoded by the gene, selected from any of the genes or proteins specifically identified by a sequence described herein.

In the method of the present invention wherein the goals are to reduce angiogenesis in a tissue, decrease tumorigenicity of tumor cells, decrease tumor burden, increase survival, or reduce the potential for the development of tumor cells, preferably, cells that are targeted by the method are cells which, prior to the application of the present method, are exhibiting inappropriate (malignant) cell growth or a potential therefore, or cells in a tissue where it is desirable to inhibit angiogenesis. Preferred cells to regulate according to this aspect of the present invention include tumor cells. Cells in which it is desirable to inhibit tumorigenicity or tissues in which inhibition of angiogenesis is desired can be identified, for example, using the method for assessing the presence of cancer cells or biomarker expression and activity of the present invention as described in detail above. Such methods are particularly use-

ful in patients where increased tumorigenicity (or simply tumor growth) or angiogenesis is, or is predicted to become, problematic. Therefore, such a method is particularly useful to treat patients that have, or are at a risk of developing, tumor cells (i.e., a cancer), or to treat any other patients having a condition characterized by undesirable cell growth (e.g., lymphoproliferative disorders). Other diseases and conditions in which inhibition of tumorigenicity or angiogenesis would be desirable will be apparent to those of skill in the art (many are discussed below) and are intended to be encompassed by the 10 present invention.

Similarly, in the method of the present invention wherein the goals are to enhance or promote angiogenesis in a tissue, preferably, cells that are targeted by the method are cells in a tissue where it is desirable to promote angiogenesis. Preferred 15 cells to regulate according to this aspect of the present invention include vascular endothelial cells. Such methods are particularly useful in patients where increased angiogenesis may be useful, such as in patients that have a vascular insufficiency or where the promotion of vascular stabilization and 20 development is desired. Therefore, such a method is particularly useful to treat patients with vascular deficiencies, cardiovascular disease, or to stimulate endothelial cell activation and stabilization of newly formed microvessels or other vessels. Conditions in which promotion of angiogenesis would 25 be desirable will be apparent to those of skill in the art and are intended to be encompassed by the present invention.

Accordingly, the method of the present invention includes a step of modulating (i.e., upregulating or downregulating) biomarker expression and/or biological activity in a patient 30 that has, or is at risk of developing, inappropriate or unregulated cell growth (e.g., tumors) or angiogenesis, or a patient or subject that is in need of promotion of angiogenesis, depending on the goal of the therapy, as discussed above. Modulating biomarker expression or biological activity according to the 35 present invention can be accomplished by directly affecting biomarker expression (transcription or translation) or biological activity, or by directly affecting the ability of a regulator (inhibitor or stimulator) of the biomarker to bind to the biomarker or to activate the biomarker. Preferably, the 40 method of the present invention is targeted to a particular type of cell or tissue or region of the body in which inhibition of cell growth or regulation of angiogenesis is desired. A targeted cell, for example, could include a tumor cell, wherein the method does not substantially affect biomarker expres- 45 sion or biological activity in non-tumor cells, or in cells of a different type that the tumor cell type. Therefore, the method of the present invention, in one embodiment, is intended to be specifically targeted to biomarker expression and/or biological activity for the purpose of inhibiting or promoting cell 50 growth, or inhibiting or promoting angiogenesis by modulating biomarker expression and/or biological activity.

An increase in biomarker expression and/or biological activity is defined herein as any measurable (detectable) increase (i.e., upregulation, stimulation, enhancement) of the 55 expression or activity of the biomarker. As used herein, to increase biomarker expression and/or biological activity refers to any measurable increase in biomarker expression and/or biological activity by any suitable method of measurement. A decrease in biomarker expression and/or biological activity is defined herein as any measurable (detectable) decrease (i.e., downregulation, inhibition, reduction) of the expression or activity of biomarker. As used herein, to decrease biomarker expression and/or biological activity refers to any measurable decrease in the biomarker expression and/or biological activity by any suitable method of measurement.

38

Accordingly, one embodiment of the present invention includes the use of a variety of agents (i.e., regulatory compounds) which, by acting directly on the biomarker (or by being the biomarker gene encoding a protein or the biomarker protein itself) or by acting on inhibitors or stimulators of the biomarker or being an inhibitor or stimulator of the biomarker, modulate (regulate up or down) the expression and/or biological activity of the biomarker in a cell to produce a desired effect (e.g., inhibition of tumorigenesis or reduction of tumor burden or tumor stasis/increase of survival, inhibition or promotion of angiogenesis). Agents useful in the present invention include, for example, proteins, nucleic acid molecules, antibodies, and compounds that are products of rational drug design (i.e., drugs). Such compounds can be identified using the method of identifying compounds for regulating tumor cell growth and malignancy or for regulating angiogenesis as described above. Moreover, the expression or biological activity of the biomarker in a cell can be determined using the methods described above.

Therefore, in one embodiment, the method of the present invention increases the transcription and/or the translation of the biomarker by a cell that naturally expresses the biomarker and that is the target for growth regulation, or increases (stimulates, enhances) the biological activity of the biomarker. Methods for increasing the expression of a given biomarker include, but are not limited to, administering an agent that increases the expression or biological activity of the endogenous biomarker, administering biomarker protein or a homologue or analog (agonist) thereof to a subject, and/or overexpressing biomarker in target cells. In one aspect of this embodiment, the biomarker can be effectively overexpressed in a cell by increasing the activity of a promoter for the biomarker gene in the cell such that expression of endogenous biomarker in the cell is increased. For example, the activity of the biomarker gene promoter can be increased by methods which include, contacting the promoter with a transcriptional activator, inhibiting a biomarker promoter inhibitor, and increasing the activity of a biomarker promoter stimulator. Methods by which such compounds (e.g., transcriptional activators) can be administered to a cell are described below. In another embodiment, biomarker activity is increased by administering the biomarker or a homologue or analog (synthetic homologue or mimetic or compound) to the target cells or to the patient in an appropriate carrier or delivery vehicle.

In another embodiment, the method of the present invention decreases the transcription and/or the translation of the biomarker by a cell that naturally expresses the biomarker and that is the target for growth regulation, or inhibits the biological activity of biomarker. In this embodiment, it is desired to modify a target cell in order to decrease in biomarker gene expression, decrease the function of the gene, or decrease the function of the gene product (i.e., the protein encoded by the gene). Such methods can be referred to as inactivation (complete or partial), deletion, interruption, blockage or downregulation of a gene encoding the biomarker. In one embodiment, reduction in biomarker activity or expression is achieved by use of a biomarker antagonist, antagonists having been described above.

In one aspect of this embodiment of the present invention, the expression and/or biological activity of the biomarker is increased by overexpressing the biomarker in the cell in which angiogenesis is to be regulated. Overexpression of a biomarker refers to an increase in expression of the biomarker over a normal, endogenous level of biomarker expression. For some cell types, which do not express detectable levels of the biomarker under normal conditions, such expression can be any detectable level. For cell types which do express detect-

able levels of the biomarker under normal conditions, an overexpression is any statistically significant increase in expression of the biomarker (p<0.05) (or constitutive expression where expression is normally not constitutive) over endogenous levels of expression. One method by which 5 biomarker overexpression can be achieved is by transfecting the cell with a recombinant nucleic acid molecule encoding the biomarker operatively linked to a transcription control sequence, wherein the recombinant biomarker is expressed by the cell. As discussed previously herein, the nucleic acid 10 sequence encoding biomarker, vectors suitable for expressing such a molecule, and methods of transfection of a cell with such a molecule, including in vivo methods, are known and are described in detail below.

A recombinant nucleic acid molecule expressing the biom- 15 arker is a molecule that can include at least one of any nucleic acid sequence encoding a protein having the biomarker biological activity operatively linked to at least one of any transcription control sequence capable of effectively regulating expression of the nucleic acid molecule(s) in the cell to be 20 transfected. Although the phrase "nucleic acid molecule" primarily refers to the physical nucleic acid molecule and the phrase "nucleic acid sequence" primarily refers to the sequence of nucleotides on the nucleic acid molecule, the two phrases can be used interchangeably, especially with respect 25 to a nucleic acid molecule, or a nucleic acid sequence, being capable of encoding a protein. In addition, the phrase "recombinant molecule" primarily refers to a nucleic acid molecule operatively linked to a transcription control sequence, but can be used interchangeably with the phrase "nucleic acid mol-30" ecule" which is administered to an animal.

Preferably, a recombinant nucleic acid molecule is produced using recombinant DNA technology (e.g., polymerase chain reaction (PCR) amplification, cloning). Suitable nucleic acid sequences encoding the biomarker for use in a 35 recombinant nucleic acid molecule of the present invention include any nucleic acid sequence that encodes the biomarker protein having biological activity and suitable for use in the target host cell. For example, when the target host cell is a human cell, human biomarker-encoding nucleic acid 40 sequences are preferably used, although the present invention is not limited to strict use of naturally occurring sequences or same-species sequences.

A recombinant nucleic acid molecule includes a recombinant vector, which is any nucleic acid sequence, typically a 45 heterologous sequence, which is operatively linked to the isolated nucleic acid molecule encoding a biomarker protein, which is capable of enabling recombinant production of the biomarker protein, and which is capable of delivering the nucleic acid molecule into a host cell according to the present 50 invention. Such a vector can contain nucleic acid sequences that are not naturally found adjacent to the isolated nucleic acid molecules to be inserted into the vector. The vector can be either RNA or DNA, either prokaryotic or eukaryotic, and Recombinant vectors can be used in the cloning, sequencing, and/or otherwise manipulating of nucleic acid molecules. Recombinant vectors are preferably used in the expression of nucleic acid molecules, and can also be referred to as expression vectors. Preferred recombinant vectors are capable of 60 being expressed in a transfected host cell, and particularly, in a transfected mammalian host cell in vivo.

In a recombinant molecule of the present invention, nucleic acid molecules are operatively linked to expression vectors containing regulatory sequences such as transcription control 65 sequences, translation control sequences, origins of replication, and other regulatory sequences that are compatible with

40

the host cell and that control the expression of nucleic acid molecules of the present invention. In particular, recombinant molecules of the present invention include nucleic acid molecules that are operatively linked to one or more transcription control sequences. The phrase "operatively linked" refers to linking a nucleic acid molecule to a transcription control sequence in a manner such that the molecule is expressed when transfected (i.e., transformed, transduced or transfected) into a host cell.

Transcription control sequences are sequences that control the initiation, elongation, and termination of transcription. Particularly important transcription control sequences are those that control transcription initiation, such as promoter, enhancer, operator and repressor sequences. Suitable transcription control sequences include any transcription control sequence that can function in a host cell according to the present invention. A variety of suitable transcription control sequences are known to those skilled in the art. Preferred transcription control sequences include those which function in mammalian cells, with cell- or tissue-specific transcription control sequences being particularly preferred. Examples of preferred transcription control sequences include, but are not limited to, transcription control sequences useful for expression of a protein in epithelial cells and tumor cells and the naturally occurring biomarker promoter. Particularly preferred transcription control sequences include inducible promoters, cell-specific promoters, tissue-specific promoters (e.g., insulin promoters) and enhancers. Suitable promoters for these and other cell types will be easily determined by those of skill in the art. Transcription control sequences of the present invention can also include naturally occurring transcription control sequences naturally associated with the protein to be expressed prior to isolation. In one embodiment, a transcription control sequence includes an inducible promoter.

One type of recombinant vector useful in a recombinant nucleic acid molecule of the present invention is a recombinant viral vector. Such a vector includes a recombinant nucleic acid sequence encoding a biomarker protein of the present invention that is packaged in a viral coat that can be expressed in a host cell in an animal or ex vivo after administration. A number of recombinant viral vectors can be used, including, but not limited to, those based on alphaviruses, poxviruses, adenoviruses, herpesviruses, lentiviruses, adenoassociated viruses and retroviruses. Particularly preferred viral vectors are those based on adenoviruses and adenoassociated viruses. Viral vectors suitable for gene delivery are well known in the art and can be selected by the skilled artisan for use in the present invention. A detailed discussion of current viral vectors is provided in "Molecular Biotechnology," Second Edition, by Glick and Pasternak, ASM Press, Washington D.C., 1998, pp. 555-590, the entirety of which is incorporated herein by reference.

For example, a retroviral vector, which is useful when it is preferably in the present invention, is a virus or a plasmid. 55 desired to have a nucleic acid sequence inserted into the host genome for long term expression, can be packaged in the envelope protein of another virus so that it has the binding specificity and infection spectrum that are determined by the envelope protein (e.g., a pseudotyped virus). In addition, the envelope gene can be genetically engineered to include a DNA element that encodes and amino acid sequence that binds to a cell receptor to create a recombinant retrovirus that infects a specific cell type. Expression of the biomarker gene can be further controlled by the use of a cell or tissue-specific promoter. Retroviral vectors have been successfully used to transfect cells with a gene which is expressed and maintained in a variety of ex vivo systems

An adenoviral vector is a preferred vector for use in the present method. An adenoviral vector infects a wide range of human cells and has been used extensively in live vaccines. Adenoviral vectors used in gene therapy do not integrate into the host genome, and therefore, gene therapy using this system requires periodic administration, although methods have been described which extend the expression time of adenoviral transferred genes, such as administration of antibodies directed against T cell receptors at the site of expression (Sawchuk et al., 1996, Hum. Gene. Ther. 7:499-506). The 10 efficiency of adenovirus-mediated gene delivery can be enhanced by developing a virus that preferentially infects a particular target cell. For example, a gene for the attachment fibers of adenovirus can be engineered to include a DNA element that encodes a protein domain that binds to a cell- 15 specific receptor. Examples of successful in vivo delivery of genes has been demonstrated and is discussed in more detail

Yet another type of viral vector is based on adeno-associated viruses, which are small, nonpathogenic, single-stranded 20 human viruses. This virus can integrate into a specific site on chromosome 19. This virus can carry a cloned insert of about 4.5 kb, and has typically been successfully used to express proteins in vivo from 70 days to at least 5 months. Demonstrating that the art is quickly advancing in the area of gene 25 therapy, however, a publication by Bennett et al. reported efficient and stable transgene expression by adeno-associated viral vector transfer in vivo for greater than 1 year (Bennett et al., 1999, *Proc. Natl. Acad. Sci. USA* 96:9920-9925).

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Another type of viral vector that is suitable for use in the 30 present invention is a herpes simplex virus vector. Herpes simplex virus type 1 infects and persists within nondividing neuronal cells, and is therefore a suitable vector for targeting and transfecting cells of the central and peripheral nervous system with a biomarker protein of the present invention. 35 Preclinical trials in experimental animal models with such a vector has demonstrated that the vector can deliver genes to cells of both the brain and peripheral nervous system that are expressed and maintained for long periods of time.

Suitable host cells to transfect with a recombinant nucleic 40 acid molecule according to the present invention include any mammalian cell that can be transfected. Host cells can be either untransfected cells or cells that are already transfected with at least one nucleic acid molecule. Host cells according to the present invention can be any cell capable of producing 45 a biomarker protein as described herein or in which it is desired to produce the biomarker.

According to the present invention, a host cell can also be referred to as a target cell or a targeted cell in vivo, in which a recombinant nucleic acid molecule encoding a biomarker 50 protein having the biological activity of the biomarker is to be expressed. As used herein, the term "target cell" or "targeted cell" refers to a cell to which a recombinant nucleic acid molecule of the present invention is selectively designed to be delivered. The term target cell does not necessarily restrict the 55 delivery of a recombinant nucleic acid molecule only to the target cell and no other cell, but indicates that the delivery of the recombinant molecule, the expression of the recombinant molecule, or both, are specifically directed to a preselected host cell. Targeting delivery vehicles, including liposomes 60 and viral vector systems are known in the art. For example, a liposome can be directed to a particular target cell or tissue by using a targeting agent, such as an antibody, soluble receptor or ligand, incorporated with the liposome, to target a particular cell or tissue to which the targeting molecule can bind. 65 Targeting liposomes are described, for example, in Ho et al., 1986, Biochemistry 25: 5500-6; Ho et al., 1987a, J Biol Chem

42

262: 13979-84; Ho et al., 1987b, J Biol Chem 262: 13973-8; and U.S. Pat. No. 4,957,735 to Huang et al., each of which is incorporated herein by reference in its entirety). Ways in which viral vectors can be modified to deliver a nucleic acid molecule to a target cell have been discussed above. Alternatively, the route of administration, as discussed below, can be used to target a specific cell or tissue. For example, intracoronary administration of an adenoviral vector has been shown to be effective for the delivery of a gene cardiac myocytes (Maurice et al., 1999, JClin. Invest. 104:21-29). Intravenous delivery of cholesterol-containing cationic liposomes has been shown to preferentially target pulmonary tissues (Liu et al., Nature Biotechnology 15:167, 1997), and effectively mediate transfer and expression of genes in vivo. Other examples of successful targeted in vivo delivery of nucleic acid molecules are known in the art. Finally, a recombinant nucleic acid molecule can be selectively (i.e., preferentially, substantially exclusively) expressed in a target cell by selecting a transcription control sequence, and preferably, a promoter, which is selectively induced in the target cell and remains substantially inactive in non-target cells.

According to the method of the present invention, a host cell is preferably transfected in vivo (i.e., in a mammal) as a result of administration to a mammal of a recombinant nucleic acid molecule, or ex vivo, by removing cells from a mammal and transfecting the cells with a recombinant nucleic acid molecule ex vivo. Transfection of a nucleic acid molecule into a host cell according to the present invention can be accomplished by any method by which a nucleic acid molecule administered into the cell in vivo, and includes, but is not limited to, transfection, electroporation, microinjection, lipofection, adsorption, viral infection, naked DNA injection and protoplast fusion. Methods of administration are discussed in detail below.

In one embodiment of the present invention, a recombinant nucleic acid molecule of the present invention is administered to a patient in a liposome delivery vehicle, whereby the nucleic acid sequence encoding the biomarker protein enters the host cell (i.e., the target cell) by lipofection. A liposome delivery vehicle contains the recombinant nucleic acid molecule and delivers the molecules to a suitable site in a host recipient. According to the present invention, a liposome delivery vehicle comprises a lipid composition that is capable of delivering a recombinant nucleic acid molecule of the present invention, including both plasmids and viral vectors, to a suitable cell and/or tissue in a patient. A liposome delivery vehicle of the present invention comprises a lipid composition that is capable of fusing with the plasma membrane of the target cell to deliver the recombinant nucleic acid molecule into a cell. A liposome delivery vehicle can also be used to deliver a protein, drug, or other regulatory compound to a patient.

A liposome delivery vehicle of the present invention can be modified to target a particular site in a mammal (i.e., a targeting liposome), thereby targeting and making use of a nucleic acid molecule of the present invention at that site. Suitable modifications include manipulating the chemical formula of the lipid portion of the delivery vehicle. Manipulating the chemical formula of the lipid portion of the delivery vehicle can elicit the extracellular or intracellular targeting of the delivery vehicle. For example, a chemical can be added to the lipid formula of a liposome that alters the charge of the lipid bilayer of the liposome so that the liposome fuses with particular cells having particular charge characteristics. Other targeting mechanisms include targeting a site by addition of exogenous targeting molecules (i.e., targeting agents) to a liposome (e.g., antibodies, soluble receptors or ligands).

A liposome delivery vehicle is preferably capable of remaining stable in a patient for a sufficient amount of time to deliver a nucleic acid molecule of the present invention to a preferred site in the patient (i.e., a target cell). A liposome delivery vehicle of the present invention is preferably stable 5 in the patient into which it has been administered for at least about 30 minutes, more preferably for at least about 1 hour and even more preferably for at least about 24 hours. A preferred liposome delivery vehicle of the present invention is from about 0.01 microns to about 1 microns in size.

Suitable liposomes for use with the present invention include any liposome. Preferred liposomes of the present invention include those liposomes commonly used in, for example, gene delivery methods known to those of skill in the art. Preferred liposome delivery vehicles comprise multila- 15 mellar vesicle (MLV) lipids and extruded lipids. Methods for preparation of MLV's are well known in the art. According to the present invention, "extruded lipids" are lipids which are prepared similarly to MLV lipids, but which are subsequently extruded through filters of decreasing size, as described in 20 Templeton et al., 1997, Nature Biotech., 15:647-652, which is incorporated herein by reference in its entirety. Small unilamellar vesicle (SUV) lipids can also be used in the composition and method of the present invention. In one embodiment, liposome delivery vehicles comprise liposomes having a 25 polycationic lipid composition (i.e., cationic liposomes) and/ or liposomes having a cholesterol backbone conjugated to polyethylene glycol. In a preferred embodiment, liposome delivery vehicles useful in the present invention comprise one or more lipids selected from the group of DOTMA, DOTAP, 30 DOTIM, DDAB, and cholesterol.

Preferably, the transfection efficiency of a nucleic acid: liposome complex of the present invention is at least about 1 picogram (pg) of protein expressed per milligram (mg) of total tissue protein per microgram (μg) of nucleic acid delivered. More preferably, the transfection efficiency of a nucleic acid:liposome complex of the present invention is at least about 10 pg of protein expressed per mg of total tissue protein per μg of nucleic acid delivered; and even more preferably, at least about 50 pg of protein expressed per mg of total tissue 40 protein per μg of nucleic acid delivered; and most preferably, at least about 100 pg of protein expressed per mg of total tissue protein per μg of nucleic acid delivered.

Complexing a liposome with a nucleic acid molecule of the present invention can be achieved using methods standard in 45 the art. A suitable concentration of a nucleic acid molecule of the present invention to add to a liposome includes a concentration effective for delivering a sufficient amount of recombinant nucleic acid molecule into a target cell of a patient such that the biomarker protein encoded by the nucleic acid mol- 50 ecule can be expressed in a an amount effective to inhibit the growth of the target cell or to inhibit or promote angiogenesis at a tissue site. Preferably, from about 0.1 µg to about 10 µg of nucleic acid molecule of the present invention is combined with about 8 nmol liposomes. In one embodiment, the ratio of 55 nucleic acids to lipids (µg nucleic acid:nmol lipids) in a composition of the present invention is preferably at least from about 1:10 to about 6:1 nucleic acid:lipid by weight (i.e., 1:10=1 µg nucleic acid:10 nmol lipid).

According to the present invention, a regulatory compound 60 for regulating the expression or biological activity of a biomarker, including a recombinant nucleic acid molecule encoding the biomarker, is typically administered to a patient in a composition. In addition to the recombinant nucleic acid molecule or other biomarker regulatory compound (i.e., a 65 protein, antibody, carbohydrate, small molecule product of drug design), the composition can include, for example, a

44

pharmaceutically acceptable carrier, which includes pharmaceutically acceptable excipients and/or delivery vehicles, for delivering the recombinant nucleic acid molecule or other regulatory compound to a patient (e.g., a liposome delivery vehicle). As used herein, a pharmaceutically acceptable carrier refers to any substance suitable for delivering a therapeutic composition useful in the method of the present invention to a suitable in vivo or ex vivo site. Preferred pharmaceutically acceptable carriers are capable of maintaining a recombinant nucleic acid molecule of the present invention in a form that, upon arrival of the nucleic acid molecule to a target cell, the nucleic acid molecule is capable of entering the cell and being expressed by the cell. Suitable excipients of the present invention include excipients or formularies that transport or help transport, but do not specifically target a nucleic acid molecule to a cell (also referred to herein as non-targeting carriers). Examples of pharmaceutically acceptable excipients include, but are not limited to water, phosphate buffered saline, Ringer's solution, dextrose solution, serumcontaining solutions, Hank's solution, other aqueous physiologically balanced solutions, oils, esters and glycols. Aqueous carriers can contain suitable auxiliary substances required to approximate the physiological conditions of the recipient, for example, by enhancing chemical stability and isotonicity.

Suitable auxiliary substances include, for example, sodium acetate, sodium chloride, sodium lactate, potassium chloride, calcium chloride, and other substances used to produce phosphate buffer, Tris buffer, and bicarbonate buffer. Auxiliary substances can also include preservatives, such as thimerosal, m- or o-cresol, formalin and benzol alcohol. Compositions of the present invention can be sterilized by conventional methods and/or lyophilized.

One type of pharmaceutically acceptable carrier includes a controlled release formulation that is capable of slowly releasing a composition of the present invention into an animal. As used herein, a controlled release formulation comprises recombinant nucleic acid molecule or other biomarker regulatory compound of the present invention in a controlled release vehicle. Suitable controlled release vehicles include, but are not limited to, biocompatible polymers, other polymeric matrices, capsules, microcapsules, microparticles, bolus preparations, osmotic pumps, diffusion devices, liposomes, lipospheres, and transdermal delivery systems. Suitable delivery vehicles have been previously described herein, and include, but are not limited to liposomes, viral vectors or other delivery vehicles, including ribozymes. Natural lipidcontaining delivery vehicles include cells and cellular membranes. Artificial lipid-containing delivery vehicles include liposomes and micelles. As discussed above, a delivery vehicle of the present invention can be modified to target to a particular site in a patient, thereby targeting and making use of a nucleic acid molecule at that site. Suitable modifications include manipulating the chemical formula of the lipid portion of the delivery vehicle and/or introducing into the vehicle a targeting agent capable of specifically targeting a delivery vehicle to a preferred site, for example, a preferred cell type. Other suitable delivery vehicles include gold particles, poly-L-lysine/DNA-molecular conjugates, and artificial chromosomes

As discussed above, a composition of the present invention is administered to a patient in a manner effective to deliver the recombinant nucleic acid molecule comprising a nucleic acid sequence encoding a biomarker protein to a target cell, whereby the target cell is transfected by the recombinant molecule and whereby the biomarker protein is expressed in the target cell. When a biomarker regulatory compound is to

be delivered to a target cell in a patient, the composition is administered in a manner effective to deliver the biomarker regulatory compound to the target cell, whereby the compound can act on the cell (e.g., enter the cell and act on the biomarker or an inhibitor or stimulator thereof) so that the 5 expression or biological activity of the biomarker is increased or decreased, depending on the isoform and the goal of the therapy. Suitable administration protocols include any in vivo or ex vivo administration protocol.

According to the present invention, an effective adminis- 10 tration protocol (i.e., administering a composition of the present invention in an effective manner) comprises suitable dose parameters and modes of administration that result in transfection and expression of a recombinant nucleic acid molecule encoding a biomarker protein or another biomarker 15 regulatory compound, in a target cell of a patient, and subsequent inhibition of the growth of the target cell or inhibition or promotion of angiogenesis, preferably so that the patient obtains some measurable, observable or perceived benefit from such administration. In some situations, where the target 20 cell population is accessible for sampling, effective dose parameters can be determined using methods as described herein for assessment of tumor growth or using methods known in the art for the assessment of angiogenesis. Such methods include removing a sample of the target cell popu- 25 lation from the patient prior to and after the recombinant nucleic acid molecule is administered, and measuring changes in biomarker expression or biological activity, as well as measuring inhibition of the cell or impact on angiogenesis of a suitable cell line. Alternatively, effective dose 30 parameters can be determined by experimentation using in vitro cell cultures, in vivo animal models, and eventually, clinical trials if the patient is human. Effective dose parameters can be determined using methods standard in the art for a particular disease or condition that the patient has or is at 35 risk of developing. Such methods include, for example, determination of survival rates, side effects (i.e., toxicity) and progression or regression of disease.

According to the present invention, suitable methods of nucleic acid molecule of the present invention to a patient include any route of in vivo administration that is suitable for delivering a recombinant nucleic acid molecule into a patient. The preferred routes of administration will be apparent to those of skill in the art, depending on the type of delivery 45 vehicle used, the target cell population, whether the compound is a protein, nucleic acid, or other compound (e.g., a drug) and the disease or condition experienced by the patient. Preferred methods of in vivo administration include, but are not limited to, intravenous administration, intraperitoneal 50 administration, intramuscular administration, intracoronary administration, intraarterial administration (e.g., into a carotid artery), subcutaneous administration, transdermal delivery, intratracheal administration, subcutaneous administration, intraarticular administration, intraventricular 55 administration, inhalation (e.g., aerosol), intracerebral, nasal, oral, pulmonary administration, impregnation of a catheter, and direct injection into a tissue. In an embodiment where the target cells are in or near a tumor, a preferred route of administration is by direct injection into the tumor or tissue sur- 60 rounding the tumor. For example, when the tumor is a breast tumor, the preferred methods of administration include impregnation of a catheter, and direct injection into the tumor.

Intravenous, intraperitoneal, and intramuscular administrations can be performed using methods standard in the art. 65 Aerosol (inhalation) delivery can also be performed using methods standard in the art (see, for example, Stribling et al.,

46

Proc. Natl. Acad. Sci. USA 189:11277-11281, 1992, which is incorporated herein by reference in its entirety). Oral delivery can be performed by complexing a therapeutic composition of the present invention to a carrier capable of withstanding degradation by digestive enzymes in the gut of an animal. Examples of such carriers, include plastic capsules or tablets, such as those known in the art.

One method of local administration is by direct injection. Direct injection techniques are particularly useful for administering a recombinant nucleic acid molecule to a cell or tissue that is accessible by surgery, and particularly, on or near the surface of the body. Administration of a composition locally within the area of a target cell refers to injecting the composition centimeters and preferably, millimeters from the target cell or tissue.

Various methods of administration and delivery vehicles disclosed herein have been shown to be effective for delivery of a nucleic acid molecule to a target cell, whereby the nucleic acid molecule transfected the cell and was expressed. In many studies, successful delivery and expression of a heterologous gene was achieved in preferred cell types and/or using preferred delivery vehicles and routes of administration of the present invention. All of the publications discussed below and elsewhere herein with regard to gene delivery and delivery vehicles are incorporated herein by reference in their entirety. For example, using liposome delivery, U.S. Pat. No. 5,705, 151, issued Jan. 6, 1998, to Dow et al. demonstrated the successful in vivo intravenous delivery of a nucleic acid molecule encoding a superantigen and a nucleic acid molecule encoding a cytokine in a cationic liposome delivery vehicle, whereby the encoded proteins were expressed in tissues of the animal, and particularly in pulmonary tissues. Dow et al. also demonstrated successful in vivo delivery of a nucleic acid molecule by direct injection into a site of a tumor. As discussed above, Liu et al., 1997, ibid. demonstrated that intravenous delivery of cholesterol-containing cationic liposomes containing genes preferentially targets pulmonary tissues and effectively mediates transfer and expression of the genes in vivo. Several publications by Dzau and collaborators demonadministering a composition comprising a recombinant 40 strate the successful in vivo delivery and expression of a gene into cells of the heart, including cardiac myocytes and fibroblasts and vascular smooth muscle cells using both naked DNA and Hemagglutinating virus of Japan-liposome delivery, administered by both incubation within the pericardium and infusion into a coronary artery (intracoronary delivery) (See, for example, Aoki et al., 1997, J Mol. Cell, Cardiol. 29:949-959; Kaneda et al., 1997, Ann N.Y. Acad. Sci. 811: 299-308; and von der Leyen et al., 1995, Proc Natl Acad Sci USA 92:1137-1141).

> As discussed above, delivery of numerous nucleic acid sequences has been accomplished by administration of viral vectors encoding the nucleic acid sequences. Using such vectors, successful delivery and expression has been achieved using ex vivo delivery (See, of many examples, retroviral vector; Blaese et al., 1995, Science 270:475-480; Bordignon et al., 1995, Science 270:470-475), nasal administration (CFTR-adenovirus-associated vector), intracoronary administration (adenoviral vector and Hemagglutinating virus of Japan, see above), intravenous administration (adeno-associated viral vector; Koeberl et al., 1997, Proc Natl Acad Sci USA 94:1426-1431). A publication by Maurice et al., 1999, ibid. demonstrated that an adenoviral vector encoding a β2-adrenergic receptor, administered by intracoronary delivery, resulted in diffuse multichamber myocardial expression of the gene in vivo, and subsequent significant increases in hemodynamic function and other improved physiological parameters. Levine et al. describe in vitro, ex vivo and in vivo

delivery and expression of a gene to human adipocytes and rabbit adipocytes using an adenoviral vector and direct injection of the constructs into adipose tissue (Levine et al., 1998, *J. Nutr. Sci. Vitaminol.* 44:569-572).

In the area of neuronal gene delivery, multiple successful in 5 vivo gene transfers have been reported. Millecamps et al. reported the targeting of adenoviral vectors to neurons using neuron restrictive enhancer elements placed upstream of the promoter for the transgene (phosphoglycerate promoter). Such vectors were administered to mice and rats intramuscularly and intracerebrally, respectively, resulting in successful neuronal-specific transfection and expression of the transgene in vivo (Millecamps et al., 1999, *Nat. Biotechnol.* 17:865-869). As discussed above, Bennett et al. reported the use of adeno-associated viral vector to deliver and express a 15 gene by subretinal injection in the neural retina in vivo for greater than 1 year (Bennett, 1999, ibid.).

Gene delivery to synovial lining cells and articular joints has had similar successes. Oligino and colleagues report the use of a herpes simplex viral vector that is deficient for the 20 immediate early genes, ICP4, 22 and 27, to deliver and express two different receptors in synovial lining cells in vivo (Oligino et al., 1999, Gene Ther. 6:1713-1720). The herpes vectors were administered by intraarticular injection. Kuboki et al. used adenoviral vector-mediated gene transfer and 25 intraarticular injection to successfully and specifically express a gene in the temporomandibular joints of guinea pigs in vivo (Kuboki et al., 1999, Arch. Oral. Biol. 44:701-709). Apparailly and colleagues systemically administered adenoviral vectors encoding IL-10 to mice and demonstrated suc- 30 cessful expression of the gene product and profound therapeutic effects in the treatment of experimentally induced arthritis (Apparailly et al., 1998, *JImmunol*. 160:5213-5220). In another study, murine leukemia virus-based retroviral vector was used to deliver (by intraarticular injection) and 35 express a human growth hormone gene both ex vivo and in vivo (Ghivizzani et al., 1997, Gene Ther. 4:977-982). This study showed that expression by in vivo gene transfer was at least equivalent to that of the ex vivo gene transfer. As discussed above, Sawchuk et al. has reported successful in vivo 40 adenoviral vector delivery of a gene by intraarticular injection, and prolonged expression of the gene in the synovium by pretreatment of the joint with anti-T cell receptor monoclonal antibody (Sawchuk et al., 1996, ibid. Finally, it is noted that ex vivo gene transfer of human interleukin-1 receptor antago- 45 nist using a retrovirus has produced high level intraarticular expression and therapeutic efficacy in treatment of arthritis. and is now entering FDA approved human gene therapy trials (Evans and Robbins, 1996, Curr. Opin. Rheumatol. 8:230-234). Therefore, the state of the art in gene therapy has led the 50 FDA to consider human gene therapy an appropriate strategy for the treatment of at least arthritis. Taken together, all of the above studies in gene therapy indicate that delivery and expression of an biomarker-encoding recombinant nucleic acid molecule according to the present invention is feasible. 55

Another method of delivery of recombinant molecules is in a non-targeting carrier (e.g., as "naked" DNA molecules, such as is taught, for example in Wolff et al., 1990, *Science* 247, 1465-1468). Such recombinant nucleic acid molecules are typically injected by direct or intramuscular administration. Recombinant nucleic acid molecules to be administered by naked DNA administration include a nucleic acid molecule of the present invention, and preferably includes a recombinant molecule of the present invention that preferably is replication, or otherwise amplification, competent. A naked 65 nucleic acid reagent of the present invention can comprise one or more nucleic acid molecule of the present invention in

48

the form of, for example, a dicistronic recombinant molecule. Naked nucleic acid delivery can include intramuscular, subcutaneous, intradermal, transdermal, intranasal and oral routes of administration, with direct injection into the target tissue being most preferred. A preferred single dose of a naked nucleic acid vaccine ranges from about 1 nanogram (ng) to about $100\,\mu g$, depending on the route of administration and/or method of delivery, as can be determined by those skilled in the art. Suitable delivery methods include, for example, by injection, as drops, aerosolized and/or topically. In one embodiment, pure DNA constructs cover the surface of gold particles (1 to 3 μm in diameter) and are propelled into skin cells or muscle with a "gene gun."

In accordance with the present invention, a suitable single dose of a recombinant nucleic acid molecule encoding a biomarker protein as described herein is a dose that is capable of transfecting a host cell and being expressed in the host cell at a level sufficient, in the absence of the addition of any other factors or other manipulation of the host cell, to regulate angiogenesis and/or the tumorigenicity of the host cell when administered one or more times over a suitable time period. Doses can vary depending upon the cell type being targeted, the route of administration, the delivery vehicle used, and the disease or condition being treated.

In one embodiment, an appropriate single dose of a nucleic acid:liposome complex of the present invention is from about 0.1 μg to about 100 μg per kg body weight of the patient to which the complex is being administered. In another embodiment, an appropriate single dose is from about 1 μg to about 10 μg per kg body weight. In another embodiment, an appropriate single dose of nucleic acid:lipid complex is at least about 0.1 μg of nucleic acid, more preferably at least about 1 μg of nucleic acid, even more preferably at least about 10 μg of nucleic acid, and even more preferably at least about 100 μg of nucleic acid, and even more preferably at least about 100 μg of nucleic acid.

Preferably, an appropriate single dose of a recombinant nucleic acid molecule encoding a biomarker protein of the present invention results in at least about 1 pg of protein expressed per mg of total tissue protein per µg of nucleic acid delivered. More preferably, an appropriate single dose is a dose which results in at least about 10 pg of protein expressed per mg of total tissue protein per µg of nucleic acid delivered; and even more preferably, at least about 50 pg of protein expressed per mg of total tissue protein per µg of nucleic acid delivered; and most preferably, at least about 100 pg of protein expressed per mg of total tissue protein per µg of nucleic acid delivered; and most preferably, at least about 100 pg of protein expressed per mg of total tissue protein per µg of nucleic acid delivered.

When the biomarker regulatory agent is a protein, small molecule (i.e., the products of drug design) or antibody, a preferred single dose of such a compound typically comprises between about 0.01 microgram×kilogram⁻¹ and about 10 milligram×kilogram⁻¹ body weight of an animal. A more preferred single dose of an agent comprises between about 1 microgram×kilogram⁻¹ and about 10 milligram×kilograms⁻¹ body weight of an animal. An even more preferred single dose of an agent comprises between about 5 microgram×kilograms⁻¹ and about 7 milligram×kilograms⁻¹ body weight of an animal. An even more preferred single dose of an agent comprises between about 10 microgram×kilogram⁻¹ and about 5 milligram×kilograms⁻¹ body weight of an animal. Another particularly preferred single dose of an agent comprises between about 0.1 microgram×kilograms⁻¹ and about 10 microgram×kilograms⁻¹ body weight of an animal, if the agent is delivered parenterally.

In another embodiment, a targeting vector can be used to deliver a particular nucleic acid molecule into a recombinant

host cell, wherein the nucleic acid molecule is used to delete or inactivate an endogenous gene (e.g., biomarker-encoding gene) within the host cell or microorganism (i.e., used for targeted gene disruption or knock-out technology). Such a vector may also be known in the art as a "knock-out" vector. 5 In one aspect of this embodiment, a portion of the vector, but more typically, the nucleic acid molecule inserted into the vector (i.e., the insert), has a nucleic acid sequence that is homologous to a nucleic acid sequence of a target gene in the host cell (i.e., a gene which is targeted to be deleted or inactivated). The nucleic acid sequence of the vector insert is designed to bind to the target gene such that the target gene and the insert undergo homologous recombination, whereby the endogenous target gene is deleted, inactivated or attenuated (i.e., by at least a portion of the endogenous target gene 15 being mutated or deleted).

Compositions of the present invention can be administered to any mammalian patient, and preferably to humans. According to the present invention, administration of a composition is useful to inhibit the tumorigenicity of a target cell 20 or to treat cancer, or to inhibit angiogenesis in a tissue of a patient. Typically, it is desirable to inhibit the growth of a target cell, or to reduce tumor burden in the patient (tumor numbers and/or volume), or to prevent further growth of the tumor in the patient (tumor stasis), or to obtain any therapeu- 25 tic benefit in the patient (e.g., increased survival). In one embodiment, patients whom are suitable candidates for the method of the present invention include, but are not limited to, patients that have, or are at risk of developing (e.g., are predisposed to), cancer or a lymphoproliferative disease, or any 30 condition in which regulation of angiogenesis might be beneficial. In another embodiment, patients whom are suitable candidates for a method of the invention include, but are not limited to: patients with vascular deficiencies, cardiovascular disease, or patients in whom stimulation of endothelial cell 35 activation and stabilization of newly formed microvessels or other vessels would be beneficial. Increasing or decreasing the expression or biological activity of various biomarkers to inhibit or promote angiogenesis in the absence of obtaining some therapeutic benefit is useful for the purposes of deter- 40 mining factors involved (or not involved) in a disease and preparing a patient to more beneficially receive another therapeutic composition. In a preferred embodiment, however, the methods of the present invention are directed to the inhibition of cancer or inhibition or promotion of angiogenesis in a 45 tissue, which is useful in providing some therapeutic benefit to a patient.

As such, a therapeutic benefit is not necessarily a cure for a particular disease or condition, but rather, preferably encompasses a result which most typically includes allevia- 50 tion of the disease or condition or increased survival, elimination of the disease or condition, reduction of a symptom associated with the disease or condition (e.g. reduced tumor burden), prevention or alleviation of a secondary disease or condition resulting from the occurrence of a primary disease 55 or condition (e.g., metastatic tumor growth resulting from a primary cancer), and/or prevention of the disease or condition. As used herein, the phrase "protected from a disease" refers to reducing the symptoms of the disease; reducing the occurrence of the disease, and/or reducing the severity of the 60 disease. Protecting a patient can refer to the ability of a composition of the present invention, when administered to a patient, to prevent a disease from occurring and/or to cure or to alleviate disease symptoms, signs or causes. As such, to protect a patient from a disease includes both preventing 65 disease occurrence (prophylactic treatment) and treating a patient that has a disease (therapeutic treatment). In particu50

lar, protecting a patient from a disease is accomplished by inhibiting the tumorigenicity of a target cell in the patient or inhibiting or promoting angiogenesis in the cells or tissues of a patient by regulating biomarker expression or biological activity such that a beneficial effect is obtained. A beneficial effect can easily be assessed by one of ordinary skill in the art and/or by a trained clinician who is treating the patient. The term, "disease" refers to any deviation from the normal health of a mammal and includes a state when disease symptoms are present, as well as conditions in which a deviation (e.g., infection, gene mutation, genetic defect, etc.) has occurred, but symptoms are not yet manifested.

One embodiment of the present invention relates to a method (i.e., an assay) for diagnosing or assessing tumor cells (cancer) or the potential therefore in a patient. In one aspect of this embodiment, the method includes the steps of: (a) detecting a level of expression or activity of one or more biomarkers of the present invention in a test sample from a patient to be diagnosed; and (b) comparing the level of expression or activity of the biomarker(s) in the test sample to a normal level of biomarker expression or activity established from a control sample. For example, it is noted that the present inventor has determined that expression of MAGP-2 is upregulated in uterine tumor cells. According to the present invention, detection of the biomarker can be achieved by any method that detects the expression of the biomarker. Detection of a statistically significant difference in biomarker expression or activity in the test sample, as compared to the control level of biomarker expression or biological activity, is an indicator of a difference in the tumorigenicity or potential therefore of cells in the test sample as compared to cells in the control sample. The expression of the biomarker may be cell- and context-specific. Therefore, biomarker expression or activity could be either upregulated or downregulated in a cell as compared to the control. Typically, the biomarker is upregulated or downregulated in the manner associated with the expression of the biomarker during angiogenesis as represented in any one or more of the Tables or experiments described herein. The method of the present invention can be used for any type of tumor wherein the biomarker expression or activity is found to be statistically significantly changed in tumor cells as compared to the corresponding normal cells.

According to the present invention, the phrase "tumorigenicity" refers primarily to the tumor status of a cell or cells (e.g., the extent of neoplastic transformation of a cell, the malignancy of a cell, the propensity for a cell to form a tumor and/or have characteristics of a tumor, or simply the presence or absence of tumor cells in a patient or tissue/organ), which is reflective of a change of a cell or population of cells from a normal to malignant state. Tumorigenicity indicates that tumor cells are present in a sample, and/or that the transformation of cells from normal to tumor cells is in progress, as may be confirmed by any standard of measurement of tumor development. The change typically involves cellular proliferation at a rate which is more rapid than the growth observed for normal cells under the same conditions, and which is typically characterized by one or more of the following traits: continued growth even after the instigating factor (e.g., carcinogen, virus) is no longer present; a lack of structural organization and/or coordination with normal tissue, and typically, a formation of a mass of tissue, or tumor. A tumor, therefore, is most generally described as a proliferation of cells (e.g., a neoplasia, a growth, a polyp) resulting from neoplastic growth and is most typically a malignant tumor. In the case of a neoplastic transformation, a neoplasia is malignant or is predisposed to become malignant. Malignant tumors are typically characterized as being anaplastic (primi-

tive cellular growth characterized by a lack of differentiation), invasive (moves into and destroys surrounding tissues) and/or metastatic (spreads to other parts of the body). As used herein, reference to a "potential for neoplastic transformation", "potential for tumorigenicity" or a "potential for tumor 5 cell growth" refers to an expectation or likelihood that, at some point in the future, a cell or population of cells will display characteristics of neoplastic transformation, including rapid cellular proliferation characterized by anaplastic, invasive and/or metastatic growth.

This method of the present invention has several different uses. First, the method can be used to diagnose tumorigenicity, or the potential for tumorigenicity, or simply the presence or absence of tumor cells, in a subject. The subject can be an individual who is suspected of having a tumor, or an indi- 15 vidual who is presumed to be healthy, but who is undergoing a routine or diagnostic screening for the presence of a tumor (cancer). The subject can also be an individual who has previously been diagnosed with cancer and treated, and who is now under surveillance for recurring tumor growth. The 20 terms "diagnose", "diagnosis", "diagnosing" and variants thereof refer to the identification of a disease or condition on the basis of its signs and symptoms. As used herein, a "positive diagnosis" indicates that the disease or condition, or a potential for developing the disease or condition, has been 25 identified. In contrast, a "negative diagnosis" indicates that the disease or condition, or a potential for developing the disease or condition, has not been identified. Therefore, in the present invention, a positive diagnosis (i.e., a positive assessment) of tumor growth or tumorigenicity (i.e., malignant or 30 inappropriate cell growth or neoplastic transformation), or the potential therefore, means that the indicators (e.g., signs, symptoms) of tumor presence and/or growth according to the present invention (i.e., a change in biomarker expression or biological activity as compared to a baseline control) have 35 been identified in the sample obtained from the subject. Such a subject can then be prescribed treatment to reduce or eliminate the tumor growth. Similarly, a negative diagnosis (i.e., a negative assessment) for tumor growth or a potential therefore or the absence of tumor cells means that the indicators of 40 tumor growth or tumor presence or a likelihood of developing tumors as described herein (i.e., a change in biomarker expression or biological activity as compared to a baseline control) have not been identified in the sample obtained from the subject. In this instance, the subject is typically not pre- 45 scribed any treatment, but may be reevaluated at one or more timepoints in the future to again assess tumor growth. Baseline levels for this particular embodiment of the method of assessment of tumorigenicity of the present invention are typically based on a "normal" or "healthy" sample from the 50 same bodily source as the test sample (i.e., the same tissue, cells or bodily fluid), as discussed in detail below.

In a second embodiment, the method of the present invention can be used more specifically to "stage" a tumor in a patient. Therefore, the patient can be diagnosed as having a 55 tumor or. potential therefore by the method as discussed above, or by any other suitable method (e.g., physical exam, X-ray, CT scan, blood test for a tumor antigen, surgery), and then (or at the same time, when the present method is also used as a diagnostic), the method of the present invention can 60 be used to determine the stage of progression of tumor growth in an individual. For most cancer types, standard staging criteria exist and are known in the art. For example, in breast tumors, there are five different general stages of tumor development which are known and acknowledged in the art as stages 0, I, II, III and IV (although these stages can be grouped into more complex subgroups based on more specific indica-

52

tors). In this embodiment of the method of the present invention, the biomarker expression and/or biological activity in the patient sample is compared to a panel of several different "baseline" levels of biomarker expression or biological activity, wherein each baseline level represents a previously established level for a given stage of the cancer being diagnosed. The ability to "stage" a tumor in the method of the present invention allows the physician to more appropriately prescribe treatment for the patient.

In a third embodiment of this method of the present invention, the method is used to monitor the success, or lack thereof, of a treatment for cancer in a patient that has been diagnosed as having cancer. In this embodiment, the baseline or control level of biomarker expression or biological activity typically includes the previous level of biomarker expression or biological activity in a sample of the patient's tumor, so that a new level of biomarker expression or biological activity can be compared to determine whether tumor cell growth is decreasing, increasing, or substantially unchanged as compared to the previous, or first sample (i.e., the initial sample which presented a positive diagnosis). In addition, or alternatively, a baseline established as a "normal" or "healthy" level of biomarker expression or biological activity can be used in this embodiment, particularly to determine in what manner the biomarker expression is regulated in tumors for the given cell type. This embodiment allows the physician to monitor the success, or lack of success, of a treatment that the patient is receiving for cancer, and can help the physician to determine whether the treatment should be modified. In one embodiment of the present invention, the method includes additional steps of modifying cancer treatment for the patient based on whether an increase or decrease in tumor cell growth is indicated by evaluation of biomarker expression and/or biological activity in the patient.

The first step of the method of the present invention includes detecting biomarker expression or biological activity in a test sample from a patient. According to the present invention, the term "test sample" can be used generally to refer to a sample of any type which contains cells or products that have been secreted from cells (e.g., some biomarkers of the invention are secreted proteins and so one can evaluate a cell supemate, bodily fluid or other media into which such biomarkers may have been secreted by a cell) to be evaluated by the present method, including but not limited to, a sample of isolated cells, a tissue sample and/or a bodily fluid sample. According to the present invention, a sample of isolated cells is a specimen of cells, typically in suspension or separated from connective tissue which may have connected the cells within a tissue in vivo, which have been collected from an organ, tissue or fluid by any suitable method which results in the collection of a suitable number of cells for evaluation by the method of the present invention. The cells in the cell sample are not necessarily of the same type, although purification methods can be used to enrich for the type of cells that are preferably evaluated. Cells can be obtained, for example, by scraping of a tissue, processing of a tissue sample to release individual cells, or isolation from a bodily fluid. A tissue sample, although similar to a sample of isolated cells, is defined herein as a section of an organ or tissue of the body which typically includes several cell types and/or cytoskeletal structure which holds the cells together. One of skill in the art will appreciate that the term "tissue sample" may be used, in some instances, interchangeably with a "cell sample", although it is preferably used to designate a more complex structure than a cell sample. A tissue sample can be obtained by a biopsy, for example, including by cutting, slicing, or a punch. A bodily fluid sample, like the tissue sample, contains

the cells to be evaluated for biomarker expression or biological activity and/or contains the soluble biomarker secreted by cells, and is a fluid obtained by any method suitable for the particular bodily fluid to be sampled. Bodily fluids suitable for sampling include, but are not limited to, blood, mucous, 5 seminal fluid, saliva, breast milk, bile and urine.

In general, the sample type (i.e., cell, tissue or bodily fluid) is selected based on the accessibility and structure of the organ or tissue to be evaluated for tumor cell growth and/or on what type of cancer is to be evaluated. For example, if the 10 organ/tissue to be evaluated is the breast, the sample can be a sample of epithelial cells from a biopsy (i.e., a cell sample) or a breast tissue sample from a biopsy (a tissue sample). The sample that is most useful in the present invention will be cells, tissues or bodily fluids isolated from a patient by a 15 biopsy or surgery or routine laboratory fluid collection.

Once a sample is obtained from the patient, the sample is evaluated for detection of biomarker expression or biological activity in the cells of the sample. The phrase "biomarker expression" can generally refer to biomarker mRNA transcription or biomarker protein translation. Preferably, the method of detecting biomarker expression or biological activity in the patient is the same or qualitatively equivalent to the method used for detection of biomarker expression or biological activity in the sample used to establish the baseline 25 level.

Methods suitable for detecting biomarker transcription include any suitable method for detecting and/or measuring mRNA levels from a cell or cell extract. Such methods include, but are not limited to: polymerase chain reaction 30 (PCR), reverse transcriptase PCR (RT-PCR), in situ hybridization, Northern blot, sequence analysis, gene microarray analysis (gene chip analysis) and detection of a reporter gene. Such methods for detection of transcription levels are well known in the art, and many of such methods are described in 35 detail in the attached examples, in Sambrook et al., *Molecular Cloning: A Laboratory Manual*, Cold Spring Harbor Labs Press, 1989 and/or in Glick et al., *Molecular Biotechnology: Principles and Applications of Recombinant DNA*, ASM Press, 1998; Sambrook et al., ibid., and Glick et al., ibid. are 40 incorporated by reference herein in their entireties.

Measurement of biomarker transcription is suitable when the sample is a cell or tissue sample; therefore, when the sample is a bodily fluid sample containing cells or cellular extracts, the cells are typically isolated from the bodily fluid 45 to perform the expression assay, or the fluid is evaluated for the presence of secreted biomarker protein.

Biomarker expression can also be identified by detection of biomarker translation (i.e., detection of biomarker protein in a sample). Methods suitable for the detection of biomarker 50 protein include any suitable method for detecting and/or measuring proteins from a cell or cell extract. Such methods include, but are not limited to, immunoblot (e.g., Western blot), enzyme-linked immunosorbant assay (ELISA), radioimmunoassay (RIA), immunoprecipitation, immunohis- 55 tochemistry and immunofluorescence. Particularly preferred methods for detection of proteins include any single-cell assay, including immunohistochemistry and immunofluorescence assays. Such methods are well known in the art. Furthermore, antibodies against certain of the biomarkers 60 described herein are known in the art and are described in the public literature, and methods for production of antibodies that can be developed against biomarkers are well known in the art.

The method of the present invention includes a step of 65 comparing the level of biomarker expression or biological activity detected in step (a) to a baseline level (also known as

54

a control level) of biomarker expression or biological activity established from a control sample. According to the present invention, a "baseline level" is a control level, and in some embodiments (but not all embodiments, depending on the method), a normal level, of biomarker expression or activity against which a test level of biomarker expression or biological activity (i.e., in the test sample) can be compared. Therefore, it can be determined, based on the control or baseline level of biomarker expression or biological activity, whether a sample to be evaluated for tumor cell growth has a measurable increase, decrease, or substantially no change in biomarker expression or biological activity, as compared to the baseline level. As discussed above, the baseline level can be indicative of different states of cell tumorigenicity or lack thereof, depending on the primary use of the assay. For example, the baseline level can be indicative of the cell growth expected in a normal (i.e., healthy, negative control, non-tumor) cell sample. Therefore, the term "negative control" or "normal control" used in reference to a baseline level of biomarker expression or biological activity typically refers to a baseline level established in a sample from the patient or from a population of individuals which is believed to be normal (i.e., non-tumorous, not undergoing neoplastic transformation, not exhibiting inappropriate cell growth). For some biomarkers, the negative control may have a higher level of biomarker expression or activity than the tumor type. In another embodiment, a baseline can be indicative of a positive diagnosis of tumor cell growth. Such a baseline level, also referred to herein as a "positive control" baseline, refers to a level of biomarker expression or biological activity established in a cell sample from the patient, another patient, or a population of individuals, wherein the sample was believed, based on data for that cell sample, to be neoplastically transformed (i.e., tumorous, exhibiting inappropriate cell growth, cancerous). In one aspect, the baseline can be indicative of a particular stage of tumor cell growth, which will allow a patient's sample to be "staged" (i.e., the stage of the cancer in the patient can be identified). In yet another embodiment, the baseline level can be established from a previous sample from the patient being tested, so that the tumor growth of a patient can be monitored over time and/or so that the efficacy of a given therapeutic protocol can be evaluated over time. Methods for detecting biomarker expression or biological activity are described in detail above.

The method for establishing a baseline level of biomarker expression or activity is selected based on the sample type, the tissue or organ from which the sample is obtained, the status of the patient to be evaluated, and, as discussed above, the focus or goal of the assay (e.g., diagnosis, staging, monitoring). Preferably, the method is the same method that will be used to evaluate the sample in the patient. In a most preferred embodiment, the baseline level is established using the same cell type as the cell to be evaluated. Baseline levels can be established from an autologous control sample obtained from the patient. According to the present invention, and as used in the art, the term "autologous" means that the sample is obtained from the same patient from which the sample to be evaluated is obtained. The control sample should be of or from the same cell type and preferably, the control sample is obtained from the same organ, tissue or bodily fluid as the sample to be evaluated, such that the control sample serves as the best possible baseline for the sample to be evaluated. In one embodiment, when the goal of the assay is diagnosis of abnormal cell growth, it is desirable to take the control sample from a population of cells, a tissue or a bodily fluid which is believed to represent a "normal" cell, tissue, or bodily fluid, or at a minimum, a cell or tissue which is least likely to be

undergoing or potentially be predisposed to develop tumor cell growth. For example, if the sample to be evaluated is an area of apparently abnormal cell growth, such as a tumorous mass, the control sample is preferably obtained from a section of apparently normal tissue (i.e., an area other than and preferably a reasonable distance from the tumorous mass) in the tissue or organ where the tumorous mass is growing.

In another embodiment, when the goal is to monitor tumor cell growth in the patient, the autologous baseline sample is typically a previous sample from the patient which was taken from an apparent or confirmed tumorous mass, and/or from apparently normal (i.e., non-tumor) tissue in the patient (or a different type of baseline for normal can be used, as discussed below). Therefore, a second method for establishing a baseline level of biomarker expression or biological activity is to establish a baseline level of biomarker expression or biological activity from at least one measurement of biomarker expression or biological activity in a previous sample from the same patient. Such a sample is also an autologous sample, 20 but is taken from the patient at a different time point than the sample to be tested. Preferably, the previous sample(s) were of a same cell type, tissue type or bodily fluid type as the sample to be presently evaluated. In one embodiment, the previous sample resulted in a negative diagnosis (i.e., no 25 tumor cell growth, or potential therefore, was identified). In this embodiment, a new sample is evaluated periodically (e.g., at annual physicals), and as long as the patient is determined to be negative for tumor development, an average or other suitable statistically appropriate baseline of the previous samples can be used as a "negative control" for subsequent evaluations. For the first evaluation, an alternate control can be used, as described below, or additional testing may be performed to confirm an initial negative diagnosis, if desired, $_{35}$ and the value for biomarker expression or biological activity can be used thereafter. This type of baseline control is frequently used in other clinical diagnosis procedures where a "normal" level may differ from patient to patient and/or where obtaining an autologous control sample at the time of 40 diagnosis is not possible, not practical or not beneficial.

In another embodiment, the previous sample from the patient resulted in a positive diagnosis (i.e., tumor growth was positively identified). In this embodiment, the baseline provided by the previous sample is effectively a positive control 45 for tumor growth, and the subsequent samplings of the patient are compared to this baseline to monitor the progress of the tumor growth and/or to evaluate the efficacy of a treatment that is being prescribed for the cancer. In this embodiment, it may also be beneficial to have a negative baseline level of 50 biomarker expression or biological activity (i.e., a normal cell baseline control), so that a baseline for remission or regression of the tumor can be set. Monitoring of a patient's tumor growth can be used by the clinician to modify cancer treatment for the patient based on whether an increase or decrease 55 in cell growth is indicated.

It will be clear to those of skill in the art that some samples to be evaluated will not readily provide an obvious autologous control sample, or it may be determined that collection of autologous control samples is too invasive and/or causes 60 undue discomfort to the patient. In these instances, an alternate method of establishing a baseline level of biomarker expression or biological activity can be used.

Another method for establishing a baseline level of biomarker expression or biological activity is to establish a baseline level of biomarker expression or biological activity from control samples, and preferably control samples that were

56

obtained from a population of matched individuals. It is preferred that the control samples are of the same sample type as the sample type to be evaluated for biomarker expression or biological activity (e.g., the same cell type, and preferably from the same tissue or organ). According to the present invention, the phrase "matched individuals" refers to a matching of the control individuals on the basis of one or more characteristics which are suitable for the type of cell or tumor growth to be evaluated. For example, control individuals can be matched with the patient to be evaluated on the basis of gender, age, race, or any relevant biological or sociological factor that may affect the baseline of the control individuals and the patient (e.g., preexisting conditions, consumption of particular substances, levels of other biological or physiological factors). For example, levels of biomarker expression in the uterine tissue of a normal individual (i.e., having uterine tissue that is not neoplastically transformed or predisposed to such transformation) may be lower or higher in individuals of a given classification (e.g., elderly vs. teenagers, smokers vs. non-smokers) (although such variation in groups is not currently known). To establish a control or baseline level of biomarker expression or biological activity, samples from a number of matched individuals are obtained and evaluated for biomarker expression or biological activity. The sample type is preferably of the same sample type and obtained from the same organ, tissue or bodily fluid as the sample type to be evaluated in the test patient. The number of matched individuals from whom control samples must be obtained to establish a suitable control level (e.g., a population) can be determined by those of skill in the art, but should be statistically appropriate to establish a suitable baseline for comparison with the patient to be evaluated (i.e., the test patient). The values obtained from the control samples are statistically processed using any suitable method of statistical analysis to establish a suitable baseline level using methods standard in the art for establishing such values.

It will be appreciated by those of skill in the art that a baseline need not be established for each assay as the assay is performed but rather, a baseline can be established by referring to a form of stored information regarding a previously determined baseline level of biomarker expression for a given control sample, such as a baseline level established by any of the above-described methods. Such a form of stored information can include, for example, but is not limited to, a reference chart, listing or electronic file of population or individual data regarding "normal" (negative control) or tumor positive (including staged tumors) biomarker expression; a medical chart for the patient recording data from previous evaluations; or any other source of data regarding baseline biomarker expression that is useful for the patient to be diagnosed.

After the level of biomarker expression or biological activity is detected in the sample to be evaluated for tumor cell growth, such level is compared to the established baseline level of biomarker expression or biological activity, determined as described above. Also, as mentioned above, preferably, the method of detecting used for the sample to be evaluated is the same or qualitatively and/or quantitatively equivalent to the method of detecting used to establish the baseline level, such that the levels of the test sample and the baseline can be directly compared. In comparing the test sample to the baseline control, it is determined whether the test sample has a measurable decrease or increase in biomarker expression or biological activity over the baseline level, or whether there is no statistically significant difference

between the test and baseline levels. After comparing the levels of biomarker expression or biological activity in the samples, the final step of making a diagnosis, monitoring, or staging of the patient can be performed as discussed above.

As discussed above, a positive diagnosis indicates that 5 increased cell growth, and possibly tumor cell growth (neoplastic transformation), has occurred, is occurring, or is statistically likely to occur in the cells or tissue from which the sample was obtained. In order to establish a positive diagnosis, the level of biomarker activity is modulated as compared 10 to the established baseline by an amount that is statistically significant (i.e., with at least a 95% confidence level, or p<0.05). Preferably, detection of at least about a 10% change in biomarker expression or biological activity in the sample as compared to the baseline level results in a positive diagnosis of cancer for said sample, as compared to the baseline. More preferably, detection of at least about a 30% change in biomarker expression or biological activity in the sample as compared to the baseline level results in a positive diagnosis of cancer for said sample, as compared to the baseline. More 20 preferably, detection of at least about a 50% change, and more preferably at least about a 70% change, and more preferably at least about a 90% change, or any percentage change between 5% and higher in 1% increments (i.e., 5%, 6%, 7%, 8%...) in biomarker expression or biological activity in the 25 sample as compared to the baseline level results in a positive diagnosis of cancer for said sample. In one embodiment, a 1.5 fold change in biomarker expression or biological activity in the sample as compared to the baseline level results in a positive diagnosis of cancer for said sample. More preferably, 30 detection of at least about a 3 fold change, and more preferably at least about a 6 fold change, and even more preferably, at least about a 12 fold change, and even more preferably, at least about a 24 fold change, or any fold change from 1.5 up in increments of 0.5 fold (i.e., 1.5, 2.0, 2.5, 3.0 . . .) in 35 biomarker expression or biological activity as compared to the baseline level, results in a positive diagnosis of cancer for said sample.

Once a positive diagnosis is made using the present method, the diagnosis can be substantiated, if desired, using 40 any suitable alternate method of detection of tumor cells, including pathology screening, blood screening for tumor antigens, and surgery.

Included in the present invention are kits for assessing angiogenesis in cells or for diagnosing tumor cells (cancer) in 45 a patient. The assay kit includes: (a) reagents for detecting biomarker expression or activity in a test sample (e.g., a probe that hybridizes under stringent hybridization conditions to a nucleic acid molecule encoding the biomarker or a fragment thereof; RT-PCR primers for amplification of mRNA encod- 50 ing the biomarker or a fragment thereof; and/or an antibody, antigen-binding fragment thereof or other antigen-binding peptide that selectively binds to the biomarker); and (b) reagents for detecting a control marker characteristic of a cell type in the test sample (e.g., a probe that hybridizes under 55 stringent hybridization conditions to a nucleic acid molecule encoding a protein marker; PCR primers which amplify such a nucleic acid molecule; and/or an antibody, antigen binding fragment thereof, or antigen binding peptide that selectively binds to the control marker in the sample).

The reagents for detecting of part (a) and or part (b) of the assay kit of the present invention can be conjugated to a detectable tag or detectable label. Such a tag can be any suitable tag which allows for detection of the reagents of part (a) or (b) and includes, but is not limited to, any composition 65 or label detectable by spectroscopic, photochemical, biochemical, immunochemical, electrical, optical or chemical

58

means. Useful labels in the present invention include biotin for staining with labeled streptavidin conjugate, magnetic beads (e.g., DynabeadsTM), fluorescent dyes (e.g., fluorescein, texas red, rhodamine, green fluorescent protein, and the like), radiolabels (e.g., ³H, ¹²⁵I, ³⁵S, ¹⁴C, or ³²p), enzymes (e.g., horse radish peroxidase, alkaline phosphatase and others commonly used in an ELISA), and colorimetric labels such as colloidal gold or colored glass or plastic (e.g., polystyrene, polypropylene, latex, etc.) beads.

In addition, the reagents for detecting of part (a) and or part (b) of the assay kit of the present invention can be immobilized on a substrate. Such a substrate can include any suitable substrate for immobilization of a detection reagent such as would be used in any of the previously described methods of detection. Briefly, a substrate suitable for immobilization of a means for detecting includes any solid support, such as any solid organic, biopolymer or inorganic support that can form a bond with the means for detecting without significantly effecting the activity and/or ability of the detection means to detect the desired target molecule. Exemplary organic solid supports include polymers such as polystyrene, nylon, phenol-formaldehyde resins, acrylic copolymers (e.g., polyacrylamide), stabilized intact whole cells, and stabilized crude whole cell/membrane homogenates. Exemplary biopolymer supports include cellulose, polydextrans (e.g., Sephadex®), agarose, collagen and chitin. Exemplary inorganic supports include glass beads (porous and nonporous), stainless steel, metal oxides (e.g., porous ceramics such as ZrO₂, TiO₂, Al_2O_3 , and NiO) and sand.

According to the present invention, the method and assay for assessing tumor cells in a patient, as well as other methods disclosed herein, are suitable for use in a patient that is a member of the Vertebrate class, Mammalia, including, without limitation, primates, livestock and domestic pets (e.g., a companion animal). Most typically, a patient will be a human patient.

The following examples are provided for the purpose of illustration and are not intended to limit the scope of the present invention. Each publication or other reference disclosed below and elsewhere herein is incorporated herein by reference in its entirety.

EXAMPLES

The following Materials and Methods were used in Examples 1-5 below.

Plasmids

All retroviral expression vectors encoding various putative angiogenic factors were generated by first PCR amplifying their full-length cDNAs from expressed sequence tags using oligonucleotides that facilitated their subsequent subcloning into the pcDNA3.1/Myc-His B vector (Invitrogen). The resulting full-length Myc-His₆-tagged cDNAs were PCR amplified using oligonucleotides that permitted their ligation into the bicistronic retroviral vector, pMSCV-IRES-YFP (Albig and Schiemann, 2005). Table II identifies all of the IMAGE clones and oligonucleotides used to synthesize these retroviral vectors. All putative angiogenic factor inserts were sequenced in their entirety on an Applied Biosystems 377A DNA sequencing machine.

TABLE II

		Cloning oligonucl	eotides
Gene name	Image clone	Oligos for subcloning to pcDNA3.1/Myc-His	Oligos for subcloning to pMSCV-YFP
Matri- lin-2	5063535	5'(NotI)GGCGGCGCGGCCGCATGGAGAAGATGTTGGTG SEQ ID NO: 57	5'(XhoI)GGCGGCCTCGAGATGGAGAAGATGTTGGTG SEQ ID NO: 59
		3'(SacII)GGCGGCCCGCGGTCTGTATTTTAGGCGATT SEQ ID NO: 58	3' (EcoRI)CCGGCCGAATTCTCAATGGTGATGGTGATGATGACC SEQ ID NO: 60
Lumican	5707371	5'(BamH1)GGCGCCGGATCCATGAATGTATGTGCGTTC SEQ ID NO: 61	5' (BgIII) GGCGCCAGATCTATGAATGTATGTGCGTTC SEQ ID NO: 63
		3'(Not1)GGCGCCGGATCCATGAATGTATGTGCGTTC SEQ ID NO: 62	3'(EcoRI)CCGGCCGAATTCTCAATGGTGATGGTGATGATGACC SEQ ID NO: 64
ECM1	5347298	5'(BamHI)GGCGGCGGATCCATGGGGACCGTATCCAGA SEQ ID NO: 65	5'(ECM1)GGCGCCAGATCTATGAATGTATGTGCGTTC SEQ ID NO: 67
		3'(SacII)GGCGGCCCGCGGTTCTTCCTTGGACCCAGG SEQ ID NO: 66	3'(HpaI)GGCCGGGTTAACTCAATGGTGATGGTGATGATG SEQ ID NO: 68
SMOC-2	3988177	5'(HindIII)GGCGGCAAGCTTATGCTGCCGCCACAGCTG SEQ ID NO: 69	5'(BgIII)GGCGGCCTCGAGATGTGGCCCCAACCACCC SEQ ID NO: 71
		3'(SacII)GGCGGCCCGCGGTCCTTGTTTCCTGGGCTG SEQ ID NO: 70	3'(EcoRI)CCGGCCGAATTCTCAATGGTGATGGTGATGATGACC SEQ ID NO: 72
MAGP-2	3469761	5'(HindIII)GGCGGCAAGCTTATGCTGTTCTTGGGGCAG SEQ ID NO: 73	5'(XhoI)GGCGGCCTCGAGATGTGGCCCCAACCACCC SEQ ID NO: 75
		3'(SacII)GGCGGCCGCGGCAGACCATCGGGTCTCTG SEQ ID NO: 74	3'(EcoRI)CCGGCCGAATTCTCAATGGTGATGGTGATGATGACC SEQ ID NO: 76
AK002276	5 1481807	5'(HindIII)GGCGGCAAGCTTATGGCGTCTCGGGAGTCA SEQ ID NO: 77	5'(EcoRI)GGCGGCGAATTCATGGCGTCTCGGGAGTCA SEQ ID NO: 79
		3'(SacIII)GGCGGCCCGCGGTGAAGCCTTGGCTTTCCG SEQ ID NO: 78	3'(EcoRI)CCGGCCGAATTCTCAATGGTGATGGTGATGATGACC SEQ ID NO: 80
CRELD-2	6336331	5'(HindIII)GGCGGCCCGCGGTGAAGCCTTGGCTTTCCG SEQ ID NO: 81	5' (BgIII) GGCGGCAGATCTATGCACCTGCTGCTTGCA SEQ ID NO: 83
		3'(SacII)GGCGGCCCGCGGCAAATCCTCACGGGAGGG SEQ ID NO: 82	3'(XhoI)CCGGCCCTCGAGTCAATGGTGATGGTGATGATGACC SEQ ID NO: 84

The Myc-tagged mammalian expression vectors encoding murine Notch1 [pCS2+mN1FL6MT; (Mumm et al, 2000)] and Jagged-1 [pCS2+Jag1-6MT; (Mumm et al, 2000)] were kindly provided by Dr. Raphael Kopan (Washington University, St. Louis, Mo.). A retroviral Notch1 ICD vector was constructed by PCR amplifying the murine Notch1 ICD domain (amino acids 1744-2531 and contained in pCS2-mN1FL6MT) using a 5' oligonucleotide that contained a unique Xho I restriction site, a Kozak consensus sequence, and a start codon:

(5'GGCGGCCTCGAGGCCACCATGGTGCTGCTGTCCCGC; SEQ ID NO: 121)

and a 3'oligonucleotide that contained a unique Hpa I restriction site, a stop codon, and the C-terminal Myc-tag:

(5'GGCGGCGTTAACTCATGAATTCAAGTCCTCTTCAGA; SEQ ID NO: 122)

The resulting PCR product was ligated into identical restriction sites in the bicistronic retroviral vector, pMSCV-IRES-GFP (Albig and Schiemann, 2005). The pHes1-luciferase, pCMV-Hes1, and pCMV-NICD plasmids were kindly provided by Dr. Jan Jensen (University of Colorado Health Science Center, Denver, Colo.).

Cell Culture and Retroviral Infections

Retroviral supernatants were produced by EcoPack2 retroviral packaging cells (Clontech, Mountain View, Calif.) and used to infect MB114 cells as described previously (Albig et al, 2006; Albig and Schiemann, 2004). Infected cells were analyzed 48 h post-infection and the highest 10% of GFP-expressing cells were collected on a MoFlo cell sorter (Cytomation, Fort Collins, Colo.). Afterward, isolated cells were expanded to yield stable polyclonal populations that were ≥95% positive for transgene expression. Human kidney 293T cells were cultured in DMEM media supplemented with 10% fetal bovine serum (FBS), while human umbilical vein ECs (HUVEC; passages 3-6) were maintained in EGM-2 media (Cambrex Corp., East Rutherford, N.J.) supplemented with EC growth factors (Bullet Kit, Cambrex). Recombinant MAGP-2 Protein Production

A bacterial MAGP-2 expression vector was synthesized by PCR amplifying the full-length MAGP-2 cDNA (less its signal sequence) using oligonucleotides that incorporated unique Nde I (N-terminus) and Bam HI (C-terminus). The resulting PCR fragment was ligated into identical sites in pSBET (Schenk et al, 1995), which appended a FLAG-tag to the C-terminus of MAGP-2. FLAG-tagged recombinant MAGP-2 protein was purified by passing TBS/0.1% Triton X-100-solubilized bacterial cell extracts over a column containing immobilized FLAG-M2 monoclonal antibodies (Sigma, St. Louis, Mo.). Bound proteins were washed ini-

tially with 10 column volumes of TBS/0.1% Triton X-100, followed by an additional 20 column volumes of TBS. Afterward, recombinant MAGP-2 was eluted by addition of 2.5 column volumes of FLAG M2 peptide (100 □g/ml), and subsequently was concentrated by centrifugation against PBS (5 kDa cutoff; Sartorius, Goettingen, Germany). EC Activity Assays

The effect putative angiogenic agents had on MB114 cell activities were determined as follows: (i) cell proliferation using a [³H]thymidine incorporation assay as described (Albig et al, 2006; Albig and Schiemann, 2004; Albig and Schiemann, 2005); (ii) cell invasion through Matrigel matrices using a modified Boyden-chamber assay as described (Albig et al, 2006; Albig and Schiemann, 2005); (iii) p38 MAPK phosphorylation using immunoblot analyses as described (Albig et al, 2006; Albig and Schiemann, 2004; Albig and Schiemann, 2004; Albig and Schiemann, 2005); (iv) angiogenic sprouting in rat tail collagen matrices as described (Albig et al, 2006; Albig and Schiemann, 2004); and (v) Hes1- and SBE-driven luciferase reporter gene assays as described (Albig et al, 2006; Albig and Schiemann, 2004; 20 Albig and Schiemann, 2005).

Notch1 Processing Assay

To monitor the effects of MAGP-2 on the processing and S3 cleavage of Notch1, human kidney 293T cells were transiently transfected in 6-well plates with LT-1 liposomes containing 0.5 µg/well of Notch1 (pCS2+mN1FL6MT), 0.5 μg/well Jagged-1 (pCS2+Jag1-6MT), or 1.5 μg/well of MAGP-2 (pcDNA3.1-MAGP-2/Myc-His) in all combinations. Forty-eight h post-transfection, the cells were washed with ice-cold PBS, lysed immediately in Buffer H/1% Triton X-100 [500 µl/well; (Schiemann et al, 2002)], and incubated on ice for 30 min. Afterward, insoluble material was removed by microcentrifugation and 100 µl of the resulting clarified extract was fractionated through 6% SDS-PAGE gels. The fractionated proteins were transferred electrophoretically to nitrocellulose and probed with anti-Myc $9E\hat{1}0$ monoclonal 35 antibodies (Covance, Princeton, N.J.) to visual Notchi cleavage species.

Matrigel Plug Implantation Assay

The effect of MAGP-2 on vessel formation and infiltration into Matrigel plugs implanted into genetically normal mice

was determined as described (Albig et al, 2006). Briefly, phenol red-free Matrigel (BD biosciences, Bedford, Mass.) was mixed with PBS (diluent), bFGF (50 or 300 ng/ml; R&D Systems, Minneapolis, Minn.), or recombinant MAGP-2 (1 μg/ml) together with bFGF (50 ng/ml), and the resulting mixtures were injected twice subcutaneously in the ventral groin area (400 µl/injection) of C57BL/6 mice. The mice were sacrificed 10 days post-implantation and the Matrigel plugs were dissected, fixed overnight in 10% formalin, and sectioned in the National Jewish Histology Laboratory. Afterward, Masson's trichrome staining was performed to visualize infiltrating vessels, which were quantified under a light microscope by determining the average number of vessels present in 5 random fields (200× magnification). Only those fields that contained at least one vessel in the area underlying the skin were tallied. Two mice were used per experimental condition and this experiment was performed three times in its entirety. All animal studies were performed according to protocol procedures approved by the Animal Care and Use Committee at National Jewish Medical and Research Center. Semi-Quantitative Real-Time PCR

62

Semi-quantitative real-time PCR was performed as previously described (Albig et al, 2006; Albig and Schiemann, 2005). Briefly, MB114 cells were induced to tubulate on Matrigel matrices for 1-25 h, whereupon total RNA was isolated using the RNAqueous kit, followed by an additional round of phenol/chloroform extraction and ethanol precipitation as described above. Total RNA (1 µg) was reverse transcribed with random hexamers and iScript reverse transcriptase according to the manufacturer's recommendations (BioRad, Hercules, Calif.). The resulting cDNA reaction mixtures were diluted 40-fold in H₂O and employed in semiquantitative real-time PCR reactions (25 µl) that used the SYBR Green PCR system (Applied Biosystems, Foster City, Calif.) supplemented with 10 µl of diluted cDNA and 0.1 µM of the oligonucleotide pairs listed in Table III. PCR reactions were performed and analyzed on an ABI 7000 sequence detection system (Applied Biosystems). Differences in RNA concentrations were controlled by normalizing individual gene signals to their corresponding GAPDH RNA signals.

TABLE III

Real-Time PCR oligonucleotides								
Gene name	Real Time PCR Forward Oligonucleotide	Real-Time PCR Reverse Oligonucleotide						
ADAMts1	5' AATGTTTGATGGACAAGCCCC SEQ ID NO: 85	5' TGCTTGGATTCCTCTCCGAA SEQ ID NO: 86						
ADAMts7	5' ACCAGGAACGCCTACCTTTTC SEQ ID NO: 87	5' TCCAGTTTCCTACTTGCCAGC SEQ ID NO: 88						
CTGF	5' CTGCCAGTGGAGTTCAAATGC SEQ ID NO: 89	5' TCATTGTCCCCAGGACAGTTG SEQ ID NO: 90						
Decorin	5' GGCATTCAAACCTCTCGTGAA SEQ ID NO: 91	5' TCATGGACACGAAGTTCCTGG SEQ ID NO: 92						
ECM1	5' CGGAGGAATTCGTGGAAAGA SEQ ID NO: 93	5' CCACTAAAGCCACGTTCCTCA SEQ ID NO: 94						
Inhibin β-a	5' TCCCCAAGGCTAACAGAACCA SEQ ID NO: 95	5' CCCCTTTAAGCCCATTTCCTC SEQ ID NO: 96						
Inhibin β -b	5' CAGACATCGCATCCGCAAA SEQ ID NO: 97	5' AATGATCCAGTCGTTCCAGCC SEQ ID NO: 98						
Integrin α -3	5' AACCCCTTCAAACGGAACCA SEQ ID NO: 99	5' TCGACGTGGACAGCTGAAGAA SEQ ID NO: 100						

TABLE III-continued

	Real-Time PCR oligonu	cleotides
Gene name	Real Time PCR Forward Oligonucleotide	Real-Time PCR Reverse Oligonucleotide
Integrin α -6	5' CTCGTTCTTCGTTCCAGGTTG SEQ ID NO: 101	5' AGCAGCAGCGGTGACATCTAT SEQ ID NO: 102
Lipocalin-7	5' GGACAACTGCAATCGATGCA SEQ ID NO: 103	5' GCCTCGGTTGATGGCTTTAAT SEQ ID NO: 104
LoxI-3	5' AAGTGTGACAGAATGCGCCTC SEQ ID NO: 105	5' ACTTGCAACTGATGCTCCACC SEQ ID NO: 106
Lumican	5' AGTGTGCCAATGGTTCCTCCT SEQ ID NO: 107	5' TGCAGGTCTGTGACGTTCTCA SEQ ID NO: 108
Matrilin-2	5' CACAGGCATCCTGATCTTTGC SEQ ID NO: 109	5' TGAAATTGGCCACCAGGAAG SEQ ID NO: 110
Nephronectin	5' GGTGATGGAGGACATGCGAAT SEQ ID NO: 111	5' TTGTTGGCTTGGAAGTAGGCC SEQ ID NO: 112
SerpinE-2	5' AATCTGATCGATGGTGCCCTT SEQ ID NO: 113	5' CGAATGTCCGTTTCTTTGTGC SEQ ID NO: 114
SMOC-2	5' CACCAAATGGAAGACCCATCA SEQ ID NO: 115	5' ATCATCTGCTTTCCCTGCTCC SEQ ID NO: 116
CRELD-2	5' GCAGAGGAACGAGACCCACAGCATC SEQ ID NO: 117	5' GTGCCCAGCCCACTTCACACTG SEQ ID NO: 118
MAGP-2	5' GCTTGTCTTGGCAGTCAGCATCCC SEQ ID NO: 119	5' GGTCGTCTGTGAATGTCTCAGGCAC SEQ ID NO: 120

Oligonucleotide Microarray Analysis

Murine brain microvascular MB114 ECs were cultured as previously described (Albig et al, 2006; Albig and Schiemann, 2004). To identify genes differentially expressed during angiogenesis, log phase-growing MB114 cells (2×10⁶ cells/plate) were plated onto 10-cm plates that contained 4 ml of solidified Matrigel matrices [diluted 5:3 in serum-free media (SFM)]. Tubulogenesis was allowed to proceed for 1, $_{40}$ 5, 15, or 25 h, at which point the cells were gently washed twice with ice-cold PBS, and subsequently were scraped, together with their Matrigel cushions, into 16 ml of lysis/ binding buffer to isolate total RNA using the RNAqueous kit (Ambion, Austin, Tex.). Isolated total RNA samples were 45 subjected to phenol:choloroform extraction and ethanol precipitation, followed by additional purification using the RNeasy kit (Qiagen, Valencia, Calif.). Afterward, the quality and integrity of purified total RNA (1.5 µg/lane) was analyzed on an Agilent 2100 Bioanalyzer (Agilent Technologies, Palo 50 Alto, Calif.). Biotin-labeled cRNA probes were synthesized using 8 µg of total RNA that was primed with olido-dT and reverse transcribed with Superscript II (Invitrogen, Carlsbad, Calif.), and subsequently were fragmented and hybridized overnight to Affymetrix MOE430A GeneChips according to 55 the manufacturer's recommendations (Affymetrix, Santa Clara, Calif.) in the University of Colorado Health Sciences Center Microarray Core Facility. The microarrays were scanned (2.5-3µ resolution) on a Affymetrix GeneChip Scanner 3000, and differentially expressed mRNAs were identi- 60 fied using GeneSpring 6.0 software (Agilent Technologies). In doing so, individual time points were first compiled into a single experiment that was filtered on flags (i.e., 6 out of 12 flags needed to pass filter). The remaining genes then were filtered by expression levels such that only those genes that 65 were differentially regulated ≥3-fold≤ in at least one time point were considered significant.

Example 1

The following example describes the identification of secretory proteins differentially expressed in tubulating ECs.

To characterize the secretome of ECs undergoing tumorinduced angiogenesis, murine brain microvascular MB114 cells were cultured on tumor-derived basement membranes (i.e., Matrigel matrices) to stimulate angiogenesis activation and the formation of capillary-like structures in vitro. MB114 cells cultured onto Matrigel matrices for 0-25 hours as indicated in FIG. 6 spontaneously reorganized into elongated, capillary-like structures, a response that was readily detected by 5 h, and one that continued to develop over the next 20 h (FIG. 6). Total RNA was isolated at various times after the initiation of tubulogenesis in MB114 cells, and subsequently was used to synthesize biotinylated cRNA probes that were hybridized to Affymetrix MOE430 GeneChips (see Materials and Methods). In doing so, 308 genes were identified whose expression in angiogenic ECs was altered ≥3-fold≤. Of these differentially-expressed genes, 63 genes (~20%) encoded EC secretory proteins (Table I), 35 genes (~11%) encoded transmembrane or membrane-associated proteins (Table V), and 210 genes encoded non-secretory proteins (Table IV). This approach identified several secretory proteins known to be associated with angiogenesis and/or microenvironment remodeling, including ADAMTS1 (Iruela-Arispe et al, 2003), CTGF (Brigstock, 2002), HGF (Gao and Vande Woude, 2005), MMPs 3 and 9 (Heissig et al, 2003), thrombospondins 1 and 2 (Armstrong and Bornstein, 2003), and TIMP3 (Qi et al, 2003) (Table I, bold type face). In addition, numerous secretory proteins not previously associated with angiogenesis were identified (Table I, regular text face). The differential expression of 19 individual genes was verified by semi-quantitative real-time PCR (see Materials and Methods). These analyses showed significant concor-

dance in the expression profiles measured either by real-time PCR or microarray analyses (Table VI), indicating that these

(and other) genes are indeed bona fide targets of angiogenic. signaling systems in tubulating ECs.

 $TABLE\ I$

		TT		tayler 1	during	
				tubulog		_
Name	GenBank#	1	5	15	25	Description
9130213B05Rik	BC006604	1.0	0.3	0.3	0.6	RIKEN cDNA 9130213B05 gene (has signal peptide)
Adamts1	D67076	1.0	0.3	0.6	0.8	Adamts1
Adamts7	AL359935	1.0	2.4	4.8		Adamts7
C1r	NM_023143	1.0	1.0	2.3		complement component 1, r
						subcomponent
C1s	BC022123	1.0	1.0	3.8	10.7	complement component 1, s subcomponent
C3	K02782	1.0	0.7	7.9	23.0	complement component 3
Ccl2	AF065933	1.0	0.7	0.2	0.1	chemokine (C-C motif) ligand
Ccl5	NM_013653	1.0	2.7	3.4		chemokine (C-C motif) ligand
Ccl7	AF128193		0.5	0.3		chemokine (C-C motif) ligand
Ccl8	NM_021443	1.0	1.1	2.8		chemokine (C-C motif) ligand
Cfh	AI987976	1	1.5	3.4		complement component factor
Clu	NM _013492	1.0	0.9	1.7		clusterin
Col3a1	AW550625	1.0	1.3	3.0		procollagen, type III, alpha 1
Col9a3	BG074456	1.0	0.8	6.7		procollagen, type IX, alpha 3
Creld2	AK017880	1.0	3.1	1.0		cysteine-rich with EGF-like domains 2
Csf3	NM_009971	1.0	1.7	0.3	0.3	colony stimulating factor 3 (granulocyte)
Ctgf	NM_010217	1.0	0.2	0.3	0.3	connective tissue growth factor
Cxcl16	BC019961	1	3.9	3.4	3.0	chemokine (C-X-C motif) ligand 16
Cxcl2	NM_009140	1.0	1.2	0.2	0.1	chemokine (C-X-C motif)
CC1	NIM 010516	1.0	0.5	0.2	0.2	ligand 2
Cyr61 Den	NM_010516 NM_007833	$1.0 \\ 1.0$	0.5 2.1	0.3 6.9		cysteine rich protein 61 decorin
Ecm1	NM_007899	1.0	1.7	2.9		extracellular matrix protein 1
F3	BC024886	1.0	0.2	0.2		coagulation factor III
Grem1	BC024880 BC015293	1.0	3.8	2.5		cysteine knot superfamily 1,
3141111	20010200		0.0		0.0	BMP antagonist 1
Hgf	AF042856	1.0	1.2	4.4	5.0	hepatocyte growth factor
Igfbp4	BC019836	1.0	1.7	3.5		insulin-like growth factor
0 1						binding protein 4
Igfbp5	NM_010518	1.0	0.9	3.8	5.2	insulin-like growth factor
						binding protein 5
II6	NM_031168	1.0	3.1	3.2	2.7	interleukin 6
Inhba	NM_008380	1.0	1.9	0.4	0.3	inhibin beta-A
Lbp	NM_008489	1.0	0.7	2.3	5.2	lipopolysaccharide binding
						protein
Lcn2	X14607	1.0	1.3	24.7		lipocalin 2
Len7	BC005738	1.0	0.6	0.3	0.3	lipocalin 7
Lif	AF065917	1.0	0.6	0.2	0.1	leukemia inhibitory factor
Lox13	NM_013586	1.0	1.2	4.0	4.7	lysyl oxidase-like 3
Lum	AK014312	1.0	1.1	1.8	3.2	lumican
MFAP5 (MAGP-2)	NM_015776	1.0	3.2	1.0	1.2	microfibrillar associated
						protein 5
Matn2	BC005429	1.0	1.4	6.4	9.9	Matrilin-2 matrix gamma-
Mglap	NM_008597	1.0	1.9	7.4	17.8	carboxyglutamate (gla)
						protein
Mmp10	NM_019471	1.0	5.4	11.8	12.1	matrix metalloproteinase 10
Mmp11	NM_008606	1.0	1.4	4.9	9.4	matrix metalloproteinase 11
Mmp19	AF153199	1.0	1.9	5.7	9.4	matrix metalloproteinase 19
Мтр3	NM_010809	1.0	1.6	3.5	10.3	matrix metalloproteinase 3
Мтр9	NM_013599	1.0	4.4	5.7	3.6	matrix metalloproteinase 9
Naga	BC021631	1.0	1.6	4.4	8.2	N-acetyl galactosaminidase, alpha
Nbl1	NM_008675	1.0	1.2	2.8	5.8	neuroblastoma, suppression
North	NIM 012600	1.0	0.3	0.1	0.1	of tumorigenicity 1
Ngfb	NM_013609	1.0	0.3	0.1		nerve growth factor, beta
Npnt	AA223007	1	0.6	0.2		Nephronectin
	NM_008728	1.0	0.6	0.2		natriuretic peptide receptor 3
Npr3			1.5	3.6	3.2	olfactomedin 1
Npr3 Olfm1	D78264	1.0				
	D78264 NM_008873	1.0	0.9	0.2		plasminogen activator,
Npr3 Olfm1					0.3	

TABLE I-continued

Secreted proteins differentially regulated during MB114 tubulogenesis.										
Hours of tubulogenesis										
Name	GenBank#	1	5	15	25	Description				
Serpinb2	NM_011111	1.0	1.8	1.4	1.9	proteinase inhibitor, clade B, member 2 serine (or cysteine)				
Serpine1	NM_008871	1.0	0.6	0.2	0.1	proteinase inhibitor, clade E, member 1				
Serpine2	NM_009255	1.0	3.6	16.3	29.5	,				
Sfrp2	NM_009144	1.0	0.8	4.1	5.5	secreted frizzled-related sequence protein 2				
Slpi	NM_011414	1.0	1.2	3.7	6.9					
Smoc2	NM_022315	1.0	7.2	10.6	5.5	Secreted modular calcium binding protein-2				
Tgfb3	BC014690	1.0	5.4	2.2	2.8	transforming growth factor, beta 3				
Thbs1	AI385532	1.0	0.2	0.4	0.5	thrombospondin 1				
Thbs2	NM_011581	1.0	0.9	3.6	6.6	thrombospondin 2				
Timp3	BI111620	1.0	0.6	0.2	0.1	tissue inhibitor of				
						metalloproteinase 3				
U90926	NM_020562	1.0	1.0	0.3	0.3	cDNA sequence U90926 (predicted signal peptide)				
Wisp1	NM_018865	1.0	0.9	0.4	0.2	WNT1 inducible signaling pathway protein 1				

Shown in Table I are differentially-expressed genes that encode for secretory proteins whose expression was altered at least 3-fold in at least one time point during the angiogenic timecourse. in tubulating ECs. Identified genes encoding shown in regular text face.

TABLE IV

		Hou	ırs of T	ubulog	enesis	_				
Name	GenBank#	1	5	15	25	Description				
Abca1	BB144704	1.0	1.6	4.8		ATP-binding cassette, sub-family A (ABC1), member 1				
Abca7	NM_013850	1.0	1.2	3.4	4.1	ATP-binding cassette, sub-family A (ABC1), member 7				
Abcb1a	M30697	1.0	3.6	4.1	2.7	ATP-binding cassette, sub-family B (MDR/TAP), member 1A				
Abhd4	NM_134076	1.0	1.1	3.4	3.8	abhydrolase domain containing 4				
Abtb1	NM_030251	1.0	1.9	5.0	5.4	ankyrin repeat and BTB (POZ) domain containing 1				
Acta2	NM_007392	1.0	0.7	0.2	0.2	actin, alpha 2, smooth muscle, aorta				
Actg2	NM_009610	1.0	0.7	0.3	0.3	actin, gamma 2, smooth muscle, enteric				
Ahi1	BQ175532	1.0	3.2	3.4	2.5	Abelson helper integration site				
Akr1c18	NM_134066	1.0	1.9	6.1		aldo-keto reductase family 1, member C18				
Ampd3	D85596	1.0	1.0	3.7	3.7	AMP deaminase 3				
Ankrd1	AK009959	1.0	0.3	0.3	0.2	ankyrin repeat domain 1 (cardiac muscle)				
Aox1	NM_009676	1.0	1.0	6.7	11.8	aldehyde oxidase 1				
Apbb3	BC024809	1.0	2.0	4.2	6.1	amyloid beta (A4) precursor protein-binding, family B, member 3				
Aps	NM_018825	1.0	2.5	4.1	3.5	adaptor protein with pleckstrin homology and src				
Arc	NM_018790	1.0	0.3	0.2	0.1	activity regulated cytoskeletal-associated protein				
Arg2	NM_009705	1.0	1.4	4.1	5.2	arginase type II				
Ass1	NM_007494	1.0	1.7	3.0	3.7	argininosuccinate synthetase 1				
Bckdha	NM_007533	1.0	1.6	3.3	3.3	branched chain ketoacid dehydrogenase E1, alpha polypeptide				
Atoh8	AK016909	1.0	8.5	9.3		atonal homolog 8 (Drosophila)				
Bbs2	AF342737	1.0	1.8	3.6		Bardet-Biedl syndrome 2 homolog (human)				
Bhlhb2	NM_011498	1.0	0.3	0.2		basic helix-loop-helix domain containing, class B2				
Bst1	AI647987	1.0	1.4	3.9		bone marrow stromal cell antigen 1				
Cbfa2t1h	X79989	1.0	0.4	4.7		CBFA2T1 identified gene homolog (human)				
Cbr2	BC010758	1.0	1.1	5.3		carbonyl reductase 2				
Cenb1	AU015121	1.0	0.9	0.3		cyclin B1				
Ccng2	U95826	1.0	1.7	3.4		cyclin G2				
Cdc6	NM_011799	1.0	0.7	0.2	0.1	cell division cycle 6 homolog (S. cerevisiae)				
Cdk5r	BB177836	1.0	0.5	0.2		cyclin-dependent kinase 5, regulatory subunit (p35)				
Cdkn1a	AK007630	1.0	1.9	0.2	0.1	cyclin-dependent kinase inhibitor 1A (P21)				
Cebpd	BB831146	1.0	3.6	6.5	8.8	CCAAT/enhancer binding protein (C/EBP), delta				
Chc1	NM 133878	1.0	1.0	0.3		chromosome condensation 1				

TABLE IV-continued

	Non-s	ecreto	ry pro	eins dif	ferentia	ally regulated during MB114 tubulogenesis
0.7						
Cit Cte1	AF086823 NM_012006	1.0 1.0	4.0 1.0	3.5 5.0		citron mitochondrial acyl-CoA thioesterase 1
Cyp51	NM_020010	1.0	0.5	0.2		cytochrome P450, 51
Cyp7b1	NM_007825	1.0	3.5	3.9		cytochrome P450, family 7, subfamily b, polypeptide 1
Dbp	BB550183	1.0	0.6	5.1		D site albumin promoter binding protein
Dck	BB030204	1.0	1.0	0.3		deoxycytidine kinase
Dexr	BC012247	1.0	2.3	8.4	20.7	dicarbonyl L-xylulose reductase
Dhrs7	AK009385	1.0	1.8	3.5		dehydrogenase/reductase (SDR family) member 7
Dhrs8	NM_053262	1.0	0.9	4.8		dehydrogenase/reductase (SDR family) member 8
Diap3	NM_019670	1.0	0.5	0.2		diaphanous homolog 3 (Drosophila)
Dio2	AF177196	1.0	0.5	5.5		deiodinase, iodothyronine, type II
Dscr1 Dusp2	AF282255 L11330	1.0	0.5	0.2 0.2		Down syndrome critical region homolog 1 (human) dual specificity phosphatase 2
Dusp2 Dusp9	AV295798	1.0	1.0	0.2		dual specificity phosphatase 9
Ech1	NM_016772	1.0	1.5	3.1		enoyl coenzyme A hydratase 1, peroxisomal
Egr1	NM_007913	1.0	0.2	0.3		early growth response 1
Egr2	X06746	1.0	0.2	0.2		early growth response 2
Erdr1	AJ007909	1.0	0.6	0.3	0.3	DNA segment, Chr 14, Wayne State University 89,
						expressed
Fabp5	BC002008	1.0	1.0	0.3		fatty acid binding protein 5, epidermal
Fbxo32	AF441120	1.0	1.4	9.3		F-box only protein 32
Fos	AV026617	1.0	0.2	0.2		FBJ osteosarcoma oncogene
Fosl1 Foxm1	U34245 NM_008021	1.0	0.8	0.2		fos-like antigen 1 forkhead box M1
Gabpb1	NM_010249	1.0	0.9	0.3		GA repeat binding protein, beta 1
Ggtl3	BC005772	1.0	2.4	3.3		gamma-glutamyltransferase-like 3
Gjb3	NM_008126	1.0	0.9	0.2		gap junction membrane channel protein beta 3
Gstt3	BC003903	1.0	1.3	3.7		glutathione S-transferase, theta 3
Hbp1	BC026853	1.0	1.1	3.0	3.5	high mobility group box transcription factor 1
Hdac11	BC016208	1.0	0.7	4.2		histone deacetylase 11
Hmgcr	BB123978	1.0	0.5	0.3		3-hydroxy-3-methylglutaryl-Coenzyme A reductase
Hmgcs1	BB705380	1.0	0.3	0.3		3-hydroxy-3-methylglutaryl-Coenzyme A synthase 1
Hnrpab Hs6st2	NM_010448 AW536432	1.0	0.8	0.3 0.3		heterogeneous nuclear ribonucleoprotein A/B heparan sulfate 6-O-sulfotransferase 2
Hsd17b7	NM_010476	1.0	0.4	0.3		hydroxysteroid (17-beta) dehydrogenase 7
Idi1	BC004801	1.0	0.5	0.2		isopentenyl-diphosphate delta isomerase
Ier2	NM_010499	1.0	0.5	0.3		immediate early response 2
Ier5	BF147705	1.0	0.5	0.3		immediate early response 5
Ifi203	M74124	1.0	8.5	8.4	7.2	interferon activated gene 205
Ifrd1	NM_013562	1.0	0.4	0.2		interferon-related developmental regulator 1
Junb	NM_008416	1.0	0.4	0.3		Jun-B oncogene
Kenip1	NM_027398	1.0	0.8	0.2		Ky channel-interacting protein 1
Klf4 Kpnb1	BG069413 NM_008379	1.0 1.0	0.5 0.6	0.2 0.3		Kruppel-like factor 4 (gut) karyopherin (importin) beta 1
Lhx1	AV335209	1.0	0.4	0.2		LIM homeobox protein 1
Lyar	NM_025281	1.0	1.0	0.3		Ly1 antibody reactive clone
Mafk	NM_010757	1.0	0.3	0.3		v-maf musculoaponeurotic fibrosarcoma oncogene family,
						protein K (avian)
Map3k5	NM_008580	1.0	3.4	4.2		mitogen activated protein kinase kinase kinase 5
Mark1	BM213279	1.0	1.7	8.6		MAP/microtubule affinity-regulating kinase 1
Mcm3	B1658327	1.0	0.8	0.3 11.2		minichromosome maintenance deficient 3 (S. cerevisiae)
Mgst2 Mthfd2	AV066880 BG076333	1.0	2.3 1.4	0.2		microsomal glutathione S-transferase 2 methylenetetrahydrofolate dehydrogenase (NAD+
Williaz	D G070333	1.0	1.7	0.2	0.2	dependent), methenyltetrahydrofolate cyclohydrolase
Mybl2	NM_008652	1.0	0.9	0.3	0.2	myeloblastosis oncogene-like 2
Myd116	NM_008654	1.0	0.4	0.3		myeloid differentiation primary response gene 116
Myl9	AK007972	1.0	0.6	0.1	0.3	myosin, light polypeptide 9, regulatory
Narg2	BE952805	1.0	4.1	3.7		NMDA receptor-regulated gene 2
Ndrg2	NM_013864	1.0	1.5	5.0		N-myc downstream regulated 2
Ndrg4	AV006122	1.0	1.5	3.8		N-myc downstream regulated 4
Nfatc4	BF227641	1.0	0.9	4.4	5.1	nuclear factor of activated T-cells, cytoplasmic, calcineurin-dependent 4
Nfkbia	NM 010907	1.0	0.5	0.3	0.3	nuclear factor of kappa light chain gene enhancer in B-
1 TIMOTO	1414_010507	1.0	0.5	0.5	0.0	cells inhibitor, alpha
Nolc1	BM213850	1.0	0.8	0.3	0.2	nucleolar and coiled-body phosphoprotein 1
Nr4a2	NM_013613	1.0	1.7	4.3		nuclear receptor subfamily 4, group A, member 2
Nudt7	AK011172	1.0	2.2	3.1	4.1	nudix (nucleoside diphosphate linked moiety X)-type
						motif 7
Pa2g4	AA672939	1.0	0.8	0.3		proliferation-associated 2G4
Parc	BC026469	1.0	1.4	3.4		p53-associated parkin-like cytoplasmic protein
Paxip1	AW742928	1.0	0.8	0.3	0.3	PAX interacting (with transcription-activation domain) protein 1
Pdk2	NM_133667	1.0	0.9	3.7	4.6	pyruvate dehydrogenase kinase, isoenzyme 2
Pdzrn3	NM_018884	1.0	0.7	3.1		semaF cytoplasmic domain associated protein 3
Phyh	NM_010726	1.0	1.3	3.5		phytanoyl-CoA hydroxylase
Plk4	AI385771	1.0	0.7	0.3		polo-like kinase 4 (Drosophila)
Pprc1	BM199989	1.0	0.6	0.3		cDNA sequence BC013720
Ptp4a1	BC003761	1.0	0.4	0.3	0.3	protein tyrosine phosphatase 4a1

TABLE IV-continued

	Non-s	ecreto	ry prote	eins dif	ferentia	ally regulated during MB114 tubulogenesis
Ptpre	U35368	1.0	1.0	0.3		protein tyrosine phosphatase, receptor type, E
Ran	AV090150	1.0	0.9	0.3		RAN, member RAS oncogene family
Rgs16	U94828	1.0	1.3	0.2	0.2	regulator of G-protein signaling 16
Rgs2	AF215668	1.0	2.4	7.9	12.4	regulator of G-protein signaling 2
Rgs5	NM_133736	1.0	2.0	4.8	6.5	regulator of G-protein signaling 5
Rin2	AK014548	1.0	2.1	3.4	4.5	Ras and Rab interactor 2
Rnase4	BC005569	1.0	1.0	5.0	9.2	RIKEN cDNA C730049F20 gene
Rps10	AV283093	1.0	0.7	0.3	0.3	RIKEN cDNA 2210402A09 gene
Sc4mol	AK005441	1.0	0.6	0.2	0.2	sterol-C4-methyl oxidase-like
Sdpr	BE197945	1.0	0.2	0.2	0.2	serum deprivation response
Sesn1	AV016566	1.0	1.2	3.3	3.4	sestrin 1
Shmt1	AF237702	1.0	1.0	0.3	0.1	serine hydroxymethyl transferase 1 (soluble)
Sil	BC004585	1.0	0.7	0.3	0.2	Tall interrupting locus
Snrpa1	BC013777	1.0	0.9	0.3	0.2	small nuclear ribonucleoprotein polypeptide A'
Socs3	BB241535	1.0	2.1	4.3	6.3	suppressor of cytokine signaling 3
Sox9	BC024958	1.0	0.4	0.3	0.3	SRY-box containing gene 9
Srm	NM_009272	1.0	0.9	0.3	0.3	spermidine synthase
T2bp	BB277065	1.0	1.2	4.9	7.1	Traf2 binding protein
Tagln	BB114067	1.0	0.9	0.2		transgelin
Tcofl	AW209012	1.0	0.8	0.3	0.2	Treacher Collins Franceschetti syndrome 1, homolog
Timm8a	W82151	1.0	1.1	0.2		translocase of inner mitochondrial membrane 8 homolog a (yeast)
Tiparp	BB707122	1.0	0.3	0.2	0.2	TCDD-inducible poly(ADP-ribose)polymerase
Tle2	AU067681	1.0	0.9	4.2		transducin-like enhancer of split 2, homolog of Drosophila E(spl)
Tle6	NM_053254	1.0	1.1	3.2	3.5	transducin-like enhancer of split 6, homolog of Drosophila E(spl)
Tnfaip3	NM 009397	1.0	0.4	0.1	0.1	tumor necrosis factor, alpha-induced protein 3
Tnnt2	NM_011619	1.0	10.6	9.1		troponin T2, cardiac
Tprt	AK011869	1.0	0.8	0.3		trans-prenyltransferase
Trib1	AV237242	1.0	0.5	0.2		tribbles homolog 1 (Drosophila)
Trip13	AK010336	1.0	1.0	0.3		thyroid hormone receptor interactor 13
Txnip	AF173681	1.0	2.8	4.3		thioredoxin interacting protein
Ugt1a2	BC019434	1.0	2.4	4.2		UDP glycosyltransferase 1 family, polypeptide A6
Uhrfl	BB702754	1.0	0.7	0.3		ubiquitin-like, containing PHD and RING finger domains, 1
Ung	BC004037	1.0	0.5	0.2		uracil-DNA glycosylase
Xdh	AV286265	1.0	1.1	9.2		xanthine dehydrogenase
Zfp36	X14678	1.0	0.3	0.3		TIS11 (AA 1-183); Mouse TPA-induced TIS11 mRNA.
Zfp36l2	BG094962	1.0	0.3	0.4		zinc finger protein 36, C3H type-like 2
Zfp60	NM_009560	1.0	4.5	6.2		zinc finger protein 60

			Hours of T	ubulogenesis	
				acaregenesis	
Name	GenBank#	1	5	15	25
	AA223007	1.0	0.6	0.2	0.2
	AA414485	1.0	0.7	0.3	0.3
	AA672926	1.0	0.5	0.3	0.2
	AI324124	1.0	0.3	0.2	0.2
	AK009010	1.0	0.6	0.2	0.2
	AK011311	1.0	1.2	0.3	0.2
	AK012043	1.0	0.6	0.3	0.3
	AK014587	1.0	0.4	0.3	0.2
	AK015966	1.0	0.7	0.3	0.3
	AK017688	1.0	2.5	5.9	3.8
	AK018202	1.0	1.7	3.4	3.6
	AU017197	1.0	0.7	0.3	0.1
	AU018569	1.0	1.0	0.3	0.2
	AV167760	1.0	0.5	0.3	0.3
	AV171622	1.0	1.6	3.5	5.3
	AV171622	1.0	1.6	4.6	6.1
	AV171622	1.0	1.5	4.6	7.7
	AV209892	1.0	1.9	3.6	4.3
	AV221013	1.0	0.7	0.3	0.3
	AV232798	1.0	0.5	0.2	0.3
	AV371987	1.0	1.9	3.8	6.3
	AV374246	1.0	0.5	0.3	0.3
	AW488471	1.0	0.8	0.2	0.2
	AW554921	1.0	1.1	0.2	0.0
	AW744519	1.0	5.6	14.8	18.5
	AW744519	1.0	2.1	5.5	6.0
	AY029778	1.0	1.1	16.1	24.5
	BB010153	1.0	1.9	3.1	4.2
	BB042892	1.0	0.3	0.3	0.1
	BB230053	1.0	1.0	0.2	0.2
	BB332449	1.0	1.1	5.8	9.7

TABLE IV-continued

Non-secretory pr	Non-secretory proteins differentially regulated during MB114 tubulogenesis									
BB371300	1.0	3.9	4.5	5.1						
BB377340	1.0	1.2	3.3	4.8						
BB407228	1.0	0.7	0.3	0.3						
BB530223	1.0	1.3	5.0	4.7						
BB550907	1.0	0.5	9.1	32.0						
BB628049	1.0	1.3	3.2	3.5						
BC006604	1.0	0.3	0.3	0.6						
BC006717	1.0	1.8	4.8	5.6						
BC011479	1.0	1.3	3.9	3.8						
BC021353	1.0	0.2	0.2	0.2						
BC021353	1.0	0.3	0.2	0.2						
BC021353	1.0	0.3	0.3	0.3						
BC021407	1.0	1.2	4.4	3.7						
BC021429	1.0	0.9	0.3	0.3						
BC021522	1.0	2.3	4.0	4.1						
BC021842	1.0	1.8	4.5	6.6						
BC022135	1.0	0.6	0.3	0.3						
BC025169	1.0	0.7	0.2	0.1						
BC026867	1.0	0.8	0.3	0.2						
BF118393	1.0	0.7	0.3	0.2						
BF578669	1.0	0.5	0.3	0.2						
BG064632	1.0	10.2	14.7	16.6						
BG066982	1.0	0.8	0.3	0.2						
BG075321	1.0	0.7	3.7	5.0						
BG080055	1.0	0.7	0.3	0.2						
BG143461	1.0	0.5	0.3	0.2						
BG868949	1.0	1.3	3.6	3.5						
BG868949	1.0	1.3	4.4	4.3						
BI251603	1.0	1.9	4.0	3.8						
BI454991	1.0	2.0	3.7	4.2						
BI466783	1.0	0.5	0.2	0.2						
BI558298	1.0	1.1	0.3	0.2						
BI660196	1.0	1.2	3.6	4.4						
BM117243	1.0	1.4	3.3	4.1						
BM117243	1.0	1.6	3.6	3.9						
BM200151	1.0	1.0	0.3	0.3						
BM213835	1.0	0.8	0.3	0.2						
BM247465	1.0	0.5	0.2	0.1						
C78203	1.0	2.5	3.7	3.4						
NM_020562	1.0	1.0	0.3	0.3						
NM_026235	1.0	1.3	3.1	7.2						
NM 026839	1.0	0.7	0.3	0.2						
NM_030697	1.0	0.5	3.4	5.0						
NM_054098	1.0	2.1	15.0	24.3						
NM_133706	1.0	1.0	0.3	0.2						
NM_133775	1.0	1.8	3.2	4.3						

Genes encoding non-secretory proteins that demonstrated at least 3-fold differential expression in at least one time-point over a 25 h angiogenesis timecourse.

TABLE V

Transme	Transmembrane proteins differentially regulated during MB114 tubulogenesis								
Hours of tubulogenesis									
Name	GenBank	1	5	15	25	Description			
0610007C21Rik	AK002276	1.0	1.5	2.1	3.3	Clone IMAGE: 1513950, mRNA (predicted transmembrane)			
1810014L12Rik	NM_133706	1.0	1.0	0.3	0.2	RIKEN cDNA 1810014L12 gene (predicted transmembrane)			
Alcam	U95030	1	3.4	4.2	2.6	activated leukocyte cell adhesion molecule			
Anpep	NM_008486	1	3.5	7.0	9.3	alanyl (membrane) aminopeptidase			
Areg	NM_009704	1	0.7	0.2	0.1	amphiregulin calcium channel, voltage-			
Cacna2d1	NM_009784	1.0	2.3	3.9	4.3	dependent, alpha2/delta subunit 1			
Cd14	NM_009841	1.0	2.0	4.0	6.4	CD14 antigen			
Cd38	BB256012	1.0	4.5	4.8	5.1	CD38 antigen			
Cd44	X66083	1.0	1.2	0.3	0.2	CD44 antigen			
Cd53	NM_007651	1.0	2.0	9.6	10.4	CD53 antigen			
Dtr	L07264	1.0	0.4	0.1	0.1	diphtheria toxin receptor			
Emp2	AF083876	1	2.6	3.1	3.2	epithelial membrane protein 2			

75

TABLE V-continued

Tı	ansmembrane proteins	differe	ntiall	y regu	lated	during MB114 tubulogenesis
		Hours	of tu	bulog	enesis	<u>s_</u>
Name	GenBank	1	5	15	25	Description
Epha2	NM_010139	1.0	0.4	0.2	0.2	Eph receptor A2
Fegrt	NM_010189	1.0	1.1	2.5	6.1	Fc receptor, IgG, alpha chain
Islr	NM_012043	1.0	1.2	2.4	4.2	transporter immunoglobulin superfamily containing leucine-rich repeat
Itga3	NM_013565	1.0	0.9	0.3	0.2	integrin alpha 3
Itga6	BM935811	1.0	1.3		0.1	integrin alpha 6
Ldlr	AF425607	1.0	0.2		0.2	low density lipoprotein receptor
Lrp1	NM_008512	1.0	1.3	3.2	5.5	low density lipoprotein receptor- related protein 1
Lrp2	C80829	1.0	0.5	0.3	0.2	low density lipoprotein receptor- related protein 2
Ly6a	BC002070	1.0	0.7	2.3	4.8	lymphocyte antigen 6 complex,
Npr3	NM 008728	1	0.5	0.2	0.2	natriuretic peptide receptor 3
P2rx4	AJ251462	1	1.1	3.2	5.2	purinergic receptor P2X, ligand- gated ion channel 4
Pedh18	AK014140	1.0	0.2	0.3	0.3	protocadherin 18
Pedhb9	NM_053134	1.0	1.1	3.2	4.7	protocadherin beta 9
Ptpre	U35368	1.0	1.0	0.3	0.3	protein tyrosine phosphatase, receptor type, E
Ramp1	NM_016894	1.0	1.3	4.0	5.9	receptor (calcitonin) activity modifying protein 1
Sele	NM 011345	1.0	1.3	0.3	0.3	selectin, endothelial cell
Slc4a3	NM_009208	1	1.7	3.1	4.5	solute carrier family 4 (anion exchanger), member 3 solute carrier family 7 (cationic
Slc7a5	BC026131	1	1.4	0.3	0.1	amino acid transporter, y+ system), member 5
Tfre	AK011596	1.0	1.1	0.3	0.3	transferrin receptor
Tm4sf12	BB072896	1.0	2.3	3.3	3.5	transmembrane 4 superfamily member 12
Tmc6	BC004840	1.0	2.1	3.4	3.3	transmembrane channel-like gene family 6

Genes encoding transmembrane or membrane-associated proteins that demonstrated at least 3-fold differential expression in at least one time-point over the 25 h angiogenesis timecourse. Identified genes encoding known angiogenic 40 regulators are shown in bold type face. Identified genes encoding putative angiogenic regulators are shown in regular text face.

TABLE VI

Real-Time PCR analysis of select proteins												
		Hrs. of Tu	ıbulogenesis									
Name	1	5	15	25								
ADAMts1	1.0	0.4	1.6	2.4								
ADAMts7	1.0	2.0	4.9	5.1								
CRELD-2	1.0	11.4	5.8	10.0								
CTGF	1.0	0.3	0.4	0.3								
Decorin	1.0	3.6	8.4	16.6								
ECM1	1.0	4.3	6.3	9.0								
Inhibin β-a (Inhβ-a)	1.0	4.9	1.4	1.1								
Inhibin β-b (Inhβ-b)	1.0	0.1	0.5	0.7								
Integrin α-3	1.0	1.4	0.8	0.3								
Integrin α-6	1.0	1.2	0.6	0.4								
Lipocalin-7	1.0	0.9	0.6	0.6								
Loxl-3	1.0	2.8	18.0	17.9								
Lumican	1.0	0.4	0.9	1.7								
MAGP-2	1.0	8.4	2.3	4.2								
Matrilin-2	1.0	1.6	6.7	8.0								
Nephronectin	1.0	0.9	0.5	0.5								
SerpinE2	1.0	0.8	5.1	10.1								
SMOC-2	1.0	21.5	58.3	13.1								
TIMP-3	1.0	2.5	0.5	0.5								

Real-time PCR analysis was conducted to confirm differential expression of selected genes from microarray analysis.

Example 2

The following example describes the effects of putative angiogenic gene expression on EC activities-coupled to angiogenesis.

The microarray analyses described in Example 1 identified numerous genes whose expression is regulated by angiogenesis, indicating that the expression of these genes is required during vessel formation. To test this hypothesis and to identify novel regulators of EC activities-coupled to angiogenesis, a series of in vitro assays was performed that modeled angiogenesis activation in ECs (Albig et al, 2006; Albig and Schiemann, 2004; Albig and Schiemann, 2005). In doing so, bicistronic retroviral transduction of MB114 cells was used to 55 stably express six identified secretory proteins, namely matrilin-2, CRELD-2 (cysteine-rich with EGF-Like domains-2), MAGP-2, lumican, SMOC-2 (secreted modular calciumbinding protein-2), and ECM-1, (extracellular protein-1), and one putative transmembrane protein, AK002276. Immunoblotting and semi-quantitative real-time PCR analyses both showed that the expression of all individual transgenes were readily detected in MB114 cells (FIGS. 7A and 7B). In these experiments, MB114 cells were infected with retrovirus encoding either GFP (i.e., control) or various potential angiogenic agents as indicated. Afterward, infected cells were FACS-sorted by GFP expression (highest 10%) to establish stable polyclonal populations of transgenic MB114 cells.

Transgene expression was detected by immunoblotting nickel-captured secretory proteins with anti-Myc antibodies, except AK002276 which was captured from detergent-solubilized cell extracts (FIG. 7A) and by performing semi-quantitative real-time PCR (FIG. 7B).

FIG. 1A show results from an experiment in which serumstarved MB114 cells, stably expressing either GFP or various putative angiogenic agents, were stimulated in the absence or presence of either bFGF (50 ng/ml) or EGF (10 ng/ml) for 24 h at 37° C. Differences in MB114 cell DNA synthesis was 10 determined by measuring [3H]thymidine incorporation into cellular DNA. Functionally, MAGP-2 and SMOC-2 expression significantly enhanced the proliferative response of MB114 cells to bFGF, while MAGP-2 and AK002276 expression significantly enhanced that to EGF (FIG. 1A). In 15 contrast, expression of all other transgenes failed to effect the proliferative response of MB114 cells to either bFGF or EGF (data not shown). FIG. 1B shows that SMOC-2, MAGP-2, and CRELD-2 expression all significantly induced MB114 cell invasion through synthetic basement membranes, a 20 response that was not mimicked by expression of additional transgenes (data not shown). In this experiment, invasion of MB114 cells expressing either GFP or various putative angiogenic agents through synthetic basement membranes was determined over 48 h using a modified Boyden-chamber 25

The inventors' previous studies have associated stimulation of p38 MAPK activity with angiogenesis of MB114 cells and, conversely, inhibition of p38 MAPK activity with angiostasis of MB114 cells (Albig et al, 2006; Albig and 30 Schiemann, 2004; Albig and Schiemann, 2005). Serumstarved MB114 cells expressing MAGP-2 (FIG. 1C) or lumican (FIG. 1D) were stimulated with either bFGF (50 ng/ml) or EGF (10 ng/ml) 0-15 min as indicated in the figures. The phosphorylation status of p38 MAPK was determined by 35 immunoblotting whole cell lysates with phospho-specific p38 MAPK antibodies (p38-P). Differences in protein loading were monitored by reprobing stripped membranes with anti-p38 MAPK polyclonal antibodies (p38). FIG. 1C shows that MAGP-2 expression significantly enhanced p38 MAPK 40 phosphorylation in MB114 cells stimulated with either bFGF or EGF stimulation. In contrast, lumican expression significantly inhibited p38 MAPK activation in MB114 cells treated with either growth factor (FIG. 1D).

Finally, it was determined whether expression of these 45 putative angiogenic factors could effect the angiogenic sprouting of quiescent MB114 cells monolayers. MB114 cells expressing either GFP or various putative angiogenic agents were grown to confluency, and subsequently were overlaid with rat tail collagen matrices. Angiogenic sprouting 50 by quiescent EC monolayers was stimulated by inclusion of 10% FBS and allowed to proceed for 5 days. The quantity of invading angiogenic sprouts was determined by manual counting under a light microscope. FIG. 1E shows that expression of CRELD-2, matrillin-2, or AK002276 failed to 55 significantly affect MB114 cell angiogenic sprouting in response to serum. In stark contrast, expression of MAGP-2 or SMOC-2 both significantly increased the sprouting of MB114 cells cell sprouting, while that of lumican and ECM-1 significantly decreased the ability of MB114 cells to form 60 angiogenic sprouts in collagen matrices (FIG. 1E).

Collectively, these findings demonstrate that tubulating ECs upregulate expression of lumican and ECM-1 during the latter stages of angiogenesis, consistent with their involvement in mediating angiogenesis resolution. Accordingly, both 65 proteins antagonized angiogenic sprouting in MB114 cells, and as such, the inventors propose lumican and ECM-1 as

78

novel mediators of angiostasis. Conversely, tubulating ECs were observed to upregulate expression of MAGP-2 and SMOC-2 during the early stages of angiogenesis, implicating their involvement in mediating angiogenesis activation. Indeed, both proteins stimulated various angiogenic activities, including angiogenic sprouting in MB114 cells. Thus, it is proposed herein that MAGP-2 and SMOC-2 are novel mediators of angiogenesis. Because MAGP-2 was the only protein to exhibit angiogenic activity in all measured indices in vitro, the inventors chose to further characterize the molecular mechanisms whereby MAGP-2 induces angiogenesis in quiescent ECs.

Example 3

The following example demonstrates that MAGP-2 promotes angiogenesis in vivo.

The ability of MAGP-2 to stimulate EC activities coupled to angiogenesis in vitro indicated that MAGP-2 may function to induce vessel formation in vivo. The inventors tested this hypothesis by utilizing the Matrigel plug implantation assay, which monitors the ability of various angiogenic agents to alter vessel formation and infiltration into Matrigel plugs implanted subcutaneously into normal mice. In doing so, first, recombinant FLAG-tagged MAGP-2 (rMAGP-2) was expressed and purified from bacterial cells (FIG. 2A). More particularly, recombinant FLAG-tagged MAGP-2 (rMAGP-2) was purified from detergent-solubilized bacterial cell extracts by anti-FLAG chromatography. MAGP-2 purity was monitored by coomassie staining, and by immunoblotting with anti-FLAG M2 monoclonal antibodies (FIG. 2A; right panel). rMAGP-2 (1 µg/ml) stimulated angiogenic sprouting of quiescent MB114 cell monolayers (FIG. 2A; left panel). Similar to its constitutive expression in MB114 cells, purified rMAGP-2 protein (1 µg/ml) also was found to stimulate angiogenic sprouting of quiescent MB114 cells, thereby demonstrating that these rMAGP-2 preparations were biologically active (FIG. 2A). To further demonstrate that MAGP-2 promotes angiogenesis in vivo, C57BL/6 female mice were injected subcutaneously with Matrigel supplemented either with diluent (D), bFGF (50 ng/ml, LD; or 300 ng/ml, HD), or bFGF (50 ng/ml) in combination with MAGP-2 (1 μg/ml). Mice were sacrificed on day 10 and the plugs harvested and photographed (FIG. 2B; left panels). Afterward, the Matrigel plugs were fixed, sectioned, and stained with Masson's trichrome to visualize infiltrating blood vessels (FIG. 2B; right panels; arrows denote blood vessels), which were quantified by manual counting under a light microscope. FIG. 2B shows that bFGF dose-dependently stimulated significant vascularization of implanted Matrigel plugs. Importantly, rMAGP-2 administration (1 μg/ml) significantly increased the development and infiltration of vessels into Matrigel plugs supplemented with bFGF as compared to those solely containing bFGF (FIG. 2B). Collectively, these findings, together with the in vitro analyses, provide strong evidence implicating MAGP-2 as a bona fide promoter of angiogenesis.

Example 4

The following example demonstrates that MAGP-2 inhibits Notch1 signaling.

MAGP-2 can interact physically with Notch1 and its ligand, Jagged-1 (Miyamoto et al, 2006; Nehring et al, 2005), resulting in the ectodomain shedding of both molecules from the cell surface. Notch signaling also plays an essential role in regulating normal vessel development and angiogenesis in

mammals (Leong and Karsan, 2005; Shawber and Kitajewski, 2004). Given these two facts, the inventors hypothesized that MAGP-2 promotes angiogenesis by modulating Notch1 signaling. To test this hypothesis, first measured were changes in luciferase expression driven by a Hes1-luciferase 5 reporter gene whose expression is induced by NotchI activation (Iso et al, 2003). MB114 and HUVEC cells were transiently transfected either with pHesI-luciferase, pCMV-βgal, and MAGP-2 cDNAs, or with pHes1-luciferase and pCMV-β-gal cDNAs and subsequently stimulated with rMAGP-2 (1 or 5 μg/ml). Afterward, luciferase and β-gal activities contained in detergent-solubilized cell extracts were measured. In addition, GFP- and MAGP-2-expressing MB114 cells were transiently transfected with pHes1-luciferase and pCMV-β-gal cDNAs, together with or without 15 angiogenesis by antagonizing Notch signaling. Jagged-1 cDNA as indicated. Afterward, luciferase and β-gal activities were measured as above. FIG. 3A shows that MAGP-2 expression in or rMAGP-2 treatment of either MB114 or HUVEC cells repressed Hes1-driven luciferase activity. More importantly, MAGP-2 expression abrogated 20 the ability of Jagged-1 to induce Hes1-luciferase activity in MB114 cells (FIG. 3B), suggesting that MAGP-2 functions to antagonize Jagged-1 and, consequently, Notch1 signaling in

Activation of Notch1 signaling involves three proteolytic 25 processing events, termed S1, S2, and S3, that produce three distinct Notch1 fragments, termed TMIC, NEXT, and NICD, respectively (Mumm et al, 2000). NICD production is mediated by a gamma-secretase cleavage reaction that cuts Notch1 at a membrane proximal cytoplasmic site (Mumm et al, 30 2000), resulting in the release and subsequent translocation of NICD to the nucleus where it regulates the expression of Notch1-responsive genes, including Hes1 (Iso et al, 2003). The findings described above indicate that MAGP-2 antagonizes Notch1 signaling, and as such, indicate that MAGP-2 35 may do so by inhibiting Notch1 proteolytic processing. The inventors tested this possibility by transiently transfecting human 293T cells with cDNAs encoding Myc-tagged versions of Notch1, Jagged-1, and MAGP-2 in all combinations, and subsequently monitored changes in NICD production 40 and accumulation by immunoblot analyses using anti-Myc monoclonal antibodies. As expected, Jagged-1 expression significantly enhanced Notch1 processing and the production of NICD as compared to cells solely expressing Notch1 (FIG. 4A). Importantly, the ability of Jagged-1 to induce Notch1 45 cleavage and NICD production in 293T cells was reduced significantly by co-expression of MAGP-2 (FIG. 4A). Thus, these findings indicate that MAGP-2 inhibits Notch1 signaling and Hes1 expression in part by preventing Notch1 processing and NICD production.

To further investigate the impact of MAGP-2 on Notch1 processing and NICD accumulation, the inventors took advantage of recent findings showing that the ability of TGF-β to induce Hes1 promoter activity requires Smad3 to interact physically with NICD (Blokzijl et al, 2003), a reac-5 tion that is dispensable for canonical Smad3-mediated signaling stimulated by TGF-β (Blokzijl et al, 2003). It was therefore reasoned that the ability of MAGP-2 to inhibit NICD production in ECs would reduce the capacity of TGF-β to induce luciferase expression driven by the Hes1 promoter, 6 but not that driven by the synthetic Smad2/3-binding element (SBE). GFP- and MAGP-2-expressing MB114 cells were transiently transfected with either pHesI- or pSBE-luciferase, both together with pCMV-β-gal as indicated in FIG. 4B. Afterward, the resulting transfectants were stimulated over- 65 Expression of various components of the Notch signaling night with increasing concentrations of TGF-β1 (0-5 ng/ml). MAGP-2 expression in MB114 cells significantly decreased

80

the ability of TGF-β to stimulate Hes1-luciferase activity, but had no effect on its stimulation of SBE-luciferase activity (FIG. 4B). Similar effects of MAGP-2 on TGF-β-stimulated Hes1- and SBE-luciferase activities also were observed in HUVEC cells, indicating that MAGP-2-mediated inhibition of Notch1 processing and NICD production was not restricted solely to MB114 cells (data not shown). Collectively, these findings demonstrate that MAGP-2 antagonizes Notch1 signaling by preventing its cleavage and ultimate release of the Notch1 signaling fragment, NICD.

Example 5

The following examples shows that MAGP-2 promotes

Based on the findings described in the Examples above, the inventors hypothesized that MAGP-2 promotes angiogenesis by antagonizing Notch1 signaling. To test this hypothesis, it was first determined whether inhibiting Notch signaling in MB114 cells would enhance their angiogenic sprouting. In doing so, MB114 cells were transiently transfected with the Hes1-luciferase reporter gene (and pCMV-β-gal cDNA as control), and subsequently were treated overnight with or without the highly specific gamma-secretase inhibitor, DAPT (Sastre et al, 2001), which inhibits S3-mediated cleavage of Notch1 and, consequently, NICD-mediated induction of Hes1 expression. Afterward, luciferase and β-gal activities were determined. As expected, DAPT administration (10 μM) significantly inhibited Hes1 promoter activity in MB114 cells (FIG. 5A). More importantly, MB114 cells treated with DAPT formed significantly more angiogenic sprouts than did their untreated counterparts (FIG. 5B). In this experiment, quiescent MB114 cell monolayers were overlaid with rat tail collagen matrices, and were induced to form angiogenic sprouts by addition of 10% FBS supplemented with or without DAPT (10 μ M). Five days later the number of invading angiogenic spouts were quantified by manual counting on a light microscope. Based on these findings, the inventors conclude that Notch activation functions in mediating angiostasis in MB114 cells. This conclusion is bolstered further by the inventors' observation that the Notch ligands Jagged-1 and Delta-like-4, and the Hes1 transcription factor were all strongly downregulated in tubulating MB114 cells (Table VII). Collectively, these findings indicate that Notch1 signaling antagonizes angiogenic sprouting in MB114 cells, and that downregulation of Notch1 signaling components is necessary for angiogenesis activation in MB114 cells.

TABLE VII

		F	Hours of Tu	<u>bulogenesi</u>	s
Name	Genbank	1	5	15	25
DII1	NM_007865	1.0	0.9	1.3	1.1
DII3	AB013440	1.0	0.8	0.6	0.9
DII4	AK004739	1.0	1.2	0.3	0.4
Jag1	AA880220	1.0	0.7	0.2	0.2
Jag2	AV264681	1.0	0.4	0.7	1.3
Notch1	NM_008714	1.0	1.4	0.6	0.7
Notch2	D32210	1.0	1.1	1.1	1.2
Notch3	NM_008716	1.0	0.9	1.3	1.5
Notch4	NM_010929	1.0	1.1	1.1	1.1
Hes1	BC018375	1.0	0.5	0.2	0.1

pathway during MB114 cell tubulogenesis on Matrigel matri-

Having shown that Notch1 signaling mediates angiostasis in MB114 cells, the inventors next asked whether MAGP-2 promotes angiogenesis in MB114 cells via its ability to antagonize Notch signaling. To do so, MAGP-2-expressing MB114 cells were engineered to constitutively express active 5 Notch1 NICD fragment in an attempt to overcome the block of Notch processing mediated by MAGP-2. More particularly, GFP-, MAGP-2-, and MAGP-2/N1ICD-expressing MB114 cells were transiently transfected with pHes1-luciferase and pCMV-β-gal cDNAs. Luciferase and P-gal 10 activities were determined 48 h post-transfection. As the inventors observed previously, MAGP-2 expression reduced Hes1-luciferase activity in MB114 cells (FIG. 5C), a reaction that was bypassed by co-expression of NICD in these cells (FIG. 5C). More importantly, the ability of MAGP-2 to pro- 15 mote angiogenic sprouting was prevented completely by constitutive N1ICD expression in MB114 cells (FIG. 5D). In this experiment, quiescent monolayers of GFP-, MAGP-2-, and MAGP-2/N1ICD-expressing MB114 cells were overlaid with rat tail collagen matrices and incubated in the absence or 20 presence of 10% FBS for 5 days. Afterward, the number of invading angiogenic sprouts were determined by manual counting under a light microscope. Taken together, these results demonstrate that Notch1 activation antagonizes angiogenesis in MB114 cells, and most notably, that 25 MAGP-2 promotes angiogenesis in part via its ability to antagonize Notch1 processing and signaling in ECs.

Example 6

The following example shows that MAGP-2 is expressed aberrantly in the majority of human uterine tumors.

Radiolabeled cDNA probes corresponding to either murine MAGP-2 (FIG. **8**A; upper panel) or human ubiquitin (FIG. **8**A; lower panel) were hybridized to matched human normal:tumor cDNA array. The resulting phosphor-images depict MAGP-2 and ubiquitin expression in paired normal (upper spot) and malignant (bottom spot) uterine tissue. MAGP-2 expression was normalized to that of ubiquitin, followed by a determination of tumor:normal tissue MAGP-2 expression ratios. Ratios ≥ 2 or ≤ 0.5 were considered significant. The results showed that MAGP-2 is expressed aberrantly in the majority of human uterine tumors tested.

Each publication or other reference disclosed below and elsewhere herein is incorporated herein by reference in its entirety.

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 Williams et al., (2006) Blood 107: 931-939
 Zimrin et al., (1996) J Biol Chem 271: 32499-32502

60 U.S. Provisional Application No. 60/722,694 U.S. Provisional Application No. 60/816,969

While various embodiments of the present invention have been described in detail, it is apparent that modifications and adaptations of those embodiments will occur to those skilled

65 in the art. It is to be expressly understood, however, that such modifications and adaptations are within the scope of the present invention, as set forth in the following claims.

SEQUENCE LISTING

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Phe	Pro	Ile	Ala	Thr 85	Tyr	Ala	Phe	Leu	Lys 90	Gly	Ser	Cys	Lys	Trp 95	Ile
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Ser 145	Val	Tyr	Ala	Pro	Tyr 150	Leu	Leu	Ile	Pro	Phe 155	Ile	Leu	Leu	Ile	Phe 160
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Asn	Phe	Ser 35	Asp	Pro	Lys	Pro	Asp 40		Ser	Ser	Ala	Phe 45	Asn	Сув	Leu
Gly	Ala 50	Ala	Asp	Val	Glu	Leu 55	Tyr	Gly	Gly	Gly	Arg 60	Ser	Leu	Glu	Tyr
Leu 65	Leu	Lys	Arg	Val	Asp 70	Thr	Glu	Ala	Asp	Leu 75	Gly	Gln	Phe	Thr	Asp
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Pro	Pro 130	Pro	Leu	Leu	Glu	Ala 135	Thr	Gly	Pro	Asp	Leu 140	Asn	Ile	Leu	Asn
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Ile	Ser 210	Ala	Leu	СЛа	Pro	Leu 215	Lys	Phe	Pro	Thr	Leu 220	Gln	Val	Leu	Ala
Leu	Arg	Asn	Ala	Gly	Met	Glu	Thr	Pro	Ser	Gly	Val	Cys	Ser	Ala	Leu

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						89						aon	tin	uod.	
225					230					235		COII	CIII	uea	240
	Ala	Ala	Arg		Gln	Leu	Gln	Gly			Leu	Ser	His		
Leu	Arq	Asp	Ala	245 Ala	Gly	Ala	Pro	Ser	250 Cys	Asp	Trp	Pro	Ser	255 Gln	Leu
	_	_	260		-			265	-		_		270		
Asn	ser	ьец 275	Asn	ьeu	Ser	Pne	280	GIĀ	Leu	гув	GIN	285	Pro	гув	GIĀ
Leu	Pro 290	Ala	Lys	Leu	Ser	Val 295	Leu	Asp	Leu	Ser	Tyr 300	Asn	Arg	Leu	Asp
Arg 305	Asn	Pro	Ser	Pro	Asp 310	Glu	Leu	Pro	Gln	Val 315	Gly	Asn	Leu	Ser	Leu 320
Lys	Gly	Asn	Pro	Phe 325	Leu	Asp	Ser	Glu	Ser 330	His	Ser	Glu	Lys	Phe 335	Asn
Ser	Gly	Val	Val 340	Thr	Ala	Gly	Ala	Pro 345	Ser	Ser	Gln	Ala	Val 350	Ala	Leu
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His	Lys	Phe 35	Asp	Val	Lys	Gln	Ile 40	Gly	Ala	Gln	Leu	Leu 45	Pro	Pro	Leu
Tyr	Ser 50	Leu	Val	Phe	Ile	Phe 55	Gly	Phe	Val	Gly	Asn 60	Met	Leu	Val	Val
Leu 65	Ile	Leu	Ile	Asn	Cys 70	Lys	Lys	Leu	Lys	Сув 75	Leu	Thr	Asp	Ile	Tyr 80
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Ala 145	Val	Phe	Ala	Leu	Lys 150	Ala	Arg	Thr	Val	Thr 155	Phe	Gly	Val	Val	Thr 160
Ser	Val	Ile	Thr	Trp 165	Leu	Val	Ala	Val	Phe 170	Ala	Ser	Val	Pro	Gly 175	Ile
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Tyr	Phe	Pro 195	Arg	Gly	Trp	Asn	Asn 200	Phe	His	Thr	Ile	Met 205	Arg	Asn	Ile
Leu	Gly 210	Leu	Val	Leu	Pro	Leu 215	Leu	Ile	Met	Val	Ile 220	CAa	Tyr	Ser	Gly
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Ala Val Arg Val Ile Phe Thr Ile Met Ile Val Tyr Phe Leu Phe Trp

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Phe Leu Thr Leu Gly Asn Ile Leu Val Ile Val Gly Ser Ile Ile Met

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Val	Thr	Ile	Ala 100	Ile	Leu	Leu	Phe	Val 105	Tyr	Glu	Gln	Lys	Leu 110	Asn	Thr
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Asn	Ser 130	Thr	Met	Lys	Ala	Trp 135	Asp	Phe	Ile	Gln	Thr 140	Gln	Leu	Gln	Cys
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CÀa	Val	Ile 195	Gln	Val	Leu	Gly	Met 200	Ser	Phe	Ala	Leu	Thr 205	Leu	Asn	Cys
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Tyr	Leu	Leu 35	Ile	His	Asn	Asn	Phe 40	Gly	Val	Leu	Phe	His 45	Asn	Leu	Pro
Ser	Leu 50	Thr	Leu	Gly	Asn	Val 55	Phe	Val	Ile	Val	Gly 60	Ser	Ile	Ile	Met
Val 65	Val	Ala	Phe	Leu	Gly 70	CAa	Met	Gly	Ser	Ile 75	Lys	Glu	Asn	Lys	Cys
Leu	Leu	Met	Ser	Phe 85	Phe	Ile	Leu	Leu	Leu 90	Ile	Ile	Leu	Leu	Ala 95	Glu
Val	Thr	Leu	Ala 100	Ile	Leu	Leu	Phe	Val 105	Tyr	Glu	Gln	Lys	Leu 110	Asn	Glu
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Cys 145	Gly	Ile	Asn	Gly	Thr 150	Ser	Asp	Trp	Thr	Ser 155	Gly	Pro	Pro	Ala	Ser 160
Cys	Pro	Ser	Asp	Arg 165	Lys	Val	Glu	Gly	Cys 170		Ala	Lys	Ala	Arg 175	Leu
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Сла	Val	Ile 195	Glu	Val	Leu	Gly	Met 200	Ser	Phe	Ala	Leu	Thr 205	Leu	Asn	Сув
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n Leu Phe Arg Leu Lys Gl
n $\,$ 90 Gly Glu Arg Phe Val Leu Thr Ser Ile Ile Gln Leu Met Ser Cys Leu 100 105 Cys Val Met Ile Gly Ala Ser Ile Tyr Thr Asp Arg Arg Gln Asp Leu His Gln Gln Asn Arg Lys Leu Tyr Tyr Leu Leu Gln Glu Gly Ser Tyr 135 Gly Tyr Ser Phe Ile Leu Ala Trp Val Ala Phe Ala Phe Thr Phe Ile Ser Gly Leu Met Tyr Met Ile Leu Arg Lys Arg Lys <210> SEQ ID NO 11 <211> LENGTH: 167 <212> TYPE: PRT <213 > ORGANISM: Homo sapiens <400> SEQUENCE: 11 Met Leu Val Leu Leu Ala Phe Ile Ile Ala Phe His Ile Thr Ser Ala 10 Ala Leu Leu Phe Ile Ala Thr Val Asp Asn Ala Trp Trp Val Gly Asp 25 Glu Phe Phe Ala Asp Val Trp Arg Ile Cys Thr Asn Asn Thr Asn Cys 40 Thr Val Ile Asn Asp Ser Phe Gln Glu Tyr Ser Thr Leu Gln Ala Val 55 Gln Ala Thr Met Ile Leu Ser Thr Ile Leu Cys Cys Ile Ala Phe Phe Ile Phe Val Leu Gln Leu Phe Arg Leu Lys Gln Gly Glu Arg Phe Val Leu Thr Ser Ile Ile Gln Leu Met Ser Cys Leu Cys Val Met Ile Ala 100 105 Ala Ser Ile Tyr Thr Asp Arg Glu Asp Ile His Asp Lys Asn Ala Lys Phe Tyr Pro Val Thr Arg Glu Gly Ser Tyr Gly Tyr Ser Tyr Ile 135 140

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360

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	2 > T 3 > OI			Mus	mus	culu	3								
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Ala	Cys 210	Thr	Thr	Pro	His	Val 215	Leu	Lys	Gly	Ile	Pro 220	Leu	Gly	Arg	Leu
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Gly	Arg 290	Ala	Leu	Pro	Gly	Ala 295	Leu	Ala	Thr	Ser	Gly 300	Gln	Pro	Arg	Phe
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Tyr	Gly 2210	Gln	Arg	Pro	ГЛа	Ile 2215	Glu	Arg	Ser	Phe	Leu 2220	Asp	CÀa	Thr
Asn	Arg 2225	Thr	Val	Leu	Val	Ser 2230	Glu	Gly	Ile	Val	Thr 2235	Pro	Arg	Gly
Leu	Ala 2240	Val	Asp	Arg	Ser	Asp 2245	Gly	Tyr	Val	Tyr	Trp 2250	Val	Asp	Asp
Ser	Leu 2255	Asp	Ile	Ile	Ala	Arg 2260	Ile	Arg	Ile	Asn	Gly 2265	Glu	Asn	Ser
Glu	Val 2270	Ile	Arg	Tyr	Gly	Ser 2275	Arg	Tyr	Pro	Thr	Pro 2280	Tyr	Gly	Ile
Thr	Val 2285	Phe	Glu	Asn	Ser	Ile 2290	Ile	Trp	Val	Asp	Arg 2295	Asn	Leu	Lys
ГÀа	Ile 2300	Phe	Gln	Ala	Ser	Lys 2305	Glu	Pro	Glu	Asn	Thr 2310	Glu	Pro	Pro
Thr	Val 2315	Ile	Arg	Asp	Asn	Ile 2320	Asn	Trp	Leu	Arg	Asp 2325	Val	Thr	Ile
Phe	Asp 2330	Lys	Gln	Val	Gln	Pro 2335	Arg	Ser	Pro	Ala	Glu 2340	Val	Asn	Asn
Asn	Pro 2345	CAa	Leu	Glu	Asn	Asn 2350		Gly	Сув	Ser	His 2355	Leu	CAa	Phe
Ala	Leu 2360	Pro	Gly	Leu	His	Thr 2365		Lys	Сув	Asp	Сув 2370	Ala	Phe	Gly
Thr	Leu 2375	Gln	Ser	Asp	Gly	Lys 2380		Cys	Ala	Ile	Ser 2385	Thr	Glu	Asn
Phe	Leu 2390	Ile	Phe	Ala	Leu	Ser 2395		Ser	Leu	Arg	Ser 2400	Leu	His	Leu
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Arg	Thr 2420	Val	Met	Ser	Leu	Asp 2425		Asp	Ser	Val	Ser 2430	Asp	Arg	Ile
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Ala	Glu 2495	Asp	Gly	Ser	Asn	Arg 2500	Thr	Val	Ile	Ala	Arg 2505	Val	Pro	Lys
Pro	Arg 2510	Ala	Ile	Val	Leu	Asp 2515	Pro	Cys	Gln	Gly	Tyr 2520	Leu	Tyr	Trp
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Met	Thr 2630	Thr	Asn	Leu	Leu	Ser 2635	Gln	Pro	Arg	Gly	Ile 2640	Asn	Thr	Val
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Ile	Pro 2795	Arg	Glu	Phe	Ile	Cys 2800	Asn	Gly	Val	Asp	Asn 2805	-	His	Asp
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Glu	Asn 2855	Pro	Thr	Tyr	СЛа	Thr 2860	Thr	His	Thr	СЛа	Ser 2865	Ser	Ser	Glu

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Сув	Gly 2900	His	Ser	Glu	Arg	Thr 2905	Cys	Leu	Ala	Asp	Glu 2910	Phe	Lys	Cys
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Leu	Met 3065	His	Leu	Cys	His	Thr 3070	Pro	Glu	Pro	Thr	Сув 3075	Pro	Pro	His
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	SEQ II														

<211> LENGTH: 339 <212> TYPE: PRT

<213 > ORGANISM: Mus musculus <400> SEQUENCE: 19 Met Ala Gly Cys Cys Ser Val Leu Gly Ser Phe Leu Phe Glu Tyr Asp Arg Val Val Gln Leu Leu Ile Leu Ala Tyr Val Ile Gly Trp Val Phe $35 \ \ \,$ 40 $\ \ \,$ 45 Val Trp Glu Lys Gly Tyr Gln Glu Thr Asp Ser Val Val Ser Ser Val Thr Thr Lys Ala Lys Gly Val Ala Val Thr Asn Thr Ser Gln Leu Gly Phe Arg Ile Trp Asp Val Ala Asp Tyr Val Val Pro Ala Gln Glu Glu Asn Ser Leu Phe Ile Met Thr Asn Met Ile Val Thr Val Asn Gln Thr 105 Gln Gly Thr Cys Pro Glu Ile Pro Asp Lys Thr Ser Ile Cys Asp Ser 120 Asp Ala Asn Cys Thr Leu Gly Ser Ser Asp Thr His Ser Ser Gly Ile 135 Gly Thr Gly Arg Cys Val Pro Phe Asn Ala Ser Val Lys Thr Cys Glu Val Ala Ala Trp Cys Pro Val Glu Asn Asp Ala Gly Val Pro Thr Arg 170 Asn Ile Leu Pro Asn Ile Thr Thr Ser Tyr Leu Lys Ser Cys Ile Tyr 185 Asn Ala Arg Thr Asp Pro Phe Cys Pro Ile Phe Arg Leu Gly Gln Ile Val Ala Asp Ala Gly His Ser Phe Gln Glu Met Ala Val Glu Gly Gly Ile Met Gly Ile Gln Ile Lys Trp Asp Cys Asn Leu Asp Arg Ala Ala Ser His Cys Leu Pro Arg Tyr Ser Phe Arg Arg Leu Asp Thr Arg Asp Leu Glu His Asn Val Ser Pro Gly Tyr Asn Phe Arg Phe Ala Lys Tyr \$260\$Tyr Arg Asp Leu Ala Gly Asn Glu Gln Arg Thr Leu Thr Lys Ala Tyr Gly Ile Arg Phe Asp Ile Ile Val Phe Gly Lys Ala Thr Val Leu Cys 295 Asp Val Ile Val Leu Tyr Cys Met Lys Lys Arg Tyr Tyr Tyr Arg Asp 310 315 Lys Lys Tyr Lys Tyr Val Glu Asp Tyr Glu Gln Gly Leu Ser Gly Glu 330 Met Asn Gln <210> SEQ ID NO 20 <211> LENGTH: 388 <212> TYPE: PRT <213> ORGANISM: Homo sapiens <400> SEQUENCE: 20 Met Ala Gly Cys Cys Ser Ala Leu Ala Ala Phe Leu Phe Glu Tyr Asp

-continued

Thr Pro Arg Ile Val Leu Ile Arg Ser Arg Lys Val Gly Leu Met Asn Arg Ala Val Gln Leu Leu Ile Leu Ala Tyr Val Ile Gly Trp Val Phe Val Trp Glu Lys Gly Tyr Gln Glu Thr Asp Ser Val Val Ser Ser Val Thr Thr Lys Val Lys Gly Val Ala Val Thr Asn Thr Ser Lys Leu Gly 65 $$ 70 $$ 75 $$ 80 Phe Arg Ile Trp Asp Val Ala Asp Tyr Val Ile Pro Ala Gln Glu Glu 90 Asn Ser Leu Phe Val Met Thr Asn Val Ile Leu Thr Met Asn Gln Thr 105 Gln Gly Leu Cys Pro Glu Ile Pro Asp Ala Thr Thr Val Cys Lys Ser 120 Asp Ala Ser Cys Thr Ala Gly Ser Ala Gly Thr His Ser Asn Gly Val 135 Ser Thr Gly Arg Cys Val Ala Phe Asn Gly Ser Val Lys Thr Cys Glu 150 Val Ala Ala Trp Cys Pro Val Glu Asp Asp Thr His Val Pro Gln Pro Ala Phe Leu Lys Ala Ala Glu Asn Phe Thr Leu Leu Val Lys Asn Asn 185 Ile Trp Tyr Pro Lys Phe Asn Phe Ser Lys Arg Asn Ile Leu Pro Asn 200 Ile Thr Thr Thr Tyr Leu Lys Ser Cys Ile Tyr Asp Ala Lys Thr Asp Pro Phe Cys Pro Ile Phe Arg Leu Gly Lys Ile Val Glu Asn Ala Gly His Ser Phe Gln Asp Met Ala Val Glu Gly Gly Ile Met Gly Ile Gln Val Asn Trp Asp Cys Asn Leu Asp Arg Ala Ala Ser Leu Cys Leu Pro Arg Tyr Ser Phe Arg Arg Leu Asp Thr Arg Asp Val Glu His Asn Val Ser Pro Gly Tyr Asn Phe Arg Phe Ala Lys Tyr Tyr Arg Asp Leu Ala 290 295295300 Gly Asn Glu Gln Arg Thr Leu Ile Lys Ala Tyr Gly Ile Arg Phe Asp 310 315 Ile Ile Val Phe Gly Lys Ala Gly Lys Phe Asp Ile Ile Pro Thr Met 325 330 Ile Asn Ile Gly Ser Gly Leu Ala Leu Leu Gly Met Ala Thr Val Leu Cys Asp Ile Ile Val Leu Tyr Cys Met Lys Lys Arg Leu Tyr Tyr Arg 360 Glu Lys Lys Tyr Lys Tyr Val Glu Asp Tyr Glu Gln Gly Leu Ala Ser 375 Glu Leu Asp Gln 385 <210> SEQ ID NO 21 <211> LENGTH: 828 <212> TYPE: PRT <213> ORGANISM: Mus musculus

<400> SEQUENCE: 21

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Val	Ser	Lys	His 20	Leu	Glu	Thr	Ala	Thr 25	Gly	Leu	Asp	Ala	Ser 30	Met	His
Phe	Leu	Ile 35	Met	Glu	Lys	Leu	Gly 40	Arg	Ile	His	Leu	Asn 45	Arg	Gln	Val
Met	Ala 50	Phe	Ile	Phe	Met	Met 55	Val	Leu	Val	Gln	Val 60	Сув	Ser	Glu	Pro
Thr 65	Ile	Arg	Tyr	Ser	Ile 70	Leu	Glu	Glu	Thr	Glu 75	Ser	Gly	Ser	Phe	Val 80
Ala	His	Leu	Ala	Lys 85	Asp	Leu	Gly	Leu	Gly 90	Ala	Arg	Glu	Leu	Ala 95	Ala
Arg	Ser	Ala	Arg 100	Val	Leu	Ser	Asp	Asp 105	Tyr	Lys	Gln	Arg	Leu 110	Leu	Leu
Asp	Pro	Glu 115	Thr	Gly	Asp	Leu	Leu 120	Leu	Arg	Glu	Lys	Val 125	Asp	Arg	Glu
Glu	Val 130	Cys	Ser	Thr	Val	Asp 135	Pro	Cys	Val	Leu	His 140	Phe	Gln	Val	Thr
Leu 145	Glu	Lys	Pro	Val	Gln 150	Tyr	Phe	Gln	Arg	Glu 155	Leu	Leu	Ile	Gln	Asp 160
Ile	Asn	Asp	His	Ala 165	Pro	Glu	Phe	Pro	Asp 170	Arg	Glu	Leu	Leu	Leu 175	Arg
Ile	Pro	Glu	Asn 180	Ser	Gln	Gln	Gly	Thr 185	Gln	Phe	Ser	Leu	Asn 190	Leu	Ala
Gln	Asp	Leu 195	Asp	Val	Gly	Ser	Asn 200	Gly	Leu	Gln	Gln	Tyr 205	Thr	Val	Ser
Pro	Asn 210	Pro	Tyr	Phe	His	Val 215	Leu	Thr	Gln	Asn	Asn 220	Ser	Lys	Gly	Lys
Lys 225	Tyr	Pro	Glu	Leu	Val 230	Gln	Asp	Arg	Gly	Leu 235	Asp	Arg	Glu	Glu	Gln 240
Ala	Glu	Leu	Ser	Leu 245	Thr	Leu	Thr	Ala	Leu 250	Asp	Gly	Gly	Ser	Pro 255	Pro
Arg	Ser	Gly	Thr 260	Ala	Leu	Val	Arg	Ile 265	Leu	Ile	Met	Asp	Ile 270	Asn	Asp
Asn	Ala	Pro 275	Glu	Phe	Val	Asn	Ser 280	Pro	Tyr	Glu	Val	Gln 285	Val	Leu	Glu
Ser	Ser 290	Pro	Pro	Asp	Ser	Pro 295	Val	Leu	Thr	Val	Leu 300	Ala	Arg	Asp	Ala
Asp 305	Ala	Gly	Asn	Phe	Gly 310	Arg	Val	Ser	Tyr	Gly 315	Phe	Phe	Gln	Ala	Ser 320
Asp	Glu	Ile	Gln	Gln 325	Thr	Phe	Ser	Ile	Asn 330	Ala	Thr	Ser	Gly	Asp 335	Met
Arg	Leu	Lys	Lys 340	Lys	Leu	Asp	Phe	Glu 345	ГÀа	Ile	Lys	Ser	Tyr 350	His	Val
Glu	Ile	Glu 355	Ala	Ile	Asp	Gly	Gly 360	Gly	Leu	Ser	Gly	Lys 365	Gly	Ser	Val
Thr	Ile 370	Glu	Val	Val	Asp	Val 375	Asn	Asp	Asn	Ala	Pro 380	Glu	Leu	Thr	Ile
Ser 385	Ser	Leu	Thr	Ser	Ser 390	Val	Pro	Glu	Asn	Ala 395	Pro	Glu	Thr	Ile	Ile 400
Ser	Ile	Phe	Arg	Val 405	Gly	Asp	Arg	Asp	Ser 410	Gly	Glu	Asn	Gly	Lys 415	Met
Val	Сув	Ser	Ile 420	Pro	Glu	Asn	Leu	Pro 425	Phe	Ile	Leu	Lys	Ser 430	Thr	Phe

Asp Ass Thr Pro Ala Phe Thr Gln Thr Ser Tyr Thr Met Phe Val Asp Arg Glu Ass Ass Ser Pro Ala Leu His So5																
Arg Leu Thr Thr Try His Thr Ile Lys Val Gln Val Ser Asp Ile Ass Ass Ass Ass Ass Ass Ass Ass Ass As	Lys	Asn		Tyr	Thr	Leu	Val		Glu	Ser	Pro	Leu	_	Arg	Glu	Ser
465 470 475 486 A89 A80 Thr Pro Ala Phe Thr Gln Thr Ser Tyr Thr Met Phe Val A95 A89 A80 A80 Pro Ala Phe Thr Gln Thr Ser Tyr Thr Met Phe Val A95 A80	Arg		Glu	Tyr	Asn	Ile		Ile	Met	Val	Ser		Met	Gly	Thr	Pro
See Asp See Gly See Asp Ala Leu His Tile Gly Thr Tile See Ala Thr Asp See See Gly See Asp Ala Leu Arg Ar	_	Leu	Thr	Thr	Trp		Thr	Ile	Lys	Val		Val	Ser	Asp	Ile	Asn 480
Ser Asp Ser Gly Ser Asn Ala His The The Tyr Ser Leu Leu Pro Pro Ser Asp Ser Gly Ser Asn Ala His The The Tyr Ser Leu Leu Pro Pro Ser Asn Ala Leu Ser Ser Leu Ile Ser Ile Asn Ala Asp Ser Ser Ser Gly Gln Leu Phe Ala Leu Arg Ala Leu Asp Tyr Glu Ala Leu Gross Ser Ser Gly Gly Ser Pro Ser Ser Ser Gln Ala Leu Val Arg Val Val Val Leu Asp Asp Asn Asp Asp Asn Asp Ser Ser Ser Gln Ala Leu Tyr Pro Met Good Phe Ser Ser Ser Gln Ala Leu Tyr Pro Met Good Phe Ser Ala Pro Phe Val Leu Tyr Pro Met Good Phe	Asp	Asn	Thr	Pro		Phe	Thr	Gln	Thr		Tyr	Thr	Met	Phe		Arg
Signature Sign	Glu	Asn	Asn		Pro	Ala	Leu	His		Gly	Thr	Ile	Ser		Thr	Asp
Asn Gly Gln Leu Phe Ala Leu Arg Ala Leu Asp Tyr Glu Ala Leu Gly Ala Leu Asp Tyr Glu Ala Leu Gly Ala Leu Asp Tyr Glu Ala Leu Gly Ala Leu Asp Asp <td>Ser</td> <td>Asp</td> <td></td> <td>Gly</td> <td>Ser</td> <td>Asn</td> <td>Ala</td> <td></td> <td>Ile</td> <td>Thr</td> <td>Tyr</td> <td>Ser</td> <td></td> <td>Leu</td> <td>Pro</td> <td>Pro</td>	Ser	Asp		Gly	Ser	Asn	Ala		Ile	Thr	Tyr	Ser		Leu	Pro	Pro
545 550 555 566 Val Phe Glu Phe His Ses Val Gly Ala Thr Asp S70 Gly Gly Ser Pro S75 Pro Pro Let S75 Ser Ser Gln Ala Leu Val Arg Val Val Val Val Val Leu Asp Asp Asp Asp Asp S80 Ala Pro Phe Val Leu Tyr Pro Met Gln Asn Ala Ser Ala Pro Phe Thr 595 Ala Pro Phe Val Leu Tyr Pro Met Gln Asn Ala Ser Ala Pro Phe Thr 605 Glu Leu Leu Pro Arg Ala Ala Glu Pro Gly Tyr Leu Val Thr Lys Val 620 Val Ala Val Asp Arg Asp Ser Gly Gln Asn Ala Trp Leu Val Thr Lys Val 640 Ser Phe Glr 646 Leu Leu Lys Ala Thr Glu Pro Gly Leu Phe Ser Val Trp Ala His Asp 645 G45 Ser Gly Glu Asn Ala Trp Leu Ser Phe Glr 640 His Arg Leu Leu Leu Leu Val Val Lys Asp Asp Asp Ser Gly Glu Pro 660 Ser Glu Arg Asp Val Pro Lys 660 Ser Glu Arg Asp Val Pro Lys 660 Tyr Leu Pro 675 Thr Thr Arg Leu Leu Leu Val Asp Gly Phe Ser Gln Arg Ser 660 Ser Glu Arg Asp Val Pro Lys 670 Ala Ser Val Thr Leu Gln Val Lys Asp Asp Asp Asp Gly Phe Ser Gln Arg 660 Ser Mal His Glu Asp Glr 720 Tyr Leu Pro Leu Pro Glu Val Ala Arg Asp Pro Ala His Glu Asp Glr 720 Asp Val Leu Thr Leu Tyr Leu Val Ala Arg Asp Pro Ala His Glu Asp Glr 730 Asp Val Leu Leu Ser Val Leu Leu Pro Gly Gly Cys Ser Met Pro Gly Gly Gly Cys Ser Met Pro Gly Gly Gly Cys Ser Met Pro Gly Gly Gly Gly Cys Ser Met Pro Gly Gly Gly Gly Cys Ser Met Pro Gly Gly Gly Gly Cys Ser Gly Thr Leu Ser Gly 750 Phe Pro Gly His Leu Val Asp Val Cys Leu Arg Gly Asp Ser Gly Thr G	His		Pro	Glu	Leu	Ala		Ser	Ser	Leu	Ile		Ile	Asn	Ala	Asp
Ser Ser Gen Ala Leu Val Arg Val Val Leu Asp Asp <td></td> <td>Gly</td> <td>Gln</td> <td>Leu</td> <td>Phe</td> <td></td> <td>Leu</td> <td>Arg</td> <td>Ala</td> <td>Leu</td> <td></td> <td>Tyr</td> <td>Glu</td> <td>Ala</td> <td>Leu</td> <td>Gln 560</td>		Gly	Gln	Leu	Phe		Leu	Arg	Ala	Leu		Tyr	Glu	Ala	Leu	Gln 560
Ala Pro Phe Val Leu Tyr Pro Met Gln Asn Ala Ser Ala Pro Phe Thr Control Pro Met Gln Asn Ala Ser Ala Pro Phe Thr Pro Phe Thr Control Phe Thr Pro Phe Gln Asn Ala Ser Phe Gln Asn Ala Thr Lys Val Asn	Val	Phe	Glu	Phe		Val	Gly	Ala	Thr		Gly	Gly	Ser	Pro		Leu
Glu Leu Leu Pro Arg Ala Ala Glu Pro Gly Tyr Leu Val Thr Lys Val 625 Val Ala Val Asp Arg Asp Ser Gly Gln Asn Ala Trp Leu Ser Phe Gly Gly Glu Leu Leu Leu Lys Ala Thr Glu Pro Gly Leu Phe Ser Val Trp Ala His Asn 655 Gly Glu Val Arg Thr Thr Arg Leu Leu Ser Gly Glu Arg Asp Val Pro Gly Gly Glu Rrg Asp 670 His Arg Leu Leu Leu Leu Val Val Lys Asp Asn Gly Glu Pro Gln Arg Ser 680 Ala Ser Val Thr Leu Gln Val Leu Leu Val Asp Gly Pro Ala His Glu Asp Gry 700 Tyr Leu Pro Leu Pro Glu Val Ala Arg Asp Val Leu Leu Val Asp Gly Rrg Glu Asp Gly Rrg Gly Asp Asp Ser Gly Asp Asp Asp Asp Ser Gly Asp Gly Rrg Gly Rrg Gly Rrg Gly Rrg Gly Rrg Gly Rrg Gly	Ser	Ser	Gln		Leu	Val	Arg	Val		Val	Leu	Asp	Asp		Asp	Asn
610 615 620 Val Ala Val Asp Arg Asp Ser Gly Gln Asn Ala Trp Leu Ser Phe Gld Leu Leu Lys Ala Thr Glu Pro Gly Leu Phe Ser Val Trp Ala His Asg Asg Leu L	Ala	Pro		Val	Leu	Tyr	Pro		Gln	Asn	Ala	Ser		Pro	Phe	Thr
625	Glu		Leu	Pro	Arg	Ala		Glu	Pro	Gly	Tyr		Val	Thr	ГÀа	Val
Gly Glu Val Arg Thr Thr Arg Leu Leu Ser Glu Arg Asp Val Pro Lys 680		Ala	Val	Asp	Arg		Ser	Gly	Gln	Asn		Trp	Leu	Ser	Phe	Gln 640
His Arg Leu Leu Leu Val Val Lys Asp Asn Gly Glu Pro Gln Arg Ser 670 Ala Ser Val Thr Leu Gln Val Leu Val Asp Gly Phe Ser Gln Ser 690 Tyr Leu Pro Leu Pro Glu Val Ala Arg Asp Pro Ala His Glu Asp Gly 720 Asp Val Leu Thr Leu Tyr Leu Val Ile Ala Leu Ala Ser Val Ser Ser 720 Leu Phe Leu Leu Ser Val Leu Leu Phe Val Gly Val Arg Leu Cys Arg 745 Arg Ala Arg Glu Val Ser Leu Gly Gly Cys Ser Met Pro Gly Glu His 755 Phe Pro Gly His Leu Val Asp Val Ser Gly Ala Gly Thr Leu Ser Gln Ser 785 Ser Tyr Gln Tyr Glu Val Cys Leu Arg Gly Asp Ser Gly Thr Gly Gly Gly Rep 795	Leu	Leu	Lys	Ala		Glu	Pro	Gly	Leu		Ser	Val	Trp	Ala		Asn
Ala Ser Val Thr Leu Gln Val Leu Leu Val Asp Gly Phe Ser Gln Ser G90 Val Thr Leu Gln Val Ala Arg Asp Pro Ala His Glu Asp Glr 705 Asp Val Leu Thr Leu Tyr Leu Val Ile Ala Leu Ala Ser Val Ser Ser 725 Leu Phe Leu Leu Ser Val Leu Leu Phe Val Gly Val Arg Leu Cys Arg 745 Arg Ala Arg Glu Val Ser Leu Gly Gly Cys Ser Met Pro Gly Glu His 755 Phe Pro Gly His Leu Val Asp Val Ser Gly Ala Gly Thr Gly Gly Ser Tyr Gln Tyr Glu Val Cys Leu Arg Gly Asp Ser Gly Thr Gly Gly Gly Gly Cys Ser Tyr Gln Tyr Glu Val Cys Leu Arg Gly Asp Ser Gly Thr Gly	Gly	Glu	Val		Thr	Thr	Arg	Leu		Ser	Glu	Arg	Asp		Pro	Lys
Tyr Leu Pro Leu Pro Glu Val Ala Arg Asp Pro Ala His Glu Asp Glu 720 Asp Val Leu Pro Leu Yal Leu Val Leu Val Leu Ala Leu Ser Val Ser Ser Ser Arg Ala Arg Leu Leu Phe Val Gly Val Arg Leu Cys Arg Arg Arg Fro Gly Gly Cys Ser Met Pro Gly Glu His Fro Gly Fro Ala Gly Fro Gly Fro Gly Fro Fro Gly Gly Fro Fro Gly Fro Gly Fro Fro Gly Fro Fro Gly Fro Fro Gly Fro Gly Fro Fro Fro Gly F	His	Arg		Leu	Leu	Val	Val		Asp	Asn	Gly	Glu		Gln	Arg	Ser
Asp Val Leu Thr Leu Tyr Leu Val Ile Ala Leu Ala Ser Val Ser 735 Leu Phe Leu Leu Ser Val Leu Leu Phe Val Gly Val Arg Leu Cys Arg 745 Arg Ala Arg Glu Val Ser Leu Gly 760 Gly Cys Ser Met 765 Pro Gly Gly Glu His Leu Val Asp 775 Phe Pro Gly His Leu Val Asp 775 Val Ser Gly Ala Gly Thr Leu Ser Gly 780 Ser Tyr Gln Tyr Glu Val Cys Leu Arg Gly Asp 795 Ser Gly Thr Gly Gly 618	Ala		Val	Thr	Leu	Gln		Leu	Leu	Val	Asp	_	Phe	Ser	Gln	Ser
Leu Phe Leu Leu Ser 740 Val Leu Leu Leu Phe 745 Val Gly Val Arg Leu Cys Arg 750 Arg Ala Arg 755 Glu Val Ser Leu Gly Gly Cys Ser Met 765 Pro Gly His Leu Val Asp Val Ser Gly Ala Gly Thr Leu Ser Gly 770 Ser Tyr Gln Tyr Glu Val Cys Leu Arg Gly Asp 795 Ser Gly Asp 795		Leu	Pro	Leu	Pro		Val	Ala	Arg	Asp		Ala	His	Glu	Asp	Glu 720
Arg Ala Arg Glu Val Ser Leu Gly Gly Cys Ser Met 755 Pro Gly Glu His Leu Val Asp Val Ser Gly Ala Gly Thr Leu Ser Glu 770 Ser Tyr Gln Tyr Glu Val Cys Leu Arg Gly Asp 795 Ser Gly Asp 795	Asp	Val	Leu	Thr		Tyr	Leu	Val	Ile		Leu	Ala	Ser	Val		Ser
755 760 765 Phe Pro Gly His Leu Val Asp Val Ser Gly Ala Gly Thr Leu Ser Glr 770 780 Ser Tyr Gln Tyr Glu Val Cys Leu Arg Gly Asp Ser Gly Thr Gly Glr 785 790 795 800	Leu	Phe	Leu		Ser	Val	Leu	Leu		Val	Gly	Val	Arg		Cys	Arg
770 775 780 Ser Tyr Gln Tyr Glu Val Cys Leu Arg Gly Asp Ser Gly Thr Gly Glu 785 790 795 800	Arg	Ala	_	Glu	Val	Ser	Leu	_	Gly	CÀa	Ser	Met		Gly	Glu	His
785 790 795 800	Phe		Gly	His	Leu	Val	_	Val	Ser	Gly	Ala		Thr	Leu	Ser	Gln
Phe Lys Phe Leu Lys Pro Met Ile Pro Asn Ala Gly Ile Gly Ile Met		Tyr	Gln	Tyr	Glu		CAa	Leu	Arg	Gly		Ser	Gly	Thr	Gly	Glu 800
805 810 815	Phe	Lys	Phe	Leu		Pro	Met	Ile	Pro		Ala	Gly	Ile	Glu		Met
Glu Ser Pro His Cys Arg Asp Ser Phe Val Phe Asn 820 825	Glu	Ser	Pro		Cys	Arg	Asp	Ser		Val	Phe	Asn				

<210> SEQ ID NO 22 <211> LENGTH: 796 <212> TYPE: PRT

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< 400	0 > S	EQUEI	NCE:	22											
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Phe	Leu	Phe	Trp 20	Gly	Val	Ser	Leu	Ala 25	Gly	Ser	Gly	Phe	Gly 30	Arg	Tyr
Ser	Val	Thr 35	Glu	Glu	Thr	Glu	Lys 40	Gly	Ser	Phe	Val	Val 45	Asn	Leu	Ala
ГÀЗ	Asp 50	Leu	Gly	Leu	Ala	Glu 55	Gly	Glu	Leu	Ala	Ala 60	Arg	Gly	Thr	Arg
Val 65	Val	Ser	Asp	Asp	Asn 70	Lys	Gln	Tyr	Leu	Leu 75	Leu	Asp	Ser	His	Thr 80
Gly	Asn	Leu	Leu	Thr 85	Asn	Glu	Lys	Leu	Asp 90	Arg	Glu	Lys	Leu	Сув 95	Gly
Pro	Lys	Glu	Pro 100	CÀa	Met	Leu	Tyr	Phe 105	Gln	Ile	Leu	Met	Asp 110	Asp	Pro
Phe	Gln	Ile 115	Tyr	Arg	Ala	Glu	Leu 120	Arg	Val	Arg	Asp	Ile 125	Asn	Asp	His
Ser	Pro 130	Val	Phe	Arg	His	Lуs 135	Glu	Met	Val	Leu	Lys 140	Ile	Ser	Glu	Asn
Thr 145	Ala	Glu	Gly	Thr	Ala 150	Phe	Arg	Leu	Glu	Arg 155	Ala	Gln	Asp	Pro	Asp 160
Glu	Gly	His	Asn	Ser 165	Ile	Gln	Asn	Tyr	Thr 170	Ile	Ser	Ser	Asn	Ser 175	Phe
Phe	His	Ile	Lys 180	Ile	Ser	Gly	Ser	Asp 185	Glu	Gly	Met	Ile	Tyr 190	Pro	Glu
Leu	Val	Leu 195	Asp	ГÀа	Ala	Leu	Asp 200	Arg	Glu	Glu	Gln	Glu 205	Glu	Leu	Ser
Leu	Thr 210	Leu	Thr	Ala	Leu	Asp 215	Gly	Gly	Ser	Pro	Ser 220	Arg	Ser	Gly	Thr
Ser 225	Thr	Ile	Arg	Ile	Val 230	Val	Leu	Asp	Val	Asn 235	Asp	Asn	Ala	Pro	Gln 240
Phe	Ala	Gln	Ala	Leu 245	Tyr	Glu	Thr	Gln	Ala 250	Pro	Glu	Asn	Ser	Pro 255	Val
Gly	Ser	Leu	Ile 260	Val	Lys	Val	Ser	Ala 265	Gly	Asp	Ala	Asp	Ser 270	Gly	Val
Asn	Ala	Glu 275	Val	Ser	Tyr	Ser	Phe 280	Phe	Asp	Ala	Ser	Glu 285	Asp	Ile	Leu
Thr	Thr 290	Phe	Gln	Ile	Asn	Pro 295	Phe	Ser	Gly	Glu	Ile 300	Phe	Leu	Arg	Glu
Leu 305	Leu	Asp	Tyr	Glu	Leu 310	Val	Asn	Ser	Tyr	Lys 315	Ile	Asn	Ile	Gln	Ala 320
Met	Asp	Gly	Gly	Gly 325	Leu	Ser	Ala	Arg	330	Thr	Val	Leu	Ile	Lys 335	Val
Leu	Asp	Ser	Asn 340	Asp	Asn	Pro	Pro	Glu 345	Leu	Ile	Ile	Ser	Ser 350	Leu	Ser
Asn	Ser	Val 355	Ala	Glu	Asn	Ser	Pro 360	Gly	Ile	Val	Leu	Ala 365	Val	Phe	ГÀа
Ile	Lys 370	Asp	Arg	Asp	Ser	Gly 375	Glu	Asn	Gly	ГÀа	Thr 380	Ile	Cys	Tyr	Val
Gln 385	Asp	Asn	Leu	Pro	Phe 390	Phe	Leu	Lys	Pro	Ser 395	Val	Asp	Asn	Phe	Tyr 400
Ile	Leu	Met	Thr	Glu	Gly	Ala	Leu	Asp	Arg	Glu	Ser	Lys	Ala	Glu	Tyr

												0011	C III.	aca	
				405					410					415	
Asn	Ile	Thr	Ile 420	Thr	Val	Thr	Asp	Leu 425	Gly	Thr	Pro	Arg	Leu 430	Lys	Thr
Glu	His	Ser 435	Ile	Thr	Leu	Gln	Val 440	Ser	Asp	Val	Asn	Asp 445	Asn	Ala	Pro
Ala	Phe 450	Thr	Gln	Thr	Ser	Tyr 455	Thr	Leu	Phe	Val	Arg 460	Glu	Asn	Asn	Ser
Pro 465	Ala	Leu	His	Ile	Gly 470	Ser	Val	Ser	Ala	Thr 475	Asp	Arg	Asp	Ser	Gly 480
Thr	Asn	Ala	Gln	Val 485	Thr	Tyr	Ser	Leu	Leu 490	Pro	Pro	Gln	Asp	Pro 495	His
Leu	Pro	Leu	Ala 500	Ser	Leu	Val	Ser	Ile 505	Asn	Ala	Asp	Asn	Gly 510	His	Leu
Phe	Ala	Leu 515	Arg	Ser	Leu	Asp	Tyr 520	Glu	Ala	Leu	Gln	Ala 525	Phe	Asp	Phe
Arg	Val 530	Gly	Ala	Ser	Asp	Arg 535	Gly	Ser	Pro	Ala	Leu 540	Ser	Ser	Glu	Ala
Leu 545	Val	Arg	Val	Leu	Val 550	Leu	Asp	Ala	Asn	Asp 555	Asn	Ser	Pro	Phe	Val 560
Leu	Tyr	Pro	Leu	Gln 565	Asn	Gly	Ser	Ala	Pro 570	Cya	Thr	Glu	Leu	Val 575	Pro
Arg	Ala	Ala	Glu 580	Pro	Gly	Tyr	Leu	Val 585	Thr	Lys	Val	Val	Ala 590	Val	Asp
Gly	Asp	Ser 595	Gly	Gln	Asn	Ala	Trp 600	Leu	Ser	Tyr	Gln	Leu 605	Leu	Lys	Ala
Thr	Glu 610	Pro	Gly	Leu	Phe	Gly 615	Val	Trp	Ala	His	Asn 620	Gly	Glu	Val	Arg
Thr 625	Ala	Arg	Leu	Leu	Ser 630	Glu	Arg	Asp	Ala	Ala 635	ГÀз	His	Arg	Leu	Val 640
Val	Leu	Val	Lys	Asp 645	Asn	Gly	Glu	Pro	Pro 650	Arg	Ser	Ala	Thr	Ala 655	Thr
Leu	His	Val	Leu 660	Leu	Val	Asp	Gly	Phe 665	Ser	Gln	Pro	Tyr	Leu 670	Pro	Leu
Pro	Glu	Ala 675	Ala	Pro	Ala	Gln	Ala 680	Gln	Ala	Asp	Leu	Leu 685	Thr	Val	Tyr
Pro	Val 690	Val	Ala	Leu	Ala	Ser 695	Val	Ser	Ser	Leu	Phe 700	Leu	Leu	Ser	Val
Leu 705	Leu	Phe	Val	Ala	Val 710	Arg	Leu	Сув	Arg	Arg 715	Ser	Arg	Ala	Ala	Ser 720
Val	Gly	Arg	Cys	Ser 725	Val	Pro	Glu	Gly	Pro 730	Phe	Pro	Gly	His	Leu 735	Val
Asp	Val	Ser	Gly 740	Thr	Gly	Thr	Leu	Phe 745	Gln	Ser	Tyr	Gln	Tyr 750	Glu	Val
Cys	Leu	Thr 755	Gly	Gly	Ser	Glu	Thr 760	Gly	Glu	Phe	Lys	Phe 765	Leu	Lys	Pro
Ile	Thr 770	Pro	His	Leu	Pro	Pro 775	His	Arg	Gly	Gly	Lys 780	Glu	Ile	Glu	Glu
Asn 785	Ser	Thr	Leu	Pro	Asn 790	Ser	Phe	Gly	Phe	Asn 795	Tyr				

<210> SEQ ID NO 23 <211> LENGTH: 699 <212> TYPE: PRT <213> ORGANISM: Mus musculus

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< 400)> SI	EQUEI	ICE :	23											
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Ala	Arg	Ala	Gly 20	Gln	Gly	Asn	Asp	Thr 25	Thr	Pro	Thr	Glu	Ser 30	Asn	Trp
Thr	Ser	Thr 35	Thr	Ala	Gly	Pro	Pro 40	Asp	Pro	Gly	Ala	Ser 45	Gln	Pro	Leu
Leu	Thr 50	Trp	Leu	Leu	Leu	Pro 55	Leu	Leu	Leu	Leu	Leu 60	Phe	Leu	Leu	Ala
Ala 65	Tyr	Phe	Phe	Arg	Phe 70	Arg	Lys	Gln	Arg	Lys 75	Ala	Val	Val	Ser	Ser 80
Asn	Asp	Lys	Lys	Met 85	Pro	Asn	Gly	Ile	Leu 90	Glu	Glu	Gln	Glu	Gln 95	Gln
Arg	Val	Met	Leu 100	Leu	Ser	Arg	Ser	Pro 105	Ser	Gly	Pro	Lys	Lys 110	Phe	Phe
Pro	Ile	Pro 115	Val	Glu	His	Leu	Glu 120	Glu	Glu	Ile	Arg	Val 125	Arg	Ser	Ala
Asp	Asp 130	Cys	Lys	Arg	Phe	Arg 135	Glu	Glu	Phe	Asn	Ser 140	Leu	Pro	Ser	Gly
His 145	Ile	Gln	Gly	Thr	Phe 150	Glu	Leu	Ala	Asn	155	Glu	Glu	Asn	Arg	Glu 160
ГÀа	Asn	Arg	Tyr	Pro 165	Asn	Ile	Leu	Pro	Asn 170	Asp	His	CÀa	Arg	Val 175	Ile
Leu	Ser	Gln	Val 180	Asp	Gly	Ile	Pro	Сув 185	Ser	Asp	Tyr	Ile	Asn 190	Ala	Ser
Tyr	Ile	Asp 195	Gly	Tyr	Lys	Glu	Lys 200	Asn	Lys	Phe	Ile	Ala 205	Ala	Gln	Gly
Pro	Lys 210	Gln	Glu	Thr	Val	Asn 215	Asp	Phe	Trp	Arg	Met 220	Val	Trp	Glu	Gln
Arg 225	Ser	Ala	Thr	Ile	Val 230	Met	Leu	Thr	Asn	Leu 235	ГÀа	Glu	Arg	Lys	Glu 240
Glu	Lys	Cys	Tyr	Gln 245	Tyr	Trp	Pro	Asp	Gln 250	Gly	CAa	Trp	Thr	Tyr 255	Gly
Asn	Ile	Arg	Val 260	Cys	Val	Glu	Asp	Сув 265	Val	Val	Leu	Val	Asp 270	Tyr	Thr
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Val 305	Pro	Phe	Thr	Pro	Ile 310	Gly	Met	Leu	Lys	Phe 315	Leu	Lys	Lys	Val	Lys 320
Thr	Leu	Asn	Pro	Ser 325	His	Ala	Gly	Pro	Ile 330	Val	Val	His	CÀa	Ser 335	Ala
Gly	Val	Gly	Arg 340	Thr	Gly	Thr	Phe	Ile 345	Val	Ile	Asp	Ala	Met 350	Met	Asp
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Ile	Arg 370	Asn	Gln	Arg	Pro	Gln 375	Met	Val	Gln	Thr	380	Val	Gln	Tyr	Thr
Phe 385	Ile	Tyr	Gln	Ala	Leu 390	Leu	Glu	Tyr	Tyr	Leu 395	Tyr	Gly	Asp	Thr	Glu 400
Leu	Asp	Val	Ser	Ser 405	Leu	Glu	Arg	His	Leu 410	Gln	Thr	Leu	His	Ser 415	Thr

Ala Thr His Phe Asp Lys Ile Gly Leu Glu Glu Glu Phe Arg Lys Leu Thr Asn Val Arg Ile Met Lys Glu Asn Met Arg Thr Gly Asn Leu Pro Ala Asn Met Lys Lys Ala Arg Val Ile Gln Ile Ile Pro Tyr Asp Phe Asn Arg Val Ile Leu Ser Met Lys Arg Gly Gln Glu Phe Thr Asp Tyr Ile Asn Ala Ser Phe Ile Asp Gly Tyr Arg Gln Lys Asp Tyr Phe Met 485 Ala Thr Gln Gly Pro Leu Ala His Thr Gly Glu Asp Phe Trp Arg Met 505 Val Trp Glu Trp Lys Ser His Thr Ile Val Met Leu Thr Glu Val Gln 520 Glu Arg Glu Gln Asp Lys Cys Tyr Gln Tyr Trp Pro Thr Glu Gly Ser 535 Val Thr His Gly Asp Ile Thr Ile Glu Ile Lys Ser Asp Thr Leu Ser 550 Glu Ala Ile Ser Val Arg Asp Phe Leu Val Thr Phe Lys Gln Pro Leu Ala Arg Gln Glu Glu Gln Val Arg Met Val Arg Gln Phe His Phe His 585 Gly Trp Pro Glu Val Gly Ile Pro Ala Glu Gly Lys Gly Ile Ile Asp 600 Leu Ile Ala Ala Val Gln Lys Gln Gln Gln Gln Thr Gly Asn His Pro Ile Thr Val His Cys Ser Ala Gly Ala Gly Arg Thr Gly Thr Phe Ile Ala Leu Ser Asn Ile Leu Glu Arg Val Lys Ala Glu Gly Leu Leu Asp Val Phe Gln Ala Val Lys Ser Leu Arg Leu Gln Arg Pro His Met Val Gln Thr Leu Glu Gln Tyr Glu Phe Cys Tyr Lys Val Val Gln Asp Phe Ile Asp Ile Phe Ser Asp Tyr Ala Asn Phe Lys <210> SEQ ID NO 24 <211> LENGTH: 642 <212> TYPE: PRT <213 > ORGANISM: Homo sapiens <400> SEQUENCE: 24 Met Ser Asn Arg Ser Ser Phe Ser Arg Leu Thr Trp Phe Arg Lys Gln 10 Arg Lys Ala Val Val Ser Thr Ser Asp Lys Lys Met Pro Asn Gly Ile 25 Leu Glu Glu Glu Gln Gln Arg Val Met Leu Leu Ser Arg Ser Pro Ser Gly Pro Lys Lys Tyr Phe Pro Ile Pro Val Glu His Leu Glu Glu 55 Glu Ile Arg Ile Arg Ser Ala Asp Asp Cys Lys Gln Phe Arg Glu Glu 70 Phe Asn Ser Leu Pro Ser Gly His Ile Gln Gly Thr Phe Glu Leu Ala

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Asn	Asp	His 115	Ser	Arg	Val	Ile	Leu 120	Ser	Gln	Leu	Asp	Gly 125	Ile	Pro	Сув
Ser	Asp 130	Tyr	Ile	Asn	Ala	Ser 135	Tyr	Ile	Asp	Gly	Tyr 140	Lys	Glu	Lys	Asn
Lys 145	Phe	Ile	Ala	Ala	Gln 150	Gly	Pro	Lys	Gln	Glu 155	Thr	Val	Asn	Asp	Phe 160
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Gln	Gly	Сув 195	Trp	Thr	Tyr	Gly	Asn 200	Ile	Arg	Val	Cys	Val 205	Glu	Asp	Cys
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Ile	Val	Val 275	His	CAa	Ser	Ala	Gly 280	Val	Gly	Arg	Thr	Gly 285	Thr	Phe	Ile
Val	Ile 290	Asp	Ala	Met	Met	Ala 295	Met	Met	His	Ala	Glu 300	Gln	Lys	Val	Asp
Val 305	Phe	Glu	Phe	Val	Ser 310	Arg	Ile	Arg	Asn	Gln 315	Arg	Pro	Gln	Met	Val 320
Gln	Thr	Asp	Met	Gln 325	Tyr	Thr	Phe	Ile	Tyr 330	Gln	Ala	Leu	Leu	Glu 335	Tyr
Tyr	Leu	Tyr	Gly 340	Asp	Thr	Glu	Leu	Asp 345	Val	Ser	Ser	Leu	Glu 350	Lys	His
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Gln	Ile	Ile	Pro	Tyr 405	Asp	Phe	Asn	Arg	Val 410	Ile	Leu	Ser	Met	Lys 415	Arg
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Arg	Gln	Lys 435	Asp	Tyr	Phe	Ile	Ala 440	Thr	Gln	Gly	Pro	Leu 445	Ala	His	Thr
Val	Glu 450	Asp	Phe	Trp	Arg	Met 455	Ile	Trp	Glu	Trp	Lys 460	Ser	His	Thr	Ile
Val 465	Met	Leu	Thr	Glu	Val 470	Gln	Glu	Arg	Glu	Gln 475	Asp	Lys	Cha	Tyr	Gln 480
Tyr	Trp	Pro	Thr	Glu 485	Gly	Ser	Val	Thr	His 490	Gly	Glu	Ile	Thr	Ile 495	Glu
Ile	Lys	Asn	Asp 500	Thr	Leu	Ser	Glu	Ala 505	Ile	Ser	Ile	Arg	Asp 510	Phe	Leu
Val	Thr	Leu 515	Asn	Gln	Pro	Gln	Ala 520	Arg	Gln	Glu	Glu	Gln 525	Val	Arg	Val

Val	Arg 530	Gln	Phe	His	Phe	His 535	Gly	Trp	Pro	Glu	Ile 540	Gly	Ile	Pro	Ala
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Lys	Ala	Glu 595	Gly	Leu	Leu	Asp	Val 600	Phe	Gln	Ala	Val	Lys 605	Ser	Leu	Arg
Leu	Gln 610	Arg	Pro	His	Met	Val 615	Gln	Thr	Leu	Glu	Gln 620	Tyr	Glu	Phe	Сув
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Glu	Glu	Asp 35	Asp	Asp	Leu	Gly	Lys 40	Thr	Leu	Ala	Val	Ser 45	Arg	Phe	Gly
Asp	Leu 50	Ile	Ser	Lys	Thr	Pro 55	Ala	Trp	Asp	Pro	Glu 60	Lys	Pro	Ser	Arg
Ser 65	Tyr	Ser	Glu	Arg	Asp 70	Phe	Glu	Phe	His	Arg 75	His	Thr	Ser	His	His 80
Thr	His	His	Pro	Leu 85	Ser	Ala	Arg	Leu	Pro 90	Pro	Pro	His	ГÀз	Leu 95	Arg
Arg	Pro	Pro	Pro 100	Thr	Ser	Ala	Arg	His 105	Thr	Arg	Arg	Lys	Arg 110	ГÀа	Lys
Glu	Lys	Thr 115	Ser	Ala	Pro	Pro	Ser 120	Glu	Gly	Thr	Pro	Pro 125	Ile	Gln	Glu
Glu	Gly 130	Gly	Ala	Gly	Ala	Glu 135	Glu	Glu	Glu	Glu	Glu 140	Glu	Glu	Glu	Glu
Glu 145	Gly	Glu	Ser	Glu	Ala 150	Glu	Pro	Val	Glu	Pro 155	Leu	Pro	Pro	Gly	Pro 160
Pro	Gln	Lys	Ala	Lys 165	Phe	Ser	Ile	Gly	Ser 170	Asp	Glu	Asp	Asp	Ser 175	Pro
Gly	Leu	Pro	Val 180	Lys	Ala	Pro	Cys	Ala 185	Lys	Ala	Leu	Pro	Ser 190	Val	Gly
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Arg	Ala 210	Arg	Ala	Ser	Arg	Ile 215	Ser	Thr	Glu	ГЛа	Ser 220	Arg	Pro	Trp	Ser
Pro 225	Ser	Ala	Ser	Tyr	Asp 230	Leu	Arg	Glu	Arg	Leu 235	CAa	Pro	Gly	Ser	Ala 240
Leu	Gly	Asn	Pro	Gly 245	Pro	Glu	Gln	Arg	Val 250	Pro	Thr	Asp	Glu	Ala 255	Glu

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Arg	Ile 290	Gln	Gly	Gly	Arg	Gly 295	Ser	Pro	Ser	Gly	Leu 300	Ala	Pro	Ile	Leu
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Phe	Val	Glu	Leu	Asn 325	Glu	Leu	Met	Leu	Asp 330	Arg	Ser	Gln	Glu	Pro 335	His
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Glu	Thr	Glu 355	Arg	Trp	Gly	ГÀа	Pro 360	His	Val	Ala	Ser	Leu 365	Ser	Phe	Arg
Ser	Leu 370	Leu	Glu	Leu	Arg	Arg 375	Thr	Ile	Ala	Gln	Gly 380	Ala	Ala	Leu	Leu
Asp 385	Leu	Glu	Gln	Thr	Thr 390	Leu	Pro	Gly	Ile	Ala 395	His	Leu	Val	Val	Glu 400
Thr	Met	Ile	Val	Ser 405	Asp	Gln	Ile	Arg	Pro 410	Glu	Asp	Arg	Ala	Ser 415	Val
Leu	Arg	Thr	Leu 420	Leu	Leu	Lys	His	Ser 425	His	Pro	Asn	Asp	Asp 430	Lys	Asp
Ser	Gly	Phe 435	Phe	Pro	Arg	Asn	Pro 440	Ser	Ser	Ser	Ser	Val 445	Asn	Ser	Val
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Pro 465	Thr	Met	Ala	Asp	Asp 470	Gln	Gly	Glu	Pro	Ala 475	Pro	Leu	Trp	Pro	His 480
Asp	Pro	Asp	Ala	Lys 485	Glu	ГÀЗ	Pro	Leu	His 490	Met	Pro	Gly	Gly	Asp 495	Gly
	_	-	500		Leu	=		505		-			510	_	
		515			Leu		520	=				525			
	530				Arg	535					540				
Leu 545	Glu	Val	Pro	Val	Pro 550	Val	Arg	Phe	Leu	Phe 555	Val	Met	Leu	Gly	Pro 560
Ser	His	Thr	Ser	Thr 565	Asp	Tyr	His	Glu	Leu 570	Gly	Arg	Ser	Ile	Ala 575	Thr
Leu	Met	Ser	Asp 580	ГÀа	Leu	Phe	His	Glu 585	Ala	Ala	Tyr	Gln	Ala 590	Asp	Asp
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Val	Ile 610	Pro	Pro	Ser	Glu	Val 615	Glu	Gly	Arg	Asp	Leu 620	Leu	Arg	Ser	Val
Ala 625	Ala	Phe	Gln	Arg	Glu 630	Leu	Leu	Arg	Lys	Arg 635	Arg	Glu	Arg	Glu	Gln 640
Thr	Lys	Val	Glu	Met 645	Thr	Thr	Arg	Gly	Gly 650	Tyr	Ala	Ala	Pro	Gly 655	ГÀа
Glu	Leu	Ser	Leu 660	Glu	Met	Gly	Gly	Ser 665	Glu	Ala	Thr	Ser	Glu 670	Asp	Asp
Pro	Leu	Gln 675	Arg	Thr	Gly	Ser	Val 680	Phe	Gly	Gly	Leu	Val 685	Arg	Asp	Val

ГЛа	Arg 690	Arg	Tyr	Pro	His	Tyr 695	Pro	Ser	Asp	Leu	Arg 700	Asp	Ala	Leu	His
Ser 705	Gln	Cys	Val	Ala	Ala 710	Val	Leu	Phe	Ile	Tyr 715	Phe	Ala	ı Ala	Leu	Ser 720
Pro	Ala	Ile	Thr	Phe 725	Gly	Gly	Leu	Leu	Gly 730	Glu	Lys	Thr	Glu	Gly 735	Leu
Met	Gly	Val	Ser 740	Glu	Leu	Ile	Val	Ser 745	Thr	Ala	Val	Leu	Gly 750		Leu
Phe	Ser	Leu 755	Leu	Gly	Ala	Gln	Pro 760	Leu	Leu	Val	Val	Gly 765		Ser	Gly
Pro	Leu 770	Leu	Val	Phe	Glu	Glu 775	Ala	Phe	Phe	Lys	Phe 780	Суя	Arg	Ala	Gln
Asp 785	Leu	Glu	Tyr	Leu	Thr 790	Gly	Arg	Val	Trp	Val 795	Gly	Leu	Trp	Leu	Val 800
Val	Phe	Val	Leu	Ala 805	Leu	Val	Ala	Ala	Glu 810	Gly	Thr	Phe	e Leu	Val 815	Arg
Tyr	Ile	Ser	Pro 820	Phe	Thr	Gln	Glu	Ile 825	Phe	Ala	Phe	Leu	Ile 830	Ser	Leu
Ile	Phe	Ile 835	Tyr	Glu	Thr	Phe	His 840	Lys	Leu	Tyr	Lys	Val 845		Thr	Glu
His	Pro 850	Leu	Leu	Pro	Phe	Tyr 855	Pro	Pro	Asp	Glu	Ala 860	Leu	ı Glu	Thr	Gly
Leu 865	Glu	Leu	Asn	Ser	Ser 870	Ala	Leu	Pro	Pro	Thr 875	Glu	Gly	Pro	Pro	Gly 880
Pro	Arg	Asn	Gln	Pro 885	Asn	Thr	Ala	Leu	Leu 890	Ser	Leu	Ile	e Leu	Met 895	Leu
Gly	Thr	Phe	Leu 900	Ile	Ala	Phe	Phe	Leu 905	Arg	Lys	Phe	Arg	Asn 910	Ser	Arg
Phe	Leu	Gly 915	Gly	Lys	Ala	Arg	Arg 920	Ile	Ile	Gly	Asp	Phe 925	Gly	Ile	Pro
Ile	Ser 930	Ile	Leu	Val	Met	Val 935	Leu	Val	Asp	Tyr	Ser 940	Ile	Thr	Asp	Thr
Tyr 945	Thr	Gln	Lys	Leu	Thr 950	Val	Pro	Thr	Gly	Leu 955	Ser	Val	Thr	Ser	Pro 960
His	Lys	Arg	Thr	Trp 965	Phe	Ile	Pro	Pro	Leu 970	Gly	Ser	Ala	Arg	Pro 975	
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Ile	Leu	Ile 995	Phe	Met	Glu	Thr	Gln 1000		e Thi	r Ala	a Le		.e V 005	al S	er Gln
ГÀа	Ala 1010		g Arc	g Lev	ı Lev	101		Ly Se	er G	ly Pi		is 020	Leu	Asp	Leu
Leu	Leu 1025		e Gly	/ Sei	. Leu	103		ly Le	eu Cy	ys G:		eu 035	Phe	Gly	Leu
Pro	Trp 1040		ı Thr	Ala	a Ala	104		al Ai	rg Se	er Va		hr 050	His	Val	Asn
Ala	Leu 1055		r Val	L Met	Arç	106		la I	le A	la P:		ly 065	Asp	Lys	Pro
Gln	Ile 1070		n Glu	ı Val	l Arg	Glu 10		ln Ai	rg Va	al Ti		ly 080	Val	Leu	Ile
Ala	Ser 1085		ı Val	l Gly	/ Let	Ser 109		Le Va	al Me	et Gi	_	la 095	Val	Leu	Arg
Arg	Ile	Pro) Lev	ı Ala	a Val	. Le	ı Pl	ne G	ly I	le Pl	ne L	eu	Tyr	Met	Gly

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Asp Leu Ile Set	r Lys Pro Pro Ala 55	Trp Asp Pro Glu 1	Lys Pro Ser Arg
Ser Tyr Ser Glu	ı Arg Asp Phe Glu 70	Phe His Arg His '	Thr Ser His His 80
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Arg Leu Pro Pro		His Thr Arg Arg 1	Lys Arg Lys Lys 110
Glu Lys Thr Set	r Ala Pro Pro Ser 120	Glu Gly Thr Pro	Pro Ile Gln Glu 125
Glu Gly Gly Ala	a Gly Val Asp Glu 135	Glu Glu Glu Glu (Glu Glu Glu Glu
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Pro Gln Lys Ala	a Lys Phe Ser Ile 165	Gly Ser Asp Glu i	Asp Asp Ser Pro 175
Gly Leu Pro Gly		Thr Lys Pro Leu 1	Pro Ser Val Gly 190
Pro His Thr Asp	o Lys Ser Pro Glr 200	His Ser Ser Ser :	Ser Pro Ser Pro 205
Arg Ala Arg Ala 210	a Ser Arg Leu Ala 215	Gly Glu Lys Ser 2	Arg Pro Trp Ser

Pro Ser Ala Ser Tyr Asp Leu Arg Glu Arg Leu Cys Pro Gly Ser Ala 225 230 230 235

Leu Gly Asn Pro Gly Gly Pro Glu Gln Gln Val Pro Thr Asp Glu Ala

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Leu 385	Asp	Leu	Glu	Gln	Thr 390	Thr	Leu	Pro	Gly	Ile 395	Ala	His	Leu	Val	Val 400
Glu	Thr	Met	Ile	Val 405	Ser	Asp	Gln	Ile	Arg 410	Pro	Glu	Asp	Arg	Ala 415	Ser
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Ala	Leu	Leu 995	Val	Leu	Ile	Leu	Ile 1000		e Me	t Glı	ı Th	r Gl:		le Th	nr Ala
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Thr	Gly 1085		l Leu	ı Ile	e Ala	109		eu Va	al G	ly L		er :	Ile V	/al M	Met

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His 945	Thr	Leu	Asp	Pro	Glu 950	ГÀа	Leu	Gly	Gln	Gly 955	Gly	Val	Gln	ГЛа	Val 960
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Thr	Gln 1055		ı Lev	ı Ala	a Phe	100		ln P	ro Se	er Se		la 065	Tyr .	Ala A	Ala
Phe	Asn 1070		n Arg	, Pro	Pro	Se: 10'		hr T	rp Le	eu Tl		la 080	Tyr '	Val V	/al
Lys	Val 1085		e Ser	: Leu	ı Ala	109		sn L	eu I	le A		le 095	Asp (Ser I	lis

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Leu	Thr 1550		n Ile	e Lys	5 Leu	Leu 155		sp As	sp P	he A	-	lu 560	Tyr	Thr	Met
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Gly	Gln 1580		n Arç	g Lys	Phe	Ile 158		er H	is I	le L		ys 590	Arg	Asn	Ala
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Asp	Gln 1640		з Туз	r Glr	ı Lys	Gln 164		78 G	lu G	lu L		ly 650	Ala	Phe	Thr
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Ile	Leu	Arg 35	Leu	Glu	Ser		Glu 40	Thr	Met	Val	Leu	Glu 45	ı Ala	His	Asp
Ala	Gln 50	Gly	Asp	Val		Val 55	Thr	Val	Thr	Val	His 60	Asp	Phe	Pro	Gly
Lys 65	Lys	Leu	Val	Leu	Ser 70	Ser	Glu	Lys	Thr	Val 75	Leu	Thr	Pro	Ala	Thr 80
Asn	His	Met	Gly	Asn 85	Val	Thr	Phe	Thr	Ile 90	Pro	Ala	Asn	a Arg	95	. Phe
Lys	Ser	Glu	Lys 100	Gly	Arg	Asn	Lys	Phe 105	Val	Thr	Val	Gln	Ala 110		Phe
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Val 145	Leu	_			150					155					160
145			Met	Val 165		Ile	Glu	Asn	Pro 170		Gly	Ile	Pro	Val 175	Lys
145 Arg	Thr	Val		165	Asn				170	Glu	_			175 Leu	Lys

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		195					200					205			
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Lys 225	Glu	Tyr	Val	Leu	Pro 230	Ser	Phe	Glu	Val	Ile 235	Val	Glu	Pro	Thr	Glu 240
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Aap	Leu	Val 515	Val	Leu	Pro	Leu	Ser 520	Ile	Thr	Thr	Asp	Phe 525	Ile	Pro	Ser
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Asn	Lys 610	Leu	Thr	Gln	Ser	Lys 615	Ile	Trp	Asp	Val	Val 620	Glu	Lys	Ala	Asp

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Leu	Lys 1595		Glu	Glu	Lys	Lys 1600		s Ty	r Le	u Me	t Trp 1605		Leu	Ser
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Lys	Asp 1625		Trp	Val	Glu	His 1630		p Pr	o Gl	u Gl	u Asp 1635		CAa	Gln
Asp	Glu 1640		. Asn	Gln	Lys	Gln 1645		s Gl	n As	p Le	u Gly 1650		Phe	Thr
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n Gl
n $\,$ Asn Tyr Gly Arg Lys Phe Val Gln Gly Lys Ser Ile Asp Val Ala Cys 410 His Pro Gly Tyr Ala Leu Pro Lys Ala Gln Thr Thr Val Thr Cys Met 425 Glu Asn Gly Trp Ser Pro Thr Pro Arg Cys Ile Arg Val Ser Phe Thr 440 Leu <210> SEQ ID NO 34 <211> LENGTH: 694 <212> TYPE: PRT <213 > ORGANISM: Mus musculus <400> SEQUENCE: 34 Met Gly Lys Ser Pro Gly Met Trp Cys Leu Val Leu Phe Ser Leu Leu Ala Ser Phe Ser Ala Glu Pro Thr Met His Gly Glu Ile Leu Ser Pro Asn Tyr Pro Gln Ala Tyr Pro Asn Asp Val Val Lys Ser Trp Asp Ile Glu Val Pro Glu Gly Phe Gly Ile His Leu Tyr Phe Thr His Val Asp Ile Glu Pro Ser Glu Ser Cys Ala Tyr Asp Ser Val Gln Ile Ile Ser 65 70 75 80 Gly Gly Ile Glu Glu Gly Arg Leu Cys Gly Gln Lys Thr Ser Lys Ser Pro Asn Ser Pro Ile Ile Glu Glu Phe Gln Phe Pro Tyr Asn Lys Leu 105 Gln Val Val Phe Thr Ser Asp Phe Ser Asn Glu Glu Arg Phe Thr Gly 120 Phe Ala Ala Tyr Tyr Thr Ala Ile Asp Ile Asn Glu Cys Thr Asp Phe 135 Thr Asp Val Pro Cys Ser His Phe Cys Asn Asn Phe Ile Gly Gly Tyr 150 155 Phe Cys Ser Cys Pro Pro Glu Tyr Phe Leu His Asp Asp Met Arg Asn

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Val Pro Val Thr Ser Leu Glu Thr Cys Lys Gln Val Lys Glu Glu Asn Pro Thr Val Arg Pro Glu Asp Tyr Val Phe Thr Asp Asn Met Ile Cys Ala Gly Glu Lys Gly Val Asp Ser Cys His Gly Asp Ser Gly Gly Ala Phe Ala Phe Gln Val Pro Asn Val Thr Val Pro Lys Phe Tyr Val Ala Gly Leu Val Ser Trp Gly Lys Arg Cys Gly Thr Tyr Gly Val Tyr Thr 665 Lys Val Lys Asn Tyr Val Asp Trp Ile Leu Lys Thr Met Gln Glu Asn 675 680 685 Ser Gly Pro Arg Lys Asp 690 <210> SEQ ID NO 35 <211> LENGTH: 688 <212> TYPE: PRT <213> ORGANISM: Homo sapiens <400> SEQUENCE: 35 Met Trp Cys Ile Val Leu Phe Ser Leu Leu Ala Trp Val Tyr Ala Glu 10 Pro Thr Met Tyr Gly Glu Ile Leu Ser Pro Asn Tyr Pro Gln Ala Tyr 25 Pro Ser Glu Val Glu Lys Ser Trp Asp Ile Glu Val Pro Glu Gly Tyr Gly Ile His Leu Tyr Phe Thr His Leu Asp Ile Glu Leu Ser Glu Asn Cys Ala Tyr Asp Ser Val Gln Ile Ile Ser Gly Asp Thr Glu Glu Gly Arg Leu Cys Gly Gln Arg Ser Ser Asn Asn Pro His Ser Pro Ile Val Glu Glu Phe Gln Val Pro Tyr Asn Lys Leu Gln Val Ile Phe Lys Ser Asp Phe Ser Asn Glu Glu Arg Phe Thr Gly Phe Ala Ala Tyr Tyr Val Ala Thr Asp Ile Asn Glu Cys Thr Asp Phe Val Asp Val Pro Cys Ser His Phe Cys Asn Asn Phe Ile Gly Gly Tyr Phe Cys Ser Cys Pro Pro Glu Tyr Phe Leu His Asp Asp Met Lys Asn Cys Gly Val Asn Cys Ser Gly Asp Val Phe Thr Ala Leu Ile Gly Glu Ile Ala Ser Pro Asn Tyr 185 Pro Lys Pro Tyr Pro Glu Asn Ser Arg Cys Glu Tyr Gln Ile Arg Leu 200 Glu Lys Gly Phe Gln Val Val Thr Leu Arg Arg Glu Asp Phe Asp Val Glu Ala Ala Asp Ser Ala Gly Asn Cys Leu Asp Ser Leu Val Phe 230 235 Val Ala Gly Asp Arg Gln Phe Gly Pro Tyr Cys Gly His Gly Phe Pro 250 Gly Pro Leu Asn Ile Glu Thr Lys Ser Asn Ala Leu Asp Ile Ile Phe 265

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Lys	Val 370	Glu	Asp	Pro	Glu	Ser 375	Thr	Leu	Phe	Gly	Ser 380	Val	Ile	Arg	Tyr
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Asp Gly Glu Ala 50	Gly Pro Pro Gl 55	ly Leu Pro Gly	Pro Pro Gly Pro Lys	
Gly Ala Pro Gly 65	Lys Pro Gly Ly 70	s Pro Gly Glu 75	Ala Gly Leu Pro Gly 80	
Leu Pro Gly Val	Asp Gly Leu Th 85	nr Gly Arg Asp 90	Gly Pro Pro Gly Pro 95	
Lys Gly Ala Pro 100	Gly Glu Arg Gl	ly Ser Leu Gly 105	Pro Pro Gly Pro Pro 110	
Gly Leu Gly Gly 115	Lys Gly Leu Pr 12	•	Gly Glu Ala Gly Val 125	
Ser Gly Pro Pro 130	Gly Gly Ile Gl 135	ly Leu Arg Gly	Pro Pro Gly Pro Ser 140	
Gly Leu Pro Gly 145	Leu Pro Gly Pr 150	ro Pro Gly Pro 155	Pro Gly Pro Pro Gly 160	
His Pro Gly Val	Leu Pro Glu Gl 165	ly Ala Thr Asp 170	Leu Gln Cys Pro Ser 175	
Ile Cys Pro Pro 180	Gly Pro Pro Gl	ly Pro Pro Gly 185	Met Pro Gly Phe Lys 190	
Gly Pro Thr Gly 195	Tyr Lys Gly Gl		Val Gly Lys Asp Gly 205	
Glu Lys Gly Asp 210	Pro Gly Pro Pr 215	ro Gly Pro Ala	Gly Leu Pro Gly Ser 220	
Val Gly Leu Gln 225	Gly Pro Arg Gl 230	ly Leu Arg Gly 235	Leu Pro Gly Pro Leu 240	

Gly Pro Pro Gly Asp Arg Gly Pro Ile Gly Phe Arg Gly Pro Pro Gly

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Arg	Ala	Gly 355	Glu	Leu	Gly	Glu	Ala 360	Gly	Pro	Ser	Gly	Glu 365	Pro	Gly	Val
Pro	Gly 370	Asp	Ala	Gly	Met	Pro 375	Gly	Glu	Arg	Gly	Glu 380	Ala	Gly	His	Arg
Gly 385	Ser	Ala	Gly	Ala	Leu 390	Gly	Pro	Gln	Gly	Pro 395	Pro	Gly	Ala	Pro	Gly 400
Val	Arg	Gly	Phe	Gln 405	Gly	Gln	Lys	Gly	Ser 410	Met	Gly	Asp	Pro	Gly 415	Leu
Pro	Gly	Pro	Gln 420	Gly	Leu	Arg	Gly	Asp 425	Val	Gly	Asp	Arg	Gly 430	Pro	Gly
Gly	Ala	Ala 435	Gly	Pro	ГÀа	Gly	Asp 440	Gln	Gly	Ile	Ala	Gly 445	Ser	Asp	Gly
Leu	Pro 450	Gly	Asp	ГÀа	Gly	Glu 455	Leu	Gly	Pro	Ser	Gly 460	Leu	Val	Gly	Pro
Lys 465	Gly	Glu	Ser	Gly	Ser 470	Arg	Gly	Glu	Leu	Gly 475	Pro	ГÀа	Gly	Thr	Gln 480
Gly	Pro	Asn	Gly	Thr 485	Ser	Gly	Val	Gln	Gly 490	Val	Pro	Gly	Pro	Pro 495	Gly
Pro	Leu	Gly	Leu 500	Gln	Gly	Val	Pro	Gly 505	Val	Pro	Gly	Ile	Thr 510	Gly	Lys
Pro	Gly	Val 515	Pro	Gly	ГÀа	Glu	Ala 520	Ser	Glu	Gln	Arg	Ile 525	Arg	Glu	Leu
Cys	Gly 530	Gly	Met	Ile	Ser	Glu 535	Gln	Ile	Ala	Gln	Leu 540	Ala	Ala	His	Leu
Arg 545	ГÀа	Pro	Leu	Ala	Pro 550	Gly	Ser	Ile	Gly	Arg 555	Pro	Gly	Pro	Ala	Gly 560
Pro	Pro	Gly	Pro	Pro 565	Gly	Pro	Pro	Gly	Ser 570	Ile	Gly	His	Pro	Gly 575	Ala
Arg	Gly	Pro	Pro 580	Gly	Tyr	Arg	Gly	Pro 585	Thr	Gly	Glu	Leu	Gly 590	Asp	Pro
Gly	Pro	Arg 595	Gly	Asn	Gln	Gly	Asp 600	Arg	Gly	Asp	rys	Gly 605	Ala	Ala	Gly
Ala	Gly 610	Leu	Asp	Gly	Pro	Glu 615	Gly	Asp	Gln	Gly	Pro 620	Gln	Gly	Pro	Gln
Gly 625	Val	Pro	Gly	Thr	Ser 630	Lys	Asp	Gly	Gln	Asp 635	Gly	Ala	Pro	Gly	Glu 640
Pro	Gly	Pro	Pro	Gly 645	Asp	Pro	Gly	Leu	Pro 650	Gly	Ala	Ile	Gly	Ala 655	Gln
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Ser Pro Pro Gln Pro Gly Ser Arg Thr Arg Gly Arg Gly Gln Gly Arg
Gly Thr Ala Met Pro Gly Glu Glu Val Leu Glu Ser Ser Gln Glu Ala
Leu His Val Thr Glu Arg Lys Tyr Leu Lys Arg Asp Trp Cys Lys Thr
Gln Pro Leu Lys Gln Thr Ile His Glu Glu Gly Cys Asn Ser Arg Thr
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Ile Ile Asn Arg Phe Cys Tyr Gly Gln Cys Asn Ser Phe Tyr Ile Pro
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Arg His Ile Arg Lys Glu Glu Gly Ser Phe Gln Ser Cys Ser Phe Cys
Lys Pro Lys Lys Phe Thr Thr Met Met Val Thr Leu Asn Cys Pro Glu
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Leu Gln Pro Pro Thr Lys Lys Lys Arg Val Thr Arg Val Lys Gln Cys
Arg Cys Ile Ser Ile Asp Leu Asp
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Ser Pro Gln Gln Pro Gly Ser Arg Asn Arg Gly Arg Gly Gln Gly Arg
Gly Thr Ala Met Pro Gly Glu Glu Val Leu Glu Ser Ser Gln Glu Ala
Leu His Val Thr Glu Arg Lys Tyr Leu Lys Arg Asp Trp Cys Lys Thr
Gln Pro Leu Lys Gln Thr Ile His Glu Glu Gly Cys Asn Ser Arg Thr
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Ile Ile Asn Arg Phe Cys Tyr Gly Gln Cys Asn Ser Phe Tyr Ile Pro
Arg His Ile Arg Lys Glu Glu Gly Ser Phe Gln Ser Cys Ser Phe Cys
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Leu	Gln	Pro	Pro	Thr 165	ГÀв	Lys	Lys	Arg	Val 170	Thr	Arg	Val	Lys	Gln 175	CAa
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Ser	Pro	Glu 35	Lys	Lys	Val	Gly	Ser 40	Gln	Gly	Leu	Arg	Phe 45	Arg	Leu	Ala
Gly	Phe 50	Pro	Arg	Lys	Pro	Tyr 55	Glu	Gly	Arg	Val	Glu 60	Ile	Gln	Arg	Ala
Gly 65	Glu	Trp	Gly	Thr	Ile 70	CÀa	Asp	Asp	Asp	Phe 75	Thr	Leu	Gln	Ala	Ala 80
His	Val	Leu	Cha	Arg 85	Glu	Leu	Gly	Phe	Thr 90	Glu	Ala	Thr	Gly	Trp 95	Thr
His	Ser	Ala	Lys	Tyr	Gly	Pro	Gly	Thr 105	Gly	Arg	Ile	Trp	Leu 110	Asp	Asn
Leu	Ser	Сув 115	Arg	Gly	Thr	Glu	Gly 120	Ser	Val	Thr	Glu	Cys 125	Ala	Ser	Arg
Gly	Trp 130	Gly	Asn	Ser	Asp	Сув 135	Thr	His	Asp	Glu	Asp 140	Ala	Gly	Val	Ile
Суз 145	Lys	Asp	Gln	Arg	Leu 150	Pro	Gly	Phe	Ser	Asp 155	Ser	Asn	Val	Ile	Glu 160
Val	Glu	His	Gln	Leu 165	Gln	Val	Glu	Glu	Val 170	Arg	Leu	Arg	Pro	Ala 175	Val
Glu	Trp	Gly	Arg 180	Arg	Pro	Leu	Pro	Val 185	Thr	Glu	Gly	Leu	Val 190	Glu	Val
Arg	Leu	Pro 195	Glu	Gly	Trp	Ser	Gln 200	Val	Càa	Asp	Lys	Gly 205	Trp	Ser	Ala
His	Asn 210	Ser	His	Val	Val	Cys 215	Gly	Met	Leu	Gly	Phe 220	Pro	Gly	Glu	Lys
Arg 225	Val	Asn	Met	Ala	Phe 230	Tyr	Arg	Met	Leu	Ala 235	Gln	ГÀв	Lys	Gln	His 240
Ser	Phe	Gly	Leu	His 245	Ser	Val	Ala	CÀa	Val 250	Gly	Thr	Glu	Ala	His 255	Leu
Ser	Leu	Cya	Ser 260	Leu	Glu	Phe	Tyr	Arg 265	Ala	Asn	Asp	Thr	Thr 270	Arg	Cys
Ser	Gly	Gly 275	Asn	Pro	Ala	Val	Val 280	Ser	Cys	Val	Leu	Gly 285	Pro	Leu	Tyr
Ala	Thr 290	Phe	Thr	Gly	Gln	Lys 295	Lys	Gln	Gln	His	Ser 300	Lys	Pro	Gln	Gly
Glu 305	Ala	Arg	Val	Arg	Leu 310	Lys	Gly	Gly	Ala	His 315	Gln	Gly	Glu	Gly	Arg 320
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Thr	Ala	Arg 355	Glu	Ala	Leu	Ser	Gly 360	Ala	Arg	Met	Gly	Gln 365	Gly	Met	Gly
Ala	Ile 370	His	Leu	Ser	Glu	Val 375	Arg	Cha	Ser	Gly	Gln 380	Glu	Pro	Ser	Leu
Trp 385	Arg	Cys	Pro	Ser	390	Asn	Ile	Thr	Ala	Glu 395	Asp	Cys	Ser	His	Ser 400
Gln	Asp	Ala	Gly	Val 405	Arg	Cys	Asn	Leu	Pro 410	Tyr	Thr	Gly	Val	Glu 415	Thr
Lys	Ile	Arg	Leu 420	Ser	Gly	Gly	Arg	Ser 425	Arg	Tyr	Glu	Gly	Arg 430	Val	Glu
Val	Gln	Ile 435	Gly	Ile	Pro	Gly	His 440	Leu	Arg	Trp	Gly	Leu 445	Ile	CAa	Gly
Asp	Asp 450	Trp	Gly	Thr	Leu	Glu 455	Ala	Met	Val	Ala	Cys 460	Arg	Gln	Leu	Gly
Leu 465	Gly	Tyr	Ala	Asn	His 470	Gly	Leu	Gln	Glu	Thr 475	Trp	Tyr	Trp	Asp	Ser 480
Gly	Asn	Val	Thr	Glu 485	Val	Val	Met	Ser	Gly 490	Val	Arg	CÀa	Thr	Gly 495	Ser
Glu	Leu	Ser	Leu 500	Asn	Gln	Cys	Ala	His 505	His	Ser	Ser	His	Ile 510	Thr	Cys
Lys	Lys	Thr 515	Gly	Thr	Arg	Phe	Thr 520	Ala	Gly	Val	Ile	Сув 525	Ser	Glu	Thr
Ala	Ser 530	Asp	Leu	Leu	Leu	His 535	Ser	Ala	Leu	Val	Gln 540	Glu	Thr	Ala	Tyr
Ile 545	Glu	Asp	Arg	Pro	Leu 550	His	Met	Leu	Tyr	Сув 555	Ala	Ala	Glu	Glu	Asn 560
Cys	Leu	Ala	Ser	Ser 565	Ala	Arg	Ser	Ala	Asn 570	Trp	Pro	Tyr	Gly	His 575	Arg
Arg	Leu	Leu	Arg 580	Phe	Ser	Ser	Gln	Ile 585	His	Asn	Leu	Gly	Arg 590	Ala	Asp
Phe	Arg	Pro 595	Lys	Ala	Gly	Arg	His 600	Ser	Trp	Val	Trp	His 605	Glu	Cys	His
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Pro 625	Asn	Gly	Thr	Lys	Val 630	Ala	Glu	Gly	His	635	Ala	Ser	Phe	CAa	Leu 640
Glu	Asp	Thr	Glu	Сув 645	Gln	Glu	Asp	Val	Ser 650	Lys	Arg	Tyr	Glu	Сув 655	Ala
Asn	Phe	Gly	Glu 660	Gln	Gly	Ile	Thr	Val 665	Gly	Cys	Trp	Asp	Leu 670	Tyr	Arg
His	Asp	Ile 675	Asp	CAa	Gln	Trp	Ile 680	Asp	Ile	Thr	Asp	Val 685	ГÀв	Pro	Gly
Asn	Tyr 690	Ile	Leu	Gln	Val	Val 695	Ile	Asn	Pro	Asn	Phe 700	Glu	Val	Ala	Glu
Ser 705	Asp	Phe	Thr	Asn	Asn 710	Ala	Met	Lys	Cys	Asn 715	Cys	Lys	Tyr	Asp	Gly 720
His	Arg	Ile	Trp	Val 725	His	Asn	Cys	His	Ile 730	Gly	Asp	Ala	Phe	Ser 735	Glu
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Pro	Glu	Lys 35	ГÀа	Ala	Gly	Ser	Gln 40	Gly	Leu	Arg	Phe	Arg 45	Leu	Ala	Gly
Phe	Pro 50	Arg	Lys	Pro	Tyr	Glu 55	Gly	Arg	Val	Glu	Ile 60	Gln	Arg	Ala	Gly
Glu 65	Trp	Gly	Thr	Ile	Cys 70	Asp	Asp	Asp	Phe	Thr 75	Leu	Gln	Ala	Ala	His 80
Ile	Leu	Cys	Arg	Glu 85	Leu	Gly	Phe	Thr	Glu 90	Ala	Thr	Gly	Trp	Thr 95	His
Ser	Ala	Lys	Tyr 100	Gly	Pro	Gly	Thr	Gly 105	Arg	Ile	Trp	Leu	Asp 110	Asn	Leu
Ser	Cys	Ser 115	Gly	Thr	Glu	Gln	Ser 120	Val	Thr	Glu	Cys	Ala 125	Ser	Arg	Gly
Trp	Gly 130	Asn	Ser	Asp	Cys	Thr 135	His	Asp	Glu	Asp	Ala 140	Gly	Val	Ile	Cys
Lys 145	Asp	Gln	Arg	Leu	Pro 150	Gly	Phe	Ser	Asp	Ser 155	Asn	Val	Ile	Glu	Ala 160
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Val	Leu	Lys	Ala 180	Ser	Thr	Trp	Gly	Thr 185	Val	Сув	Asp	Arg	Lys 190	Trp	Asp
Leu	His	Ala 195	Ala	Ser	Val	Val	Cys 200	Arg	Glu	Leu	Gly	Phe 205	Gly	Ser	Ala
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His 225	Leu	Ser	Glu	Val	Arg 230	Cys	Ser	Gly	Gln	Glu 235	Leu	Ser	Leu	Trp	Lys 240
Cys	Pro	His	Lys	Asn 245	Ile	Thr	Ala	Glu	Asp 250	Cys	Ser	His	Ser	Gln 255	Asp
Ala	Gly	Val	Arg 260	Cys	Asn	Leu	Pro	Tyr 265	Thr	Gly	Ala	Glu	Thr 270	Arg	Ile
Arg	Leu	Ser 275	Gly	Gly	Arg	Ser	Gln 280	His	Glu	Gly	Arg	Val 285	Glu	Val	Gln
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Trp 305	Gly	Thr	Leu	Glu	Ala 310	Met	Val	Ala	Cha	Arg 315	Gln	Leu	Gly	Leu	Gly 320
Tyr	Ala	Asn	His	Gly 325	Leu	Gln	Glu	Thr	Trp 330	Tyr	Trp	Asp	Ser	Gly 335	Asn
Ile	Thr	Glu	Val 340	Val	Met	Ser	Gly	Val 345	Arg	Cys	Thr	Gly	Thr 350	Glu	Leu
Ser	Leu	Asp 355	Gln	СЛа	Ala	His	His 360	Gly	Thr	His	Ile	Thr 365	Сув	Lys	Arg
Thr	Gly	Thr	Arg	Phe	Thr	Ala	Gly	Val	Ile	GÀa	Ser	Glu	Thr	Ala	Ser

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Asp	Arg	Pro	Leu	His 405	Met	Leu	Tyr	Cys	Ala 410	Ala	Glu	Glu	Asn	Cys 415	Leu
Ala	Ser	Ser	Ala 420	Arg	Ser	Ala	Asn	Trp 425	Pro	Tyr	Gly	His	Arg 430	Arg	Leu
Leu	Arg	Phe 435	Ser	Ser	Gln	Ile	His 440	Asn	Leu	Gly	Arg	Ala 445	Asp	Phe	Arg
Pro	Lys 450	Ala	Gly	Arg	His	Ser 455	Trp	Val	Trp	His	Glu 460	Сла	His	Gly	His
Tyr 465	His	Ser	Met	Asp	Ile 470	Phe	Thr	His	Tyr	Asp 475	Ile	Leu	Thr	Pro	Asn 480
Gly	Thr	Lys	Val	Ala 485	Glu	Gly	His	Lys	Ala 490	Ser	Phe	Cys	Leu	Glu 495	Asp
Thr	Glu	Cys	Gln 500	Glu	Asp	Val	Ser	Lys 505	Arg	Tyr	Glu	CÀa	Ala 510	Asn	Phe
Gly	Glu	Gln 515	Gly	Ile	Thr	Val	Gly 520	Cys	Trp	Aap	Leu	Tyr 525	Arg	His	Asp
Ile	Asp 530	CÀa	Gln	Trp	Ile	Asp 535	Ile	Thr	Asp	Val	Lys 540	Pro	Gly	Asn	Tyr
Ile 545	Leu	Gln	Val	Val	Ile 550	Asn	Pro	Asn	Phe	Glu 555	Val	Ala	Glu	Ser	Asp 560
Phe	Thr	Asn	Asn	Ala 565	Met	Lys	Cys	Asn	Сув 570	Lys	Tyr	Asp	Gly	His 575	Arg
Ile	Trp	Val	His 580	Asn	Сув	His	Ile	Gly 585	Asp	Ala	Phe	Ser	Glu 590	Glu	Ala
Asn	Arg	Arg	Phe	Glu	Arg	Tyr	Pro	Gly	Gln	Thr	Ser	Asn	Gln	Ile	Ile
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145 155 Pro Asp Gly Leu <210> SEQ ID NO 43 <211> LENGTH: 173 <212> TYPE: PRT <213 > ORGANISM: Homo sapiens <400> SEQUENCE: 43 Met Ser Leu Leu Gly Pro Lys Val Leu Leu Phe Leu Ala Ala Phe Ile 10 Ile Thr Ser Asp Trp Ile Pro Leu Gly Val Asn Ser Gln Arg Gly Asp $20 \\ 25 \\ 30$ Asp Val Thr Gln Ala Thr Pro Glu Thr Phe Thr Glu Asp Pro Asn Leu 40 Val Asn Asp Pro Ala Thr Asp Glu Thr Val Leu Ala Val Leu Ala Asp 55 Ile Ala Pro Ser Thr Asp Asp Leu Ala Ser Leu Ser Glu Lys Asn Thr Thr Ala Glu Cys Trp Asp Glu Lys Phe Thr Cys Thr Arg Leu Tyr Ser Val His Arg Pro Val Lys Gln Cys Ile His Gln Leu Cys Phe Thr Ser 105 Leu Arg Arg Met Tyr Ile Val Asn Lys Glu Ile Cys Ser Arg Leu Val 120 Cys Lys Glu His Glu Ala Met Lys Asp Glu Leu Cys Arg Gln Met Ala 135 Gly Leu Pro Pro Arg Arg Leu Arg Arg Ser Asn Tyr Phe Arg Leu Pro Pro Cys Glu Asn Val Asp Leu Gln Arg Pro Asn Gly Leu <210> SEQ ID NO 44 <211> LENGTH: 104 <213> ORGANISM: Mus musculus <400> SEQUENCE: 44 Met Lys Ser Leu Leu Pro Leu Ala Ile Leu Ala Ala Leu Ala Val Ala Thr Leu Cys Tyr Glu Ser His Glu Ser Met Glu Ser Tyr Glu Ile Ser Pro Phe Ile Asn Arg Arg Asn Ala Asn Thr Phe Met Ser Pro Gln Gln 40 Arg Trp Arg Ala Lys Ala Gln Lys Arg Val Gln Glu Arg Asn Lys Pro 55 Ala Tyr Glu Ile Asn Arg Glu Ala Cys Asp Asp Tyr Lys Leu Cys Glu 65 70 75 80 Arg Tyr Ala Met Val Tyr Gly Tyr Asn Ala Ala Tyr Asn Arg Tyr Phe 90 Arg Gln Arg Arg Gly Ala Lys Tyr 100 <210> SEQ ID NO 45 <211> LENGTH: 103 <212> TYPE: PRT <213 > ORGANISM: Homo sapiens

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Pro Phe Ile Asn Arg Arg Asn Ala Asn Thr Phe Ile Ser Pro Gln Gln
Arg Trp Arg Ala Lys Val Gln Glu Arg Ile Arg Glu Arg Ser Lys Pro
Val His Glu Leu Asn Arg Glu Ala Cys Asp Asp Tyr Arg Leu Cys Glu 65 70 75 80
Arg Tyr Ala Met Val Tyr Gly Tyr Asn Ala Ala Tyr Asn Arg Tyr Phe
Arg Lys Arg Arg Gly Ala Lys
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Cys Ile Ser Glu Arg Leu Phe Met Glu Met Ala Asp Arg Leu Ala Gln
Asp Gly Trp Arg Asp Leu Gly Tyr Val Tyr Leu Asn Ile Asp Asp Cys
Trp Ile Gly Gly Arg Asp Ala Ser Gly Arg Leu Ile Pro Asp Pro Lys
Arg Phe Pro His Gly Ile Ala Phe Leu Ala Asp Tyr Ala His Ser Leu
Gly Leu Lys Leu Gly Ile Tyr Glu Asp Met Gly Lys Met Thr Cys Met
Gly Tyr Pro Gly Thr Thr Leu Asp Lys Val Glu Leu Asp Ala Glu Thr
                       135
Phe Ala Glu Trp Lys Val Asp Met Leu Lys Leu Asp Gly Cys Phe Ser
                                       155
Ser Ser Arg Glu Arg Ala Glu Gly Tyr Pro Lys Met Ala Ala Ala Leu
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Asn Ala Thr Gly Arg Pro Ile Ala Phe Ser Cys Ser Trp Pro Ala Tyr
Glu Gly Gly Leu Pro Pro Lys Val Asn Tyr Thr Glu Val Ser Arg Val
                           200
Cys Asn Leu Trp Arg Asn Tyr Lys Asp Ile Gln Asp Ser Trp Lys Ser
                      215
Val Leu Ser Ile Leu Asp Trp Phe Val Arg His Gln Asp Val Pro Gln
                                       235
Pro Val Ala Gly Pro Gly His Trp Asn Asp Pro Asp Met Leu Leu Ile
Gly Asn Phe Gly Leu Ser Phe Asp Glu Ser Arg Ala Gln Met Ala Leu
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			260					265					270		
Trp	Thr	Val 275	Leu	Ala	Ala	Pro	Leu 280	Leu	Met	Ser	Thr	Asp 285	Leu	Arg	Thr
Ile	Ser 290	Pro	Gln	Asn	Met	Asp 295	Ile	Leu	Gln	Asn	Pro 300	Leu	Met	Ile	ГÀв
Ile 305	Asn	Gln	Asp	Pro	Leu 310	Gly	Ile	Gln	Gly	Arg 315	Arg	Ile	Leu	Lys	Ser 320
Lys	Ser	His	Ile	Glu 325	Val	Phe	Lys	Arg	Tyr 330	Leu	Ser	Asn	Gln	Ala 335	Ser
Ala	Leu	Val	Phe 340	Phe	Ser	Arg	Arg	Thr 345	Asp	Met	Pro	Phe	Arg 350	Phe	His
CÀa	Ser	Leu 355	Leu	Glu	Leu	Asn	Tyr 360	Pro	ГЛа	Gly	Arg	Val 365	Tyr	Glu	Gly
Gln	Asn 370	Val	Phe	Thr	Gly	Asp 375	Ile	Phe	Ser	Gly	Leu 380	Gln	Thr	Glu	Val
Asn 385	Phe	Thr	Val	Ile	Ile 390	Asn	Pro	Ser	Gly	Val 395	Val	Met	Trp	Tyr	Leu 400
Tyr	Pro	Ile	Lys	Asp 405	Leu	Gly	Ile	Ser	Thr 410	Met	Met	Ser	His	Trp 415	
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	2 > T) 3 > OF			Homo	sa]	piens	3								
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Met	Leu	Asp	Asn 20	Gly	Leu	Leu	Gln	Thr 25	Pro	Pro	Met	Gly	Trp	Leu	Ala
Trp	Glu	Arg 35	Phe	Arg	Cys	Asn	Ile 40	Asn	Cys	Asp	Glu	Asp 45	Pro	Lys	Asn
Cha	Ile 50	Ser	Glu	Gln	Leu	Phe 55	Met	Glu	Met	Ala	Asp 60	Arg	Met	Ala	Gln
Asp 65	Gly	Trp	Arg	Asp	Met 70	Gly	Tyr	Thr	Tyr	Leu 75	Asn	Ile	Asp	Asp	80 CÀa
Trp	Ile	Gly	Gly	Arg 85	Asp	Ala	Ser	Gly	Arg 90	Leu	Met	Pro	Asp	Pro 95	Lys
Arg	Phe	Pro	His 100	Gly	Ile	Pro	Phe	Leu 105	Ala	Asp	Tyr	Val	His 110	Ser	Leu
Gly	Leu	Lys 115	Leu	Gly	Ile	Tyr	Ala 120	Asp	Met	Gly	Asn	Phe 125	Thr	Сла	Met
Gly	Tyr 130	Pro	Gly	Thr	Thr	Leu 135	Asp	Lys	Val	Val	Gln 140	Asp	Ala	Gln	Thr
Phe 145	Ala	Glu	Trp	Lys	Val 150	Asp	Met	Leu	Lys	Leu 155	Asp	Gly	Cys	Phe	Ser 160
Thr	Pro	Glu	Glu	Arg 165	Ala	Gln	Gly	Tyr	Pro 170	Lys	Met	Ala	Ala	Ala 175	Leu
Asn	Ala	Thr	Gly 180	Arg	Pro	Ile	Ala	Phe 185	Ser	CÀa	Ser	Trp	Pro 190	Ala	Tyr
Glu	Gly	Gly 195	Leu	Pro	Pro	Arg	Val 200	Asn	Tyr	Ser	Leu	Leu 205	Ala	Asp	Ile
CAa	Asn 210	Leu	Trp	Arg	Asn	Tyr 215	Asp	Asp	Ile	Gln	Asp 220	Ser	Trp	Trp	Ser
Val	Leu	Ser	Ile	Leu	Asn	Trp	Phe	Val	Glu	His	Gln	Asp	Ile	Leu	Gln

			continued
225	230	235	

Pro Val Ala Gly Pro Gly His Trp Asn Asp Pro Asp Met Leu Leu Ile

Gly Asn Phe Gly Leu Ser Leu Glu Gln Ser Arg Ala Gln Met Ala Leu 260 265 270

Ile Ser Ala Gln Asn Met Asp Ile Leu Gln Asn Pro Leu Met Ile Lys 290 \$295\$

Ile Asn Gln Asp Pro Leu Gly Ile Gln Gly Arg Arg Ile His Lys Glu 305 310 315 320

Lys Ser Leu Ile Glu Val Tyr Met Arg Pro Leu Ser Asn Lys Ala Ser 325 \$330\$

Ala Leu Val Phe Phe Ser Cys Arg Thr Asp Met Pro Tyr Arg Tyr His

Ser Ser Leu Gly Gln Leu Asn Phe Thr Gly Ser Val Ile Tyr Glu Ala 355 360 365

Gln Asp Val Tyr Ser Gly Asp Ile Ile Ser Gly Leu Arg Asp Glu Thr 370 380

Asn Phe Thr Val Ile Ile Asn Pro Ser Gly Val Val Met Trp Tyr Leu 385 390 395 400

Tyr Pro Ile Lys Asn Leu Glu Met Ser Gln Gln 405 410

<210> SEQ ID NO 48

<211> LENGTH: 178

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<400> SEQUENCE: 48

Met Leu Trp Val Leu Val Gly Ala Val Leu Pro Val Met Leu Leu Ala 1 5 10 15

Ala Pro Pro Ile Asn Lys Leu Ala Leu Phe Pro Asp Lys Ser Ala 20 25 30

Trp Cys Glu Ala Lys Asn Ile Thr Gln Ile Val Gly His Ser Gly Cys

Glu Ala Lys Ser Ile Gln Asn Arg Ala Cys Leu Gly Gln Cys Phe Ser 50 60

Tyr Ser Val Pro Asn Thr Phe Pro Gln Ser Thr Glu Ser Leu Val His 65 70 75 80

Glu Cys Pro Asp His Glu Glu Val Pro Arg Val Asp Lys Leu Val Glu $100 \hspace{1.5cm} 105 \hspace{1.5cm} 110 \hspace{1.5cm}$

Lys Ile Val His Cys Ser Cys Gln Ala Cys Gly Lys Glu Pro Ser His 115 \$120\$ 125

Glu Gly Leu Asn Val Tyr Val Gln Gly Glu Asp Ser Pro Gly Ser Gln 130 \$135\$

Pro Gly Pro His Ser His Ala His Pro His Pro Gly Gly Gln Thr Pro 145 150 155 160

Glu Pro Glu Glu Pro Pro Gly Ala Pro Gln Val Glu Glu Glu Gly Ala 165 170 175

Glu Asp

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Lys Arg Arg Leu His Ser Pro Arg Val Leu Phe Ser Thr Gln Pro Pro

155

150

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Pro Thr Ser Ser Asp Thr Leu Asp Leu Asp Phe Gln Ala His Gly Thr Ile Pro Phe Asn Arg Thr His Arg Ser Lys Arg Ser Ser Thr His Pro Val Phe His Met Gly Glu Phe Ser Val Cys Asp Ser Val Ser Val Trp Val Gly Asp Lys Thr Thr Ala Thr Asp Ile Lys Gly Lys Glu Val Thr Val Leu Ala Glu Val Asn Ile Asn Asn Ser Val Phe Arg Gln Tyr Phe 235 230 Phe Glu Thr Lys Cys Arg Ala Ser Asn Pro Val Glu Ser Gly Cys Arg 245 250 Gly Ile Asp Ser Lys His Trp Asn Ser Tyr Cys Thr Thr Thr His Thr 265 Phe Val Lys Ala Leu Thr Thr Asp Glu Lys Gln Ala Ala Trp Arg Phe 280 Ile Arg Ile Asp Thr Ala Cys Val Cys Val Leu Ser Arg Lys Ala Thr 295 300 Arg Arg Gly 305 <210> SEQ ID NO 51 <211> LENGTH: 299 <212> TYPE: PRT <213 > ORGANISM: Homo sapiens <400> SEQUENCE: 51 Gly Arg Val Gly Ala Gly Ser Arg Arg Gly Ala Gln Arg Val Leu Ala 10 Ser Gly Arg Ala Val Gln Gly Ala Gly Trp His Ala Gly Pro Lys Leu Ser Ser Ala Ser Gly Pro Asn Asn Ser Phe Thr Lys Gly Ala Ala Phe Tyr Pro Gly His Thr Glu Val His Ser Val Met Ser Met Leu Phe Tyr Thr Leu Ile Thr Ala Phe Leu Ile Gly Ile Gln Ala Glu Pro His Ser Glu Ser Asn Val Pro Ala Gly His Thr Ile Pro Gln Val His Trp Thr Lys Leu Gln His Ser Leu Asp Thr Ala Leu Arg Arg Ala Arg Ser Ala Pro Ala Ala Ala Ile Ala Ala Arg Val Ala Gly Gln Thr Arg Asn Ile 120 Thr Val Asp Pro Arg Leu Phe Lys Lys Arg Arg Leu Arg Ser Pro Arg 135 140 Val Leu Phe Ser Thr Gln Pro Pro Arg Glu Ala Ala Asp Thr Gln Asp 150 155 Leu Asp Phe Glu Val Gly Gly Ala Ala Pro Phe Asn Arg Thr His Arg Ser Lys Arg Ser Ser Ser His Pro Ile Phe His Arg Gly Glu Phe Ser 185 Val Cys Asp Ser Val Ser Val Trp Val Gly Asp Lys Thr Thr Ala Thr 200 Asp Ile Lys Gly Lys Glu Val Met Val Leu Gly Glu Val Asn Ile Asn 215

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Asn Ser Val Phe Lys Gln Tyr Phe Phe Glu Thr Lys Cys Arg Asp Pro Asn Pro Val Asp Ser Gly Cys Arg Gly Ile Asp Ser Lys His Trp Asn Ser Tyr Cys Thr Thr Thr His Thr Phe Val Lys Ala Leu Thr Met Asp Gly Lys Gln Ala Ala Trp Arg Phe Ile Arg Ile Asp Thr Ala Cys Val Cys Val Leu Ser Arg Lys Ala Val Arg Arg Ala 290 295 <210> SEQ ID NO 52 <211> LENGTH: 592 <212> TYPE: PRT <213 > ORGANISM: Mus musculus <400> SEQUENCE: 52 Met Ala Val Leu Leu Ala Ala Val Leu Ala Ser Ser Leu Tyr Leu Gln Val Ala Ala Asp Phe Asp Gly Arg Trp Pro Arg Gln Ile Val Ser Ser 25 Ile Gly Leu Cys Arg Tyr Gly Gly Arg Ile Asp Cys Cys Trp Gly Trp 40 Ala Arg Gln Ser Trp Gly Gln Cys Gln Pro Val Cys Gln Pro Gln Cys Lys His Gly Glu Cys Val Gly Pro Asn Lys Cys Lys Cys His Pro Gly 65 70 70 75 75 80 Phe Ala Gly Lys Thr Cys Asn Gln Asp Glu Ser Phe His Pro Thr Pro Leu Asp Gln Gly Ser Glu Gln Pro Leu Phe Gln Pro Pro Asp His Gln 105 Ala Thr Asn Val Pro Ser Arg Asp Leu Asn Glu Cys Gly Leu Lys Pro Arg Pro Cys Lys His Arg Cys Met Asn Thr Phe Gly Ser Tyr Lys Cys Tyr Cys Leu Asn Gly Tyr Met Leu Leu Pro Asp Gly Ser Cys Ser Ser Ala Leu Ser Cys Ser Met Ala Asn Cys Gln Tyr Gly Cys Asp Val Val Lys Gly Gln Val Arg Cys Gln Cys Pro Ser Pro Gly Leu Gln Leu Ala Pro Asp Gly Arg Thr Cys Val Asp Ile Asp Glu Cys Ala Thr Gly Arg 200 Val Ser Cys Pro Arg Phe Arg Gln Cys Val Asn Thr Phe Gly Ser Tyr 215 Ile Cys Lys Cys His Thr Gly Phe Asp Leu Met Tyr Ile Gly Gly Lys 230 235 Tyr Gln Cys His Asp Ile Asp Glu Cys Ser Leu Gly Gln His Gln Cys Ser Ser Tyr Ala Arg Cys Tyr Asn Ile His Gly Ser Tyr Lys Cys Gln 265 Cys Arg Asp Gly Tyr Glu Gly Asp Gly Leu Asn Cys Val Tyr Ile Pro 280 Lys Val Met Ile Glu Pro Ser Gly Pro Ile His Met Pro Glu Arg Asn 295

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Gly Thr Ile Ser Lys Gly Asp Gly Gly His Ala Asn Arg Ile Pro Asp 305 310 315 320

Ala Gly Ser Thr Arg Trp Pro Leu Lys Thr Pro Tyr Ile Pro Pro Val\$325\$

				325					330					335	
Ile	Thr	Asn	Arg 340	Pro	Thr	Ser	Lys	Pro 345	Thr	Thr	Arg	Pro	Thr 350	Pro	Asn
Pro	Thr	Pro 355	Gln	Pro	Thr	Pro	Pro 360	Pro	Pro	Pro	Pro	Leu 365	Pro	Thr	Glu
Pro	Arg 370	Thr	Thr	Pro	Leu	Pro 375	Pro	Thr	Pro	Glu	Arg 380	Pro	Ser	Thr	Arg
Pro 385	Thr	Thr	Ile	Ala	Pro 390	Ala	Thr	Ser	Thr	Thr 395	Thr	Arg	Val	Ile	Thr 400
Val	Asp	Asn	Arg	Ile 405	Gln	Thr	Asp	Pro	Gln 410	Lys	Pro	Arg	Gly	Asp 415	Val
Phe	Ile	Pro	Arg 420	Gln	Pro	Thr	Asn	Asp 425	Leu	Phe	Glu	Ile	Phe 430	Glu	Ile
Glu	Arg	Gly 435	Val	Ser	Ala	Asp	Glu 440	Glu	Val	ГÀа	Asp	Asp 445	Pro	Gly	Ile
Leu	Ile 450	His	Ser	Cys	Asn	Phe 455	Asp	His	Gly	Leu	Cys 460	Gly	Trp	Ile	Arg
Glu 465	ГÀв	Asp	Ser	Asp	Leu 470	His	Trp	Glu	Thr	Ala 475	Arg	Asp	Pro	Ala	Gly 480
Gly	Gln	Tyr	Leu	Thr 485	Val	Ser	Ala	Ala	Lys 490	Ala	Pro	Gly	Gly	Lys 495	Ala
Ala	Arg	Leu	Val 500	Leu	Arg	Leu	Gly	His 505	Leu	Met	His	Ser	Gly 510	Asp	Leu
CÀa	Leu	Ser 515	Phe	Arg	His	Lys	Val 520	Thr	Gly	Leu	His	Ser 525	Gly	Thr	Leu
Gln	Val 530	Phe	Val	Arg	Lys	His 535	Gly	Thr	His	Gly	Ala 540	Ala	Leu	Trp	Gly
Arg 545	Asn	Gly	Gly	His	Gly 550	Trp	Arg	Gln	Thr	Gln 555	Ile	Thr	Leu	Arg	Gly 560
Ala	Asp	Val	Lys	Ser 565	Val	Ile	Phe	Lys	Gly 570	Glu	Lys	Arg	Arg	Gly 575	His
Thr	Gly	Glu	Ile 580	Gly	Leu	Asp	Asp	Val 585	Ser	Leu	Lys	Arg	Gly 590	Arg	Cys
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Ala	Ala	Ala	Glu 20	Phe	Asp	Gly	Arg	Trp 25	Pro	Arg	Gln	Ile	Val 30	Ser	Ser
Ile	Gly	Leu 35	Cys	Arg	Tyr	Gly	Gly 40	Arg	Ile	Asp	Cys	Cys 45	Trp	Gly	Trp
Ala	Arg 50	Gln	Ser	Trp	Gly	Gln 55	Cys	Gln	Pro	Val	Gys	Gln	Pro	Arg	Cya
Lys 65	His	Gly	Glu	Cys	Ile 70	Gly	Pro	Asn	Lys	Сув 75	Lys	Cys	His	Pro	Gly 80
Tyr	Ala	Gly	Lys	Thr 85	Сув	Asn	Gln	Asp	Leu 90	Asn	Glu	Сув	Gly	Leu 95	Lys

Pro	Arg	Pro	Cys 100	Lys	His	Arg	Cys	Met 105	Asn	Thr	Tyr	Gly	Ser 110	Tyr	Lys
Cys	Tyr	Cys 115	Leu	Asn	Gly	Tyr	Met 120	Leu	Met	Pro	Asp	Gly 125	Ser	Cys	Ser
Ser	Ala 130	Leu	Thr	Сув	Ser	Met 135	Ala	Asn	Сув	Gln	Tyr 140	Gly	Сув	Asp	Val
Val 145	Lys	Gly	Gln	Ile	Arg 150	СЛа	Gln	Сла	Pro	Ser 155	Pro	Gly	Leu	Gln	Leu 160
Ala	Pro	Asp	Gly	Arg 165	Thr	Cys	Val	Asp	Val 170	Asp	Glu	Cys	Ala	Thr 175	Gly
Arg	Ala	Ser	Cys 180	Pro	Arg	Phe	Arg	Gln 185	Cys	Val	Asn	Thr	Phe 190	Gly	Ser
Tyr	Ile	Сув 195	ГЛа	CÀa	His	ГЛа	Gly 200	Phe	Asp	Leu	Met	Tyr 205	Ile	Gly	Gly
ГÀа	Tyr 210	Gln	Cha	His	Asp	Ile 215	Asp	Glu	Cya	Ser	Leu 220	Gly	Gln	Tyr	Gln
Сув 225	Ser	Ser	Phe	Ala	Arg 230	Cys	Tyr	Asn	Ile	Arg 235	Gly	Ser	Tyr	Lys	Cys 240
Lys	Сув	Lys	Glu	Gly 245	Tyr	Gln	Gly	Asp	Gly 250	Leu	Thr	Càa	Val	Tyr 255	Ile
Pro	Lys	Val	Met 260	Ile	Glu	Pro	Ser	Gly 265	Pro	Ile	His	Val	Pro 270	ГЛа	Gly
Asn	Gly	Thr 275	Ile	Leu	ГÀа	Gly	Asp 280	Thr	Gly	Asn	Asn	Asn 285	Trp	Ile	Pro
Asp	Val 290	Gly	Ser	Thr	Trp	Trp 295	Pro	Pro	Lys	Thr	Pro 300	Tyr	Ile	Pro	Pro
Ile 305	Ile	Thr	Asn	Arg	Pro 310	Thr	Ser	Lys	Pro	Thr 315	Thr	Arg	Pro	Thr	Pro 320
ГÀа	Pro	Thr	Pro	Ile 325	Pro	Thr	Pro	Pro	Pro 330	Pro	Pro	Pro	Leu	Pro 335	Thr
Glu	Leu	Arg	Thr 340	Pro	Leu	Pro	Pro	Thr 345	Thr	Pro	Glu	Arg	Pro 350	Thr	Thr
Gly	Leu	Thr 355	Thr	Ile	Ala	Pro	Ala 360	Ala	Ser	Thr	Pro	Pro 365	Gly	Gly	Ile
Thr	Val 370	Asp	Asn	Arg	Val	Gln 375	Thr	Asp	Pro	Gln	380 TÀs	Pro	Arg	Gly	Asp
Val 385	Phe	Ile	Pro	Arg	Gln 390	Pro	Ser	Asn	Asp	Leu 395	Phe	Glu	Ile	Phe	Glu 400
Ile	Glu	Arg	Gly	Val 405	Ser	Ala	Asp	Asp	Glu 410	Ala	ГÀа	Asp	Asp	Pro 415	Gly
Val	Leu	Val	His 420	Ser	Càa	Asn	Phe	Asp 425	His	Gly	Leu	Càa	Gly 430	Trp	Ile
Arg	Glu	Lys 435	Asp	Asn	Asp	Leu	His 440	Trp	Glu	Pro	Ile	Arg 445	Asp	Pro	Ala
Gly	Gly 450	Gln	Tyr	Leu	Thr	Val 455	Ser	Ala	Ala	ГЛа	Ala 460	Pro	Gly	Gly	Lys
Ala 465	Ala	Arg	Leu	Val	Leu 470	Pro	Leu	Gly	Arg	Leu 475	Met	His	Ser	Gly	Asp 480
Leu	Сув	Leu	Ser	Phe 485	Arg	His	Lys	Val	Thr 490	Gly	Leu	His	Ser	Gly 495	Thr
Leu	Gln	Val	Phe 500	Val	Arg	Lys	His	Gly 505	Ala	His	Gly	Ala	Ala 510	Leu	Trp
Gly	Arg	Asn 515	Gly	Gly	His	Gly	Trp 520	Arg	Gln	Thr	Gln	Ile 525	Thr	Leu	Arg

Gly Ala Asp Ile Lys Ser Val Val Phe Lys Gly Glu Lys Arg Arg Gly His Thr Gly Glu Ile Gly Leu Asp Asp Val Ser Leu Lys Lys Gly His Cys Ser Glu Glu Arg <210> SEQ ID NO 54 <211> LENGTH: 457 <212> TYPE: PRT <213 > ORGANISM: Mus musculus <400> SEQUENCE: 54 Met Gln Pro Ala Arg Lys Leu Leu Ser Leu Leu Val Leu Leu Val Met 10 Gly Thr Glu Leu Thr Gln Val Leu Pro Thr Asn Pro Glu Glu Ser Trp 25 Gln Val Tyr Ser Ser Ala Gln Asp Ser Glu Gly Arg Cys Ile Cys Thr Val Val Ala Pro Gln Gln Thr Met Cys Ser Arg Asp Ala Arg Thr Lys Gln Leu Arg Gln Leu Leu Glu Lys Val Gln Asn Met Ser Gln Ser Ile Glu Val Leu Asp Arg Arg Thr Gln Arg Asp Leu Gln Tyr Val Glu Lys Met Glu Asn Gln Met Lys Gly Leu Glu Thr Lys Phe Lys Gln Val Glu 105 Glu Ser His Lys Gln His Leu Ala Arg Gln Phe Lys Ala Ile Lys Ala Lys Met Asp Glu Leu Arg Pro Leu Ile Pro Val Leu Glu Glu Tyr Lys 135 Ala Asp Ala Lys Leu Val Leu Gln Phe Lys Glu Glu Val Gln Asn Leu Thr Ser Val Leu Asn Glu Leu Gln Glu Glu Ile Gly Ala Tyr Asp Tyr Asp Glu Leu Gln Ser Arg Val Ser Asn Leu Glu Glu Arg Leu Arg Ala 185 Cys Met Gln Lys Leu Ala Cys Gly Lys Leu Thr Gly Ile Ser Asp Pro 195 200 Val Thr Val Lys Thr Ser Gly Ser Arg Phe Gly Ser Trp Met Thr Asp 215 Pro Leu Ala Pro Glu Gly Asp Asn Arg Val Trp Tyr Met Asp Gly Tyr His Asn Asn Arg Phe Val Arg Glu Tyr Lys Ser Met Val Asp Phe Met 250 Asn Thr Asp Asn Phe Thr Ser His Arg Leu Pro His Pro Trp Ser Gly 265 Thr Gly Gln Val Val Tyr Asn Gly Ser Ile Tyr Phe Asn Lys Phe Gln $\,$ 280 Ser His Ile Ile Ile Arg Phe Asp Leu Lys Thr Glu Ala Ile Leu Lys Thr Arg Ser Leu Asp Tyr Ala Gly Tyr Asn Asn Met Tyr His Tyr Ala 310 315 Trp Gly Gly His Ser Asp Ile Asp Leu Met Val Asp Glu Asn Gly Leu 330

Trp Ala Val Tyr Ala Thr Asn Gln Asn Ala Gly Asn Ile Val Ile Ser Lys Leu Asp Pro Val Ser Leu Gln Ile Leu Gln Thr Trp Asn Thr Ser 360 Tyr Pro Lys Arg Ser Ala Gly Glu Ala Phe Ile Ile Cys Gly Thr Leu Tyr Val Thr Asn Gly Tyr Ser Gly Gly Thr Lys Val His Tyr Ala Tyr Gln Thr Asn Ala Ser Thr Tyr Glu Tyr Ile Asp Ile Pro Phe Gln Asn 410 Lys Tyr Ser His Ile Ser Met Leu Asp Tyr Asn Pro Lys Asp Arg Ala 420 Leu Tyr Ala Trp Asn Asn Gly His Gln Thr Leu Tyr Asn Val Thr Leu 440 Phe His Val Ile Arg Ser Asp Glu Leu 450 <210> SEQ ID NO 55 <211> LENGTH: 485 <212> TYPE: PRT <213> ORGANISM: Homo sapiens <400> SEQUENCE: 55 Met Ser Val Pro Leu Leu Lys Ile Gly Val Val Leu Ser Thr Met Ala Met Ile Thr Asn Trp Met Ser Gln Thr Leu Pro Ser Leu Val Gly Leu 25 Asn Thr Thr Arg Leu Ser Ala Ala Ser Gly Gly Thr Leu Asp Arg Ser Thr Gly Val Leu Pro Thr Asn Pro Glu Glu Ser Trp Gln Val Tyr Ser Ser Ala Gln Asp Ser Glu Gly Arg Cys Ile Cys Thr Val Val Ala Pro Gln Gln Thr Met Cys Ser Arg Asp Ala Arg Thr Lys Gln Leu Arg Gln Leu Leu Glu Lys Val Gln Asn Met Ser Gln Ser Ile Glu Val Leu Asp 105 Arg Arg Thr Gln Arg Asp Leu Gln Tyr Val Glu Lys Met Glu Asn Gln 115 120 Met Lys Gly Leu Glu Ser Lys Phe Lys Gln Val Glu Glu Ser His Lys 135 Gln His Leu Ala Arg Gln Phe Lys Ala Ile Lys Ala Lys Met Asp Glu Leu Arg Pro Leu Ile Pro Val Leu Glu Glu Tyr Lys Ala Asp Ala Lys 170 Leu Val Leu Gln Phe Lys Glu Glu Val Gln Asn Leu Thr Ser Val Leu 185 Asn Glu Leu Gln Glu Glu Ile Gly Ala Tyr Asp Tyr Asp Glu Leu Gln 200 Ser Arg Val Ser Asn Leu Glu Glu Arg Leu Arg Ala Cys Met Gln Lys Leu Ala Cys Gly Lys Leu Thr Gly Ile Ser Asp Pro Val Thr Val Lys 235 Thr Ser Gly Ser Arg Phe Gly Ser Trp Met Thr Asp Pro Leu Ala Pro 250

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Glu Gly Asp Asn Arg Val Trp Tyr Met Asp Gly Tyr His Asn Asn Arg

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Phe Val Arg Glu Tyr Lys Ser Met Val Asp Phe Met Asn Thr Asp Asn
Phe Thr Ser His Arg Leu Pro His Pro Trp Ser Gly Thr Gly Gln Val
Val Tyr Asn Gly Ser Ile Tyr Phe Asn Lys Phe Gln Ser His Ile Ile
Ile Arg Phe Asp Leu Lys Thr Glu Thr Ile Leu Lys Thr Arg Ser Leu
                                 330
Asp Tyr Ala Gly Tyr Asn Asn Met Tyr His Tyr Ala Trp Gly Gly His
                     345
Ser Asp Ile Asp Leu Met Val Asp Glu Ser Gly Leu Trp Ala Val Tyr
Ala Thr Asn Gln Asn Ala Gly Asn Ile Val Val Ser Arg Leu Asp Pro
Val Ser Leu Gln Thr Leu Gln Thr Trp Asn Thr Ser Tyr Pro Lys Arg
                                    395
                 390
Ser Ala Gly Glu Ala Phe Ile Ile Cys Gly Thr Leu Tyr Val Thr Asn
              405
                                 410
Gly Tyr Ser Gly Gly Thr Lys Val His Tyr Ala Tyr Gln Thr Asn Ala
                              425
Ser Thr Tyr Glu Tyr Ile Asp Ile Pro Phe Gln Asn Lys Tyr Ser His
                          440
Ile Ser Met Leu Asp Tyr Asn Pro Lys Asp Arg Ala Leu Tyr Ala Trp
                     455
Asn Asn Gly His Gln Ile Leu Tyr Asn Val Thr Leu Phe His Val Ile
                470
                                      475
Arg Ser Asp Glu Leu
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                      10
Val Pro Val Ser Arg Phe Pro Gly Lys Ile Phe Leu Tyr Cys Pro Phe
                             25
Phe Asn Arg Lys His Cys Gln Arg Phe Cys Glu Phe Phe Lys Ile Cys
Arg Lys Pro Pro Leu Ser Arg Arg Thr Thr Val Val Pro Ser Phe Pro
Leu Thr Thr Glu Ala Asp Leu Ser Leu Thr Gly Gly Pro Leu Thr Pro
Thr Gly Gly Glu Ile Gln Asp Ser Arg Val Pro His Ser Pro Glu Lys
              85
                                  90
Pro Leu Pro Pro His Ser Ala His Ala Thr Val Gly Ser Cys Phe Gln
           100
                              105
Leu Leu Pro Ala Pro Gln
       115
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What is claimed is:

1. A method to promote angiogenesis in cells or a tissue of a patient that has, or is at risk of developing, a condition selected from stroke or ischemia, comprising contacting the cells or tissue from the patient with an effective amount of a MAGP-2 protein having the amino acid sequence of SEQ ID NO:43.

260

2. The method of claim 1, wherein the MAGP-2 protein is encoded by a nucleic acid molecule comprising the nucleic acid sequence of SEQ ID NO:124.